

Laboratory for Energy and NanoScience

CHAPTER 3: Getting Hamaker coefficients with an amplitude-modulation AFM in tapping mode



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Getting Hamaker coefficients with an amplitudemodulation AFM in tapping mode

We demonstrate how to get Hamaker coefficients with an amplitude-modulation AFM operated in standard tapping mode. The Hamaker coefficient is a physical parameter that provides chemical information about the surface of samples. It is based or emerges from van der Waals forces. We use a Cypher scanning probe microscope from Asylum Research to demonstrate the method. The method is based on the research carried out at LENS during the 10s decade and early 20s.

Brief introduction of Hamaker coefficient

Rapid chemical mapping of substances with nanoscale resolution has been a target of nanotechnologists¹⁻³. The broader community relies on probing and identifying chemical substances via standard spectrometry methods that exploit electromagnetic radiation generating footprints associated to a wavelength of the electromagnetic spectrum at which resonance is observed⁴. The preference for such methods is based on robust and reproducible quantification and parameterization in measurements achieved by standard spectroscopy methodologies and the possibility to directly map a physically relevant parameter to chemical substances. To advance nanotechnology or nanosciences, higher lateral resolution is often mandatory^{5,6}. While AFM methods offer the possibility to enhance lateral resolution to sub-nm levels, quantification commonly requires very specialized equipment⁷, special environmental conditions such as ultra-high vacuum^{1,3,8} or the use of atomically flat surfaces¹. Here, we set to map a parameter related to the sample's chemical composition, *i.e.* the Hamaker coefficient H, directly from the standard observables in bimodal AFM via the non-invasive non-contact mode of operation whereby mechanical contact with the sample is avoided⁹⁻¹⁷.

Steps to collect the raw data to reconstruct force profiles

1. First, insert the cantilever and fixed the cantilever holder on the Cypher head as described in the previous article.



- 2. Click on the BimodalDualAC tab (Figure 1). This indicates that we will operate the AFM with 2 frequencies.
- **3.** Use the Igor software to put the laser spot, find the focus of the tip and the focus of the sample as described in the previous article.
- 4. Click Move to pre-engage.
- 5. The cantilever should be driven at the resonance frequency. To find the natural frequency, you need to perform the Thermal test (Figure 2). After the thermal test, copy the frequency on the thermal tab and paste it in the main tab of the master panel (Figure 3) and set the drive amplitude for the 2nd mode to 0.



Figure 2: Fit the first peak in the blue oval shape.



Figure 3: Paste the Thermal test result in the frequency field (red outline). Remember to set the amplitude for second mode (yellow outline) to 0.

- Approach the tip in the same way as described in the previous article.
- Perform a force-distance curve once while setting the force distance to 30 - 50 nm.
- 8. Perform the Thermal test again. After the thermal test, calibrate the 2nd mode first: copy the frequency (the peak position should be around 6 times of the 1st mode) on the thermal tab and paste it in the main tab of the master panel (Figure 3). Divide the amplitude in volts reading by 3.473 and fit the peak again to get spring constant and Q.
- 9. Calibrate the 1st mode: fit the 1st mode peak in the spectrum and copy the frequency on the thermal tab and paste it in the main tab of the master panel (Figure 4). Set back to the original amplitude in volts reading and fit the peak again to get spring constant and Q.

	Initialize Fit		2		Initialize Fit		?
Cal Spring Constant Cal InvOLS		2	Cal Spring	Cal Spring Constant			
	Fit Thermal Data		2		Fit Thermal Data		2
Amp InvOLS	47.82 nm/V		2	An p InvOLS	13.78 nm/V	1	2
Spring Constant	3.47 nN/nm	0	2	Spring Constant	3.47 nN/nm	0	?
Thermal DC	8.74e-15	¢	2	Thermal DC	8.74e-15	0	2
Thermal Q	129.4	÷.	2	Thermal Q	129.4		?
Frequency	77.831 kHz	Ø	2	Frequency	77.831 kHz	Q.	?
White Noise	1.19e-13	÷.	2	White Noise	1.19e-13	÷.	?
Fit Width	7.982 kHz	0	2	Fit Width	7.982 kHz	٥	?
Show Fit 🗸	Show Thermal		2	Show Fit 🗸	Show Thermal]	?
Graph Log	Log/Log	•	2	Graph Log	Log/Log	-	?

Figure 1: Amp InvOLS during first calibration in red outline. Amp invOLS during second calibration (yellow outline) is derived by dividing value in red outline by 3.473.

10. Then, find the A_c as discussed in the previous article.

- **11.** Set the free amplitude to ~0.7-0.8 times of A_c , and the setpoint to ~ 0.7-0.8 times of the free amplitude for the 1st mode. For the 2nd mode, set the free amplitude to ~0.1 times of 1st mode free amplitude.
- **12.** Set the phase lag in both channel to 90°.
- **13.** Set the scan size, scan rate and gain, and start scanning.

Steps to process the raw data

Note: R studio needs to be installed and add to the path. All the source codes can be found here



 NEW_TXT folder will be generated when the code finishes running and renames the folder if needed (Figure 6).



Figure 3: Results of the extraction can be found in the "*NEW_TXT*" folder.

Copy the 5 files (without def txt file) in the NEW_TXT folder into BIMODALHAMAKER_CloseForm\
DATA_images folder and run the Matlab file:
Bimodal_Theory_2015Dec15
(Figure 7).



Figure 4: Copy all text files extracted from the raw data except the "*Def.ibw*".

4. Revise the parameters used in each experiment in the script (Figure 8).



k 1: Spring Constant first mode
Q 1: Quality Factor first mode
f 1: Frequency first mode
k 2: Spring Constant second mode
Q 2: Quality Factor second mode
A01: Free Amplitude first mode
A02: Free Amplitude second mode

Figure 5: Insert parameters obtained during the calibration stage in the parameter panel.

5. BIMODALHAMAKER_CloseForm\DA TA_images\ProcessedData folder will be generated when the code finishes running (Figure 9).
ProcessedData
ProcessedData
ProcessedData
ProcessedData
ProcessedData
ProcessedData

P1.txt

Figure 6: Results are generated in the "*ProcessedData*" folder.

What does the Hamaker mean?

The Hamaker coefficient H, sometimes termed A outside the AFM community since in AFM, A is reserved for amplitude, measures the strength between atoms due to van der Waals forces. These forces are comparatively weak as compared to ionic and covalent interactions. van der Waals (vdW) forces can refer to the London dispersion force between atoms and sometimes also includes the Debye and Keesom interactions. A standard book that covers these forces was written by J. Israelachvili¹⁸. This book is not simple since it goes into the theory in quite detail. In Figure 10 as reproduced from Ref. 19¹⁹ the long-range forces acting between atoms on the surfaces of the tip and the surface are illustrated in a). In b) the forces are illustrated for short range interactions where mechanical contact occurs. Here the Hamaker coefficient and the respective forces act as an adhesion force which is typically constant. The tip is modelled as a sphere of radius R. A discussion of our models can be found in the papers above. In c) we show the effective lateral resolution of both van der Waals, where the distance d between the tip and the surface is larger than 0, and contact forces where d<0. We see that the highest resolution would be achieved when very close to the surface. This is because the long range vdW forces also induce lateral dispersion. In d) we show the strength of the vdW interaction as parametrized by H and the effective area of interaction with radius R.



Figure 10. a) Scheme of the non-contact interaction area Snc for a tip in the proximity of a surface for the long range attractive forces (d>a0). The tip radius is termed R. The gradient shows how the effective radius rnc grows as larger fractions p of the interaction are considered. This radius, and area, are thus termed rnc (p) and Snc (p) respectively. b) Scheme of effective radius ra (p) (attractive interactions) and rr (repulsive interactions) in the contact region where indentation occurs (d<a0). The effective value reff is a combination of rr and ra (p). c) Radius of interaction r versus distance d for ra/nc (p) (squares and rhombuses), rr (outlined circles) and reff (p) (triangles) for p=0.8 (red) and 0.9 (blue). Since ra (p) and rnc (p) are obtained from the same equations (Eqns. (1) and (2)) with the only difference being that for ra (p) d=a0, both are shown with the same markers as ra/nc (p) (squares and rhombuses). In the non-contact region, nc; reff (p)= ra/nc (p). d) Relationship between r and p for the long range interaction force. The vdW and DMT forces have been used to model the nc and c interaction areas throughout. A tip radius of 20 nm has been used to produce c) and d). All references in this caption refer to the paper "How localized are energy dissipation processes in the nanoscale?"

When water layers are present the distance d is affected by the height of the water layers on the surfaces h. An example of this is shown in Figure 11 as reproduced from Ref. 20²⁰.



Figure 11. (a) Schematic representation of a cantilever probe interacting with a surface. L is the length of the probe in the vertical axis relative to the cantilever. This distance is typically of microscale dimensions and it is thus never used as a reference. (b) Schematic of the useful distances and separations for a tip interacting with the surface in dynamic AFM. Here, zc is the equilibrium tip–surface separation for the unperturbed cantilever where $zc \ll L$. The instantaneous position of the tip relative to zc is z and d is the instantaneous tip–surface distance. (c) Schematic representation of the end of the tip with effective curvature R when both the tip and the surface are hydrated and covered by a layer of water of height h. The effective distance between the water on the tip and the surface is d* where d* = d – 2h. (d) SEM image of the end of one of the sharp tips used in this work.

Examples of Hamaker maps in bimodal AM AFM

The first figures we published in 2016 on the mapping of the Hamaker coefficient while simultaneously capturing the topography of the sample come from Ref. 16. In Figure 12 we have reproduced the most illustrative figure where we show the topography and the derivative of the tip-sample force together with the Hamaker maps. These were acquired simultaneously.

The top layer corresponds to a hydrophobic (1H,1H,2H,2H perfluorodecyl) acrylate (PFDA) sample. The bottom layer corresponds to a calcite sample. The PFDA sample was selected because it presents chemical heterogeneity in the 1-2 nm range. Heterogeneity in van der Waals forces was thus demonstrated in the 1-2 nm lateral range and in the 10s of nm lateral range with the calcite sample.



Figure 12. Topography of (a) PFDA and (d) calcite obtained in bimodal AFM and corresponding (b) and (e) Fder and (c) and (f) Hamaker H.

The precursor of the theory of Hamaker reconstruction was written a few months before in Ref. 21²¹. Anybody interested in the development of the theory can check the results and discussion of this work.

Bibliography

- 1 Sugimoto, Y. *et al.* Chemical identification of individual surface atoms by atomic force microscopy. *Nature* **446**, 64-67 (2007).
- 2 Herruzo, E. T., Perrino, A. P. & Garcia, R. Fast nanomechanical spectroscopy of soft matter. *Nat Commun* **5**, 3126, doi:10.1038/ncomms4126 (2014).
- 3 Gross, L., Mohn, F., Moll, N., Liljeroth, P. & Meyer, G. The Chemical Structure of a Molecule Resolved by Atomic Force Microscopy. *Science* **325**, 1110-1114, doi:10.1126/science.1176210 (2009).
- 4 Banwell, C. N. & McCash, E. M. *Fundamentals of molecular spectroscopy*. Vol. 851 (McGraw-Hill New York, 1994).
- 5 Gross, L. *et al.* Organic structure determination using atomic-resolution scanning probe microscopy. *Nat. Chem.* **2**, 821, doi:10.1038/nchem.765

https://www.nature.com/articles/nchem.765#supplementary-information (2010).

- 6 Hadlington, S. Nanostripe controversy in new twist, <<u>http://www.rsc.org/chemistryworld/2014/11/nanostripe-stripeynanoparticle-controversy-new-twist</u>> (2014).
- 7 Sahin, O., Magonov, S., Su, C., Quate, C. F. & Solgaard, O. An atomic force microscope tip designed to measure time-varying nanomechanical forces. *Nature nanotechnology* 2, 507, doi:10.1038/nnano.2007.226

https://www.nature.com/articles/nnano.2007.226#supplementary-information (2007).

- Setvín, M. *et al.* Chemical Identification of Single Atoms in Heterogeneous III–IV Chains on Si(100) Surface by Means of nc-AFM and DFT Calculations. *ACS nano* 6, 6969-6976, doi:10.1021/nn301996k (2012).
- 9 Lai, C.-Y. *et al.* Explaining doping in material research (Hf substitution in ZnO films) by directly quantifying the van der Waals force. *Physical Chemistry Chemical Physics* 22, 4130-4137, doi:10.1039/C9CP06441A (2020).
- 10 Lu, J.-Y., Lai, C.-Y., Almansoori, I. & Chiesa, M. The evolution in graphitic surface wettability with first-principles quantum simulations: the counterintuitive role of water. *Physical Chemistry Chemical Physics* **20**, 22636-22644, doi:10.1039/C8CP03633K (2018).
- 11 Liu, J., Lai, C.-Y., Zhang, Y.-Y., Chiesa, M. & Pantelides, S. T. Water wettability of graphene: interplay between the interfacial water structure and the electronic structure. *RSC Advances* **8**, 16918-16926, doi:10.1039/C8RA03509A (2018).
- 12 Garlisi, C., Lai, C.-Y., George, L., Chiesa, M. & Palmisano, G. Relating Photoelectrochemistry and Wettability of Sputtered Cu- and N-Doped TiO2 Thin Films via an Integrated Approach. *The Journal of Physical Chemistry C* **122**, 12369-12376, doi:10.1021/acs.jpcc.8b03650 (2018).
- 13 Chiou, Y.-C. *et al.* Direct Measurement of the Magnitude of the van der Waals Interaction of Single and Multilayer Graphene. *Langmuir : the ACS journal of surfaces and colloids* **34**, 12335-12343, doi:10.1021/acs.langmuir.8b02802 (2018).
- 14 Santos, S., Lai, C.-Y., Olukan, T. & Chiesa, M. Multifrequency AFM: from origins to convergence. *Nanoscale* **9**, 5038-5043, doi:10.1039/C7NR00993C (2017).

- 15 Lai, C.-Y., Santos, S. & Chiesa, M. Systematic Multidimensional Quantification of Nanoscale Systems From Bimodal Atomic Force Microscopy Data. ACS nano 10, 6265-6272, doi:10.1021/acsnano.6b02455 (2016).
- 16 Lai, C.-Y., Perri, S., Santos, S., Garcia, R. & Chiesa, M. Rapid quantitative chemical mapping of surfaces with sub-2nm resolution. *Nanoscale*, doi:10.1039/C6NR00496B (2016).
- 17 Garlisi, C., Scandura, G., Palmisano, G., Chiesa, M. & Lai, C.-Y. Integrated Nanoand Macroscale Investigation of Photoinduced Hydrophilicity in TiO2 Thin Films. *Langmuir : the ACS journal of surfaces and colloids* **32**, 11813-11818, doi:10.1021/acs.langmuir.6b03756 (2016).
- 18 Israelachvili, J. Intermolecular & Surface Forces. 3 edn, (Academic Press, 2011).
- 19 Santos, S. *et al.* How localized are energy dissipation processes in nanoscale interactions? *Nanotechnology* **22**, 345401 (2011).
- 20 Santos, S., Stefancich, M., Hernandez, H., Chiesa, M. & Thomson, N. H. Hydrophilicity of a Single DNA Molecule. *The Journal of Physical Chemistry C* **116**, 2807-2818, doi:10.1021/jp211326c (2012).
- 21 Chia-Yun, L., Sergio, S. & Matteo, C. Reconstruction of height of sub-nanometer steps with bimodal atomic force microscopy. *Nanotechnology* **27**, 075701 (2016).