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Full Title:

A review of open top chamber (OTC) performance across the ITEX Network

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76 Abstract (200 word max):

77 Open top chambers (OTCs) were adopted as the recommended warming mechanism by the International Tundra Experiment (ITEX) network in the early 1990's. Since then, OTCs have 78 79 been deployed across the globe. Hundreds of papers have reported the impacts of OTCs on the 80 abiotic environment and the biota. Here we review the impacts of the OTC on the physical 81 environment, with comments on the appropriateness of using OTCs to characterize the response of biota to warming. The purpose of this review is to guide readers to previously published work 82 83 and to provide recommendations for continued use of OTCs to understand the implications of 84 warming on low stature ecosystems. In short, the OTC is a useful tool to experimentally 85 manipulate temperature, however the characteristics and magnitude of warming varies greatly in 86 different environments, therefore it is important to document chamber performance to maximize 87 the interpretation of biotic response. When coupled with long-term monitoring, warming 88 experiments are a valuable means to understand the impacts of climate change on natural 89 ecosystems.

90

91 Key words: Arctic, Alpine, Tundra, Warming experiment, Large-scale coordinated experiment

92 Introduction

Warming chambers have been used for many decades to study the impacts of rising
temperature on vegetation. Interest in the impacts of warming on natural ecosystems increased
greatly in the 1980's as researchers speculated on the potential effects of climate change across
the globe. Different warming experiment designs have been employed over time spanning a

97 variety of environmental gradients (Kennedy 1995; Shaver et al. 2000; Hanson and Walker 2020; Michelsen et al. 2012). Every warming mechanism has its own strengths and weaknesses. Open 98 99 top chambers (OTCs, Fig. 1) were chosen as the recommended warming mechanism for low growing tundra vegetation by the International Tundra Experiment (ITEX) network because of 100 101 their low cost, easy deployment, and relatively few experimental artifacts (Molau and Mølgaard 102 1996; Marion et al. 1997). Currently, OTCs are widely used in alpine and Arctic locations with low-stature vegetation (Henry et al. 2022). Many of the researchers using OTCs are members of 103 the ITEX network, but many are not. While the OTCs are well suited for tundra environments, 104 105 particularly at higher latitudes where diurnal contrasts in warming are small, they have been employed in other ecosystems with low-stature vegetation such as lower latitude meadows and 106 107 peatlands. Over the past three decades there have been hundreds of papers that have documented 108 the impacts of OTCs on the physical environment and the organisms living in them. Here we 109 review what has been learned about the impacts of OTCs on the physical environment and 110 provide commentary on the interpretation of the biotic response to OTCs.

111

112 Diversity of OTCs

While somewhat standardized, the ITEX OTCs are not all the same and they vary in size from approximately 1 to 2 m² (Fig. 2). The materials used have varied over time, originally most of the OTCs deployed in North America were made of fiberglass while OTCs deployed in Europe were made of plexiglass. Although these solid self-supporting materials are most commonly used, another approach utilized thin plastic wrapped around a solid metal frame (Day et al. 2008), and another modification is the use of semiflexible material wrapped into a cone

119 (Schedlbauer et al. 2018; Parker et al. 2017, 2022). Other related approaches to experimental 120 warming in tundra ecosystems have deployed plastic tents or greenhouses (Chapin & Shaver 121 1985, Havström et al. 1993, Wookey et al. 1993), although these do not clearly fall under the definition of OTC so they are not considered directly here. While there have not been detailed in 122 123 situ studies of the difference in building materials, the common assumption is that the 124 manufacturer's specifications apply and that most commercially available building materials for 125 greenhouses are suitable. The materials are chosen to block wind and allow photosynthetically active wavelengths to pass through, although the various materials differ in their transmission of 126 127 solar radiation. OTCs may need to be periodically cleaned to remove dirt and bird guano. 128 Degradation of the materials over time is another potential issue, either through 129 photodegradation, scratches by windblown snow or dust, or by staining from tannins at sites with 130 periodic standing water. Different materials likely have different degradation rates.

Over time there have been a number of suggested improvements to the basic ITEX 131 132 chamber design. These include increasing the height (Welshofer et al. 2017), addition of water 133 filled pipes -providing thermal mass- to reduce fluctuations in the magnitude of heating 134 throughout the day and night (Godfree et al. 2010), adding heating cables to ensure heating at 135 low light levels (Sun et al. 2013), or adding small legs at each corner to allow air exchange 136 (Delarue et al. 2011). Yet the basic ITEX OTC has remained one of the most commonly 137 implemented field manipulations for examining vegetation response to warming, and it continues 138 to be used in many tundra and non-tundra settings (Bokhorst et al. 2007; Aronson and McNulty 139 2009; Spence et al. 2014; Pugnaire et al. 2020; Bjorkman et al 2020).

140

141 Physics of OTC warming

142 During the day, short-wave solar radiation is largely transmitted through the OTC walls, 143 contributing to surface warming. By contrast, the OTC walls are more opaque to outgoing long-144 wave radiation, particularly in the infrared range of the electromagnetic spectrum (> 700 nm 145 wavelength), increasing the sensible heat of air inside the OTC. The increase in temperature is due to the absorption of solar radiation directly by the plant canopy and other exposed surfaces 146 147 within the OTC (soil surface, exposed rock or standing water) and the emission of long-wave 148 radiation from these surfaces. The shape of the OTC was designed to increase the boundary layer 149 and provide the opportunity for a warm "bubble" of air to develop over the surface, by greatly 150 reducing wind speed and to reduce the loss of energy from air movement (advection). The panels 151 also provide shelter from the wind reducing heat loss by convection, yet the open-top allows air 152 to flow in and out and small eddies may form.

Because OTC performance varies both temporally and among locations, we recommend direct measurements of the physical environment in individual experiments to quantify net effects. To help understand the source of these variable impacts, it is useful to review the fundamental physics of energy balance. The equation for energy balance may be expressed as follows:

net radiation absorbed (Q*) = evapotranspiration (QLE) + sensible heat flux (QH) + ground heat flux (QG) + [net energy flux by advection (QV) + net storage (\Box S)].

Generally, QV and □S are not included as they are considered to balance out over time. The
OTC warming acts by blocking the wind and interfering with loss of energy from the surface
through QV. Furthermore, the magnitude of these flows can then vary between wet and dry

surfaces. Taken together, understanding the physics behind OTC warming can help understandthe complex impacts of OTCs on air, leaf tissue, and soil temperatures (Fig. 3).

165 The impact of OTCs on humidity varies greatly between field locations (Sjögersten and 166 Wookey 2002; Bokhorst et al. 2007). It is difficult to predict the impacts of OTCs on humidity without field observations, as humidity depends on vegetation, soil properties, and soil moisture, 167 which are linked with landscape position and lateral movements of soil water. Plants and soils in 168 169 the OTCs respond to the vapor pressure deficit (VPD). In many cases air VPD increases inside 170 OTCs as a function of increasing temperature and the subsequent increase in water holding 171 potential of warmer air (Lamentowicz et al. 2016). In some locations VPD may remain the same 172 or decrease inside the OTC presumably due to sheltering from dry winds (Dorrepaal et al. 2004). 173 At temperatures lower than 10 °C VPD is generally at levels that do not constrain photosynthesis 174 (Supplementary material 1) unless relative humidity is significantly below 50 %.

175

176 Impacts on air and leaf temperatures

177 The OTCs provide passive warming; therefore, the magnitude of warming can vary 178 greatly between locations (Fig. 4). Typically, warming is greatest around solar noon on a clear 179 day with little wind and warming may be negligible when solar intensity is low (Fig. 5). At night 180 temperatures within the OTC may also be cooler than outside the OTC due to radiative heat loss 181 and reduced mixing and exchange with surrounding air (Dabros et al. 2010). The maximum 182 potential intensity of warming is greatest near summer solstice, but in most locations, the 183 variability of warming is more directly influenced by sky conditions and weather (Fig. 6; Hollister et al. 2006; Bokhorst et al. 2013; Schedlbauer et al 2018). The effectiveness of OTCs at 184

increasing air temperatures has been shown to be reduced at higher temperatures (Carlyle et al.
2011). Therefore, the net effect of OTCs can also be highly variable across time because the
warming intensity of the OTCs depends on the ambient climate. This variability may better
reflect future climate change than methods that increase temperature a constant amount.

189 Due to the nature of the warming, the daily range of temperatures is significantly greater 190 in the OTC than the nearby ambient conditions (Fig. 6). This greater range is due to multiple 191 factors, with the two main factors being reduction of wind and that the open top allows direct 192 sunlight in part of the OTC (Hollister 1998). The greater range of temperatures and the general 193 warming changes the number of freeze thaw events and other extreme temperatures experienced 194 in the OTC (Bokhorst et al. 2013). The length of the growing season may be increased due to the 195 warmer temperatures; however, snow accumulation inside the OTCs may negate the potential for 196 earlier growth (see below **Impacts on snow**) and the lack of OTC heating at night is likely to negate any differences in freeze events in the fall despite increasing average temperatures. 197

The OTC-effect on temperature depends on where the temperature is measured. Warming 198 199 is greatest near the ground surface in the center of the plot where direct sunlight enters the OTC 200 (Hollister 1998); on average, at Northern latitudes, the Northern half of the chamber warms 201 slightly more than the Southern half, although throughout the daily cycle different regions in the 202 chamber will warm more based primarily on what regions receive the most direct sunlight. 203 Cross-site analyses benefit from standardized measurements. We therefore recommend studies 204 employing OTCs deploy temperature sensors in the most commonly used location to date: 205 halfway between the northernmost edge and center of the plot (or southernmost for Southern 206 hemisphere sites), which will usually capture the largest magnitude of warming. Similarly, deployment of temperature sensors at the standardized (10-15 cm) plant height is recommended. 207

208 At many sites the ground height is variable and the temperature sensor itself is more than a few 209 cm long; therefore, an exact location is often not possible. The OTCs' effect on plant tissue and 210 leaf surface temperatures have been found to be higher than the effect on the air temperatures 211 (DeBoeck et al 2012). The range of surface temperatures is greater within OTCs than in controls 212 and results in higher maximum temperatures (Fig. 7; Healey et al. 2016; Lindwall et al. 2016) as 213 well as lower temperatures due to shading (Jónsdóttir et al. 2005; Dabros et al. 2010). Elevated leaf temperatures have important consequences for plant water status through the increase in leaf 214 215 to air VPD.

216

217 Impacts on snow

OTCs were designed to be installed year-round; however, many studies remove them in 218 219 winter. In locations where the snowpack is lower than the height of the OTC, especially 220 windswept regions with minimal snow cover, snow is trapped inside the OTCs and may 221 accumulate: nevertheless, the warmer temperatures inside the OTCs tend to melt snow faster 222 than the surrounding (Marion et al. 1997). However, without empirical evidence it is difficult to 223 determine when snowmelt will occur within the OTC relative to the surroundings. At Alexandra 224 Fiord (Ellesmere Island, Canada) and Finse (Norway) the combined effect of accumulated snow 225 and warmer temperatures resulted in similar meltout days within the OTCs and ambient plots 226 (Bjorkman et al. 2015; Klanderud personal observation). In sites with deeper snow inside the 227 OTC, the soils under the OTC are more insulated from cold winter air and the soils are warmer 228 during the winter compared to the ambient plots (Bokhorst et al. 2013; Bjorkman et al. 2015). The impacts of snow can be large and may vary greatly throughout the year and between years 229

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(Fig. 8). Greater snow accumulation in the OTCs has also the potential to increase wateravailability and nutrients, similar to snowfence manipulations (Rixen et al. 2022).

233 Impacts on soils and belowground properties

234 The impact of OTCs on soils varies greatly between locations and may result in higher 235 soil temperatures within OTCs (Marion et al. 1997; Klanderud and Totland 2005; Bokhorst et al. 236 2013) as well as a cooling of the soil due to shading (Jónsdóttir et al. 2005; Dabros et al. 2010; 237 Hollister et al. 2006; Dabros et al. 2010; Bokhorst et al. 2013), while some sites show no effect 238 on soil temperatures (Hollister et al. 2006; Delarue et al. 2011; Buttler et al. 2015; Ma et al. 2022; Björkman unpublished data). The impact on soil temperatures is complex, while air 239 240 warming generally results in soil warming, reduced direct sunlight due to shading may offset 241 increased air temperatures and the net result may be lower heat inputs into the soil, especially in 242 landscapes with bare ground (see above Physics of OTC warming and Impacts on air and leaf 243 temperatures). Cooling of the soil surface may be due to shading by the chamber walls or 244 denser plant canopies reducing incoming radiation reaching the soil surface and thus reducing 245 the warming effect (Klanderud and Totland 2005). It is also possible that vegetation changes inside the OTCs can impact the transfer of heat from the air to the soil, similar to what has been 246 247 suggested for shrubs (Blok et al. 2010), in particular a thicker moss layer may insulate the soil 248 from ambient temperatures and incoming radiation (Lett et al. 2020). Furthermore, the lateral 249 movement of soil water from outside the OTCs can negate any potential soil warming in moist, 250 wet and flooded sites (Natali et al. 2011; Lindwall et al. 2016). The magnitude of difference may vary greatly throughout the year; for example, see differences in air temperature which may 251

252 drive soil temperature (Fig. 8). While only a few OTC experiments have measured soil warming 253 at depths of, or greater than, 20 cm (but see Hollister et al. 2006; Yang et al. 2014), it is generally 254 assumed that warming effects diminish at greater soil depths due to the small size of the OTC and the hysteresis of surrounding soils. For this reason, soil temperature should be measured near 255 256 the center of the plot. Warmer soils has resulted in increased depth of seasonal thaw under OTCs 257 in Alaska (Welker et al. 2004; Hollister et al. 2006); increased thaw depth is particularly evident 258 early in the season but may be swamped by the spatial diversity of thaw across the landscape 259 (see Hinkel and Nelson 2003).

260 The OTCs tend to decrease soil moisture in drier sites, especially at the surface 261 (Sjögersten & Wookey 2002; Bokhorst et al. 2013; van Zuijlen et al. 2022; Björnsdóttir et al 2022; Jeanbille et al. 2022), although the effect is often not statistically significant and varies 262 263 greatly depending on the soil moisture of the surroundings. However, in dry communities a 264 minor lowering in soil moisture near the surface may be enough to constrain plant performance 265 (Hudson and Henry 2010; Dorji et al. 2013; Hollister et al. 2015). In moist and wet communities, 266 the impact of the OTCs on soil moisture is often negligible (Hollister et al. 2006; Bernareggi et 267 al. 2015), yet wet communities have also experienced drying in the OTCs (Jassey et al. 2011; 268 Scharn et al. 2021). Measurements of bare ground have shown increased soil moisture in OTCs 269 due to reduced losses of soil water to the atmosphere (evaporation) as a result of reduced wind 270 speed (Bernareggi et al. 2015; D'Imperio et al. 2017). It is also possible that changes in plant 271 biomass may result in changes in evapotranspiration and soil moisture. Jeanbille et al. (2022) 272 found decreased water content of the litter inside OTCs in some sites, whereas in other sites litter 273 water content was higher in OTCs than in controls. In Latnjajare (Sweden), the OTCs are 274 deployed over five plant communities following a soil moisture gradient (Scharn et al. 2021);

here, only the warmed meadow (not heath) plots had a lower soil moisture content compared to
ambient conditions. In particular for the dry and mesic meadow plots, the timing and magnitude
of snowmelt drove the soil moisture differences between warmed and ambient plots (Scharn et
al. 2021).

Studies on soil processes and the microbial communities have often found few direct 279 impacts of the OTC (Lamb et al. 2011; Andresen et al. 2022; Jeanbille et al. 2022); however, 280 281 there have been several studies that have documented changes in the microbial communities and 282 soil processes in peatlands outside the tundra (Jassey et al. 2015; Delarue et al. 2015; Binet et al. 283 2017). The lack of a response in tundra is notable, given that warming has been shown to impact 284 the quality of litter and thereby nutrient cycling (Cornelissen et al. 2007; Jeanbille et al. 2022) and impact the soil fauna (Dollery et al. 2006; Hågvar and Klanderud 2009). The reasons for a 285 286 lack of response are unclear, but are likely due to the relatively low warming impact on soil 287 temperatures, which decreases with depth, and may be masked by the heterogeneity of soils and 288 vegetation. Furthermore, the rooting zones of the plants are likely to extend well beyond the 289 chamber walls especially for plants with long rhizomes and underground stems, and below 290 ground plant biomass has been shown to be less responsive to temperature than above ground 291 biomass (Wang et al. 2016; Ma et al. 2022a, 2022b). Nevertheless, a few studies have shown 292 earlier root growth (Sullivan and Welker 2005) and changing allocation patterns in response to 293 warming (Björk et al. 2007; Hollister and Flaherty 2010; Yang et al. 2011).

294

295 Impacts on vegetation

296 The impacts of warming on tundra vegetation are the primary focus of the ITEX network 297 and as such is described elsewhere; see Henry et al. 2022, this issue, for a review of OTC 298 impacts on community composition, plant performance and carbon cycling. Here we focus on the robustness of using observations from the experimental manipulation to guide forecasts of 299 300 vegetation change due to regional climate warming. Several studies have compared the response 301 of plants in OTCs to that of a warmer year and in many cases found similar responses (Hollister 302 and Webber 2000; Elmendorf et al. 2015; Bjorkman et al. 2020). Thawing degree days (daily 303 temperatures above the lower threshold of 0°C summed daily) have been shown to provide a 304 reasonable prediction of plant responses irrespective of warming treatment (Hollister et al. 2005a), this is for instance true for inflorescence length of *Carex aquatilis* in Northern Alaska 305 306 (Fig. 9). Comparisons of vegetation change due to warming by OTCs show similar patterns to 307 regional warming and climate warming (Hollister et al. 2015; Elmendorf et al. 2015; Bjorkman 308 et al. 2020). However, phenological development in OTCs has been shown to not advance as 309 much as would be expected based on air temperatures (Hollister et al. 2005a; Oberbauer et al. 310 2013; Parker et al. 2017, 2021). Warming experiments across all biomes have been shown to under-predict phenological advance due to regional climate warming (Wolkovich et al. 2012). 311 312 There is also evidence that OTC response may vary greatly depending on the season and year, 313 these differences can be due to moisture available (Delarue et al. 2015; Jassey and Signarieux 314 2019), the responsiveness of plants has also been shown to be less during a warm year relative to 315 a cold year (Barrett and Hollister 2016; Carbognani et al. 2016; but see Collins et al 2021).

The explanation(s) for the differences between response to experimental warming and regional climate warming is not fully understood and there are likely a suite of reasons that vary between locations and species. Examining the differences between responses may further our

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understanding of the underlying mechanisms driving response to temperature. For example, the
OTCs reduce wind, and sheltering from the wind can in itself drive vegetation change (Fitzgerald
and Kirkpatrick 2017; Momberg et al. 2021). Also, the walls of the chambers may constrain seed
rain and colonization of new species, which may protect plants inside the OTCs from
interactions with new immigrants (Yang et al. 2018).

324 The magnitude and quality of OTC warming may be significantly different from the 325 warming experienced from climate change. The magnitude and timing of OTC warming varies 326 by location and is generally on average less than 2°C, this is a modest magnitude or warming that 327 is less than some regions have already experienced due to climate change (IPCC 2022). The 328 maximum temperatures experienced in warming experiments (including OTCs) may be outside 329 the range normally experienced and the response to warming may diminish if the temperature 330 optimum is exceeded (Hollister unpublished data), it is possible that the maximum temperatures 331 may negatively impact performance (Marchand et al. 2005; Shi et al. 2010). The potential 332 decoupling of air and soil warming due to OTCs described above (Impacts on soils and 333 belowground properties) may also impact plant performance. The reduction of incoming 334 photosynthetically active radiation (PAR) and other wavelengths relevant for plant development, 335 such as far-red and ultraviolet radiation, varies within the OTC. Few studies report radiation 336 measurements along with results from OTCs even though the reduced radiation and altered 337 spectral composition, especially near the chamber walls, may impact plant production and 338 change plant morphology in ways similar to shade experiments (May et al. 2022). Reductions of 339 photosynthetic photon flux density as high as 16-25% have been documented, the OTCs reduce 340 light most when the sun is at a low angle, yet the open top allows direct sunlight and reductions 341 are near zero at solar noon especially at lower latitude (Bokhorst et al. 2007; Lindwall et al.

2016; Schollert et al. 2017). It is also reasonable to assume that the vegetation response to
warming may have built in lags and that the short-term response may be different from the longterm impacts (Hollister et al. 2005b; Rozema et al 2009).

345 Cryptogam responses can vary greatly to OTC-warming, with a dominant role for competition for light between cryptogams and vascular plants (Klanderud and Totland 2005; 346 Wahren et al. 2005; Walker et al. 2006; Cornelissen et al. 2001; Day et al. 2008). In the few 347 348 studied sites where mosses and lichens dominated, responses were highly species-specific 349 (Keuper et al 2011; Dorrepaal 2007; Bokhorst et al 2015, 2016). Moreover, this relationship can 350 even be inverted in some habitats, e.g. in *Sphagnum* dominated peatlands (Dorrepaal et al. 2006), 351 often as a result of *Sphagnum* being a stronger competitor for nitrogen (Heijmans et al. 2002). 352 Future studies may consider a specific focus on cryptogam communities with little to no vascular 353 plants to better understand the moss and lichen response to climate warming without the 354 influence of faster growing vascular plants.

355

356 Impacts on herbivores and pollinators

The impacts of OTCs on herbivores depend greatly on the species of interest. Large herbivores have often avoided OTCs, although reindeer have been seen to lean in and graze the plants within (personal observation IS Jónsdóttir at Endalen, Svalbard; EJ Cooper at Adventdalen and Ny Ålesund, Svalbard; RG Björk at Latnjajaure, Sweden). The presence of large herbivores can affect the outcome of passive warming from OTCs on plant communities. In West Greenland, herbivory by caribou and muskoxen has been observed to differentially influence the biomass response of plant functional groups to OTC-induced warming (Post and Page 17 of 44

364 Pedersen 2008). After 7 years of study, grazed plots showed higher plant community stability 365 and species diversity than ungrazed plots receiving the same warming treatment. The greater stability of grazed plots has been interpreted as the result of herbivore biomass exploitation 366 mediating the effect of interspecific competition, which increases with warmer temperatures 367 368 (Post 2013). The presence of small mammals such as lemmings and voles is patchy, although 369 anecdotal evidence suggests that they may shelter in the OTCs. At Alexandra Fiord, OTCs were 370 often covered with a screen to keep song birds from perching on the chamber walls and 371 providing unwanted nutrient inputs and decimating the seed production. Juvenile snowy owls 372 have also been observed to shelter in the OTCs on cool windy days.

373 Observations of insects are complex; for some species the chamber walls provide a 374 deterrent, while other species seek out the chambers for shelter. Once in an OTC, activity is 375 greater due to the lack of wind and warmer air temperatures (e.g. Gillespie et al 2013; Birkemoe 376 et al. 2016). Observations at Alexandra Fiord showed no impact of the OTC on insect pollination 377 nor on wind pollinated species (Robinson and Henry 2018) whereas other sites have shown 378 indications of potential pollen limitations in OTCs (Jones et al. 1997; Molau and Shaver 1997; 379 Totland and Alatalo 2002; Totland and Eide 1999). OTCs have been used to demonstrate the link 380 between timing of flowering and pollination in the High Arctic (Gillespie et al 2016; Gillespie 381 and Cooper 2022).

382

383 Items to consider

Robotic tram systems in close proximity to OTCs can provide continuous objective
 measurements of fundamental micrometeorological conditions present as well as biophysical

386 properties of vegetation represented in nearby OTCs (Healey et al. 2014). Such implementations 387 may help understand the different processes occurring at different scales across the 388 heterogeneous landscape. Similarly, handheld instrumentation has also provided analysis of 389 unique spectral characteristics linked with growth, development and phenology that are 390 undetectable to the human eye (May et al. 2020). Our understanding of physiological impacts 391 induced by OTCs has also been enhanced using thermal imaging technology (Healey et al. 392 2016). Surface tissue and underlying soil or moss temperatures are key determinants of 393 metabolic activity and monitoring such phenomena is vital for comprehensive analysis of subtle, 394 yet complex, interactions among permafrost, surface moss, cryptogamic crusts and soils, and 395 tundra vegetation. Given the many factors and potential interactions between factors, we believe 396 the use of OTCs is most effective when coupled with long-term monitoring.

397 As with any long-term experiment, it is important to clearly mark the plots with 398 permanent robust markers and the corresponding precise GPS locations. Markers may include 399 anchors that serve to retain the OTCs in position during high winds that occur at many study 400 sites. How the OTCs are secured will depend on the location and the monitoring techniques 401 deployed. Sometimes removal of the OTCs is desirable or necessary to facilitate measurement of 402 the properties within. For example, measurement of vegetation solar spectral reflectance within 403 the OTCs requires removal of the OTCs because of changes in the spectrum and amount of light 404 transmitted through the chamber walls. Measurements of ecosystem trace gas fluxes within the 405 OTCs creates a dilemma, should measurements be taken with the OTCs in place or with them 406 removed. Measurements taken with the OTCs in place reflect the vegetation performance within 407 the OTC environment that might include higher air and soil temperatures and lower light, while

in cases where the focus is the vegetation potential it is preferable to remove the OTC to measureplant performance under the same environmental conditions.

410 While most experiments using OTC leave the OTCs in place year-round, many others remove them during the winter. It may be useful to deploy the OTCs during specific times of the 411 412 year to ask specific questions. For example, Gehrman et al. (2022) deployed OTCs for late 413 summer only use. Given that autumn is the season most neglected by summer-visiting 414 researchers, autumn studies could help elucidate ecological activity and thermal sensitivities 415 during the end of the growing season and during the onset of winter dormancy. However, there are caveats here related to the potential warming performance of OTCs at lower solar angles and 416 417 shorter day-lengths as the autumnal equinox approaches.

Finally, recent attempts have been made to scale up plot-level observations from OTCs to biome-wide analyses using aerial or spaceborne observations (Westergaard-Nielsen et al. 2021). Therefore, it is important to clearly document the characteristics of the study site within the heterogeneity of the landscape and region to allow for comparison across sites and scaling of observations. The continued inclusion of remote sensing observations at a variety of scales will improve future monitoring of tundra plant responses to warming scenarios that have been projected to occur with climate change.

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426 Recommendations and Concluding Remarks

427 It is important to document the impacts of the OTC on the physical environment at each428 study site. We have shown above that the impacts of OTCs vary greatly between locations in

ways that are difficult to predict without empirical observations. Therefore, any observed
biological response must be coupled with a clear understanding of the changes to the physical
environment, including measurements at standardized locations throughout the season.

432 The OTC is a cost-effective robust method of *in situ* warming of ecosystems with low 433 stature plants such as tundra environments. The response of tundra vegetation to OTC warming 434 has been shown to be similar to that of interannual variability and latitudinal gradients 435 (Elmendorf et al. 2015). However, as with any experimental manipulation, there are artifacts that 436 may be problematic depending on the situation (Ettinger et al. 2019; Kimmel et al. 2021). The 437 OTC may or may not provide a reasonable approximation of regional climate warming 438 depending on the application. For example, the increased daily range of temperatures may be 439 unrealistic, likewise air and soil warming may be decoupled. In many cases properly 440 documenting the magnitude of warming both above-ground and below-ground may be enough to 441 properly interpret the observations that the experiment was intended to examine. In other cases, it 442 may be important to document other physical factors such as plant surface temperatures, PAR, 443 wind speed, snow accumulation, nutrient inputs, or soil moisture. It may also be important to 444 account for differences in herbivory or pollination. The small scale of the OTC makes it poorly 445 suited to examine landscape dynamics such as permafrost degradation and changing migration 446 patterns (Hegland et al. 2009; Post et al. 2009). Conversely, the small scale confers the 447 advantage that OTCs can be deployed in contrasting landscape contexts, refining the process 448 understanding necessary to underpin up-scaling such as interactions between microbes and plants 449 (Jassey et al. 2015; Jeanbille et al. 2022; Klarenberg et al. 2022). Furthermore, the OTC does not 450 require electricity and can be placed in remote locations.

In general, we recommend using the findings from OTC in conjunction with those of multiple years of observation. If the same patterns are observed in a warm year at ambient plot as observed in a warmed plot in a colder year, then the difference between warmed and control plot is mostly likely due primarily to temperature (Hollister et al. 2005a, b; Hollister et al. 2015). In cases where the response to experimental warming and regional climate change are different, then the experiment may help elucidate biological processes that better our understanding of temperature relationships.

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463 Competing Interests

The authors declare there are no competing interests.

467 Data Availability

Most of the data presented in this study are continuations or redrawing of figures from published papers. All previously unpublished data are available from the corresponding author upon reasonable request.

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3 References 4 5 Andresen, L.C., Bodé, S., Björk, R.G., Michelsen, A., Aerts, R., Boeckx, P., Cornelissen, J.H.C., 6 Klanderud, K., van Logtestijn, R.S.P., and Rütting, T. 2022. Patterns of free amino acids 7 in tundra soils reflect mycorrhizal type, shrubification, and warming. Mycorrhiza. 8 doi:10.1007/s00572-022-01075-4. 9 Aronson, E.L., and McNulty, S.G. 2009. Appropriate experimental ecosystem warming methods by ecosystem, objective, and practicality. Agricultural and Forest Meteorology 149: 0 1791-1799. doi:10.1016/j.agrformet.2009.06.007. 1 2 Barrett, R.T., and Hollister, R.D. 2016. Arctic plants are capable of sustained responses to long-3 term warming. Polar Research 35: 25405. doi:10.3402/polar.v35.25405. 4 Bernareggi, G., Carbognani, M., Petraglia, A., and Mondoni, A. 2015. Climate warming could 5 increase seed longevity of alpine snowbed plants. Alp Botany 125: 69-78. 6 doi:10.1007/s00035-015-0156-0. 7 Binet, P., Rouifed, S., Jassey, V.E.J., Toussaint, M.-L., and Chiapusio, G. 2017. Experimental climate warming alters the relationship between fungal root symbiosis and Sphagnum 8 9 litter phenolics in two peatland microhabitats. Soil Biology and Biochemistry 105: 153-161. doi:10.1016/j.soilbio.2016.11.020. 0 1 Birkemoe, T., Bergmann, S., Hasle, T.E., and Klanderud, K. 2016. Experimental warming increases herbivory by leaf-chewing insects in an alpine plant community. Ecology and 2 Evolution 6: 6955–6962. doi:10.1002/ece3.2398. 3 4 Bjorkman, A.D. 2015. Ecological and Evolutionary Consequences of Experimental and Natural 5 Warming in the High Arctic Tundra. Ph.D. Thesis, The University of British Columbia. 6 Bjorkman, A.D., Elmendorf, S.C., Beamish, A.L., Vellend, M., and Henry, G.H.R. 2015. 7 Contrasting effects of warming and increased snowfall on Arctic tundra plant phenology 8 over the past two decades. Global Change Biology 21: 4651-4661. 9 doi:10.1111/gcb.13051. 0 Bjorkman, A.D., Garcia Criado, M., Myers-Smith, I.H., Ravolainen, V., Jonsdottir, I.S., 1 Westergaard, K.B., Lawler, J.P., Aronsson, M., Bennett, B., Gardfjell, H., Heidmarsson, 2 S., Stewart, L., and Normand, S. 2020. Status and trends in Arctic vegetation: Evidence 3 from experimental warming and long-term monitoring. Ambio 49: 678-692. 4 doi:10.1007/s13280-019-01161-6. 5 Björnsdóttir, K., Barrio, I.C., and Jónsdóttir, I.S. 2022. Long-term warming manipulations reveal 6 complex decomposition responses across different tundra vegetation types. Arctic 7 Science: 1-13. doi:10.1139/as-2020-0046. 8(Blok, D., Heijmans, M.M.P.D., Schaepman-Strub, G., Kononov, A.V., Maximov, T.C., and 9 Berendse, F. 2010. Shrub expansion may reduce summer permafrost thaw in Siberian tundra. Global Change Biology 16: 1296–1305. doi:10.1111/j.1365-2486.2009.02110.x. 0 1 Bokhorst, S., Convey, P., Huiskes, A., and Aerts, R. 2016. Usnea antarctica, an important 2 Antarctic lichen, is vulnerable to aspects of regional environmental change. Polar Biol

513	39 : 511–521. doi:10.1007/s00300-015-1803-z.
514 515 516 517 518	 Bokhorst, S., Huiskes, A., Aerts, R., Convey, P., Cooper, E.J., Dalen, L., Erschbamer, B., Gudmundsson, J., Hofgaard, A., Hollister, R.D., Johnstone, J., Jónsdóttir, I.S., Lebouvier, M., Van de Vijver, B., Wahren, CH., and Dorrepaal, E. 2013. Variable temperature effects of Open Top Chambers at polar and alpine sites explained by irradiance and snow depth. Glob Change Biol 19: 64–74. doi:10.1111/gcb.12028.
519 520 521	Bokhorst, S., Huiskes, A., Convey, P., and Aerts, R. 2007. The effect of environmental change on vascular plant and cryptogam communities from the Falkland Islands and the Maritime Antarctic. BMC Ecology 7: 15. doi:10.1186/1472-6785-7-15.
522 523 524 525	 Bokhorst, S., Phoenix, G.K., Berg, M.P., Callaghan, T.V., Kirby-Lambert, C., and Bjerke, J.W. 2015. Climatic and biotic extreme events moderate long-term responses of above- and belowground sub-Arctic heathland communities to climate change. Global Change Biology 21: 4063–4075. doi:10.1111/gcb.13007.
526 527 528 529	 Buttler, A., Robroek, B.J.M., Laggoun-Défarge, F., Jassey, V.E.J., Pochelon, C., Bernard, G., Delarue, F., Gogo, S., Mariotte, P., Mitchell, E.A.D., and Bragazza, L. 2015. Experimental warming interacts with soil moisture to discriminate plant responses in an ombrotrophic peatland. J Veg Sci 26: 964–974. doi:10.1111/jvs.12296.
530 531 532	 Carbognani, M., Bernareggi, G., Perucco, F., Tomaselli, M., and Petraglia, A. 2016. Micro- climatic controls and warming effects on flowering time in alpine snowbeds. Oecologia 182: 573–585. doi:10.1007/s00442-016-3669-3.
533 534 535 536	Carlyle, C.N., Fraser, L.H., and Turkington, R. 2011. Tracking Soil Temperature and Moisture in a Multi-Factor Climate Experiment in Temperate Grassland: Do Climate Manipulation Methods Produce their Intended Effects? Ecosystems 14: 489–502. doi:10.1007/s10021-011-9425-y.
537 538 539	Chapin III, F.S., and Shaver, G.R. 1985. Individualistic Growth Response of Tundra Plant Species to Environmental Manipulations in the Field. Ecology 66 : 564–576. doi:10.2307/1940405.
540 541 542	 Chapin III, F.S., Shaver, G.R., Giblin, A.E., Nadelhoffer, K.J., and Laundre, J.A. 1995. Responses of Arctic Tundra to Experimental and Observed Changes in Climate. Ecology 76: 694–711. doi:10.2307/1939337.
543 544 545 546 547 548 549 550 551	 Collins, C.G., Elmendorf, S.C., Hollister, R.D., Henry, G.H.R., Clark, K., Bjorkman, A.D., Myers-Smith, I.H., Prevéy, J.S., Ashton, I.W., Assmann, J.J., Alatalo, J.M., Carbognani, M., Chisholm, C., Cooper, E.J., Forrester, C., Jónsdóttir, I.S., Klanderud, K., Kopp, C.W., Livensperger, C., Mauritz, M., May, J.L., Molau, U., Oberbauer, S.F., Ogburn, E., Panchen, Z.A., Petraglia, A., Post, E., Rixen, C., Rodenhizer, H., Schuur, E.A.G., Semenchuk, P., Smith, J.G., Steltzer, H., Totland, Ø., Walker, M.D., Welker, J.M., and Suding, K.N. 2021. Experimental warming differentially affects vegetative and reproductive phenology of tundra plants. Nat Commun 12: 3442. doi:10.1038/s41467- 021-23841-2.
552 553 554	Cornelissen, J.H.C., Callaghan, T.V., Alatalo, J.M., Michelsen, A., Graglia, E., Hartley, A.E., Hik, D.S., Hobbie, S.E., Press, M.C., Robinson, C.H., Henry, G.H.R., Shaver, G.R., Phoenix, G.K., Gwynn Jones, D., Jonasson, S., Chapin III, F.S., Molau, U., Neill, C.,

555 556 557	Lee, J.A., Melillo, J.M., Sveinbjörnsson, B., and Aerts, R. 2001. Global change and arctic ecosystems: is lichen decline a function of increases in vascular plant biomass? Journal of Ecology 89 : 984–994. doi:10.1111/j.1365-2745.2001.00625.x.
558 559 560 561 562 563 564 565 566	 Cornelissen, J.H.C., Van Bodegom, P.M., Aerts, R., Callaghan, T.V., Van Logtestijn, R.S.P., Alatalo, J., Stuart Chapin, F., Gerdol, R., Gudmundsson, J., Gwynn-Jones, D., Hartley, A.E., Hik, D.S., Hofgaard, A., Jónsdóttir, I.S., Karlsson, S., Klein, J.A., Laundre, J., Magnusson, B., Michelsen, A., Molau, U., Onipchenko, V.G., Quested, H.M., Sandvik, S.M., Schmidt, I.K., Shaver, G.R., Solheim, B., Soudzilovskaia, N.A., Stenström, A., Tolvanen, A., Totland, Ø., Wada, N., Welker, J.M., Zhao, X., and Team, M.O.L. 2007. Global negative vegetation feedback to climate warming responses of leaf litter decomposition rates in cold biomes. Ecology Letters 10: 619–627. doi:10.1111/j.1461-0248.2007.01051.x.
567 568 569	Dabros, A., Fyles, J.W., and Strachan, I.B. 2010. Effects of open-top chambers on physical properties of air and soil at post-disturbance sites in northwestern Quebec. Plant Soil 333 : 203–218. doi:10.1007/s11104-010-0336-z.
570 571 572	Day, T.A., Ruhland, C.T., and Xiong, F.S. 2008. Warming increases aboveground plant biomass and C stocks in vascular-plant-dominated Antarctic tundra. Global Change Biology 14 : 1827–1843. doi:10.1111/j.1365-2486.2008.01623.x.
573 574	De Boeck, H.J., De Groote, T., and Nijs, I. 2012. Leaf temperatures in glasshouses and open-top chambers. New Phytol 194 : 1155–1164. doi:10.1111/j.1469-8137.2012.04117.x.
575 576 577 578	Delarue, F., Buttler, A., Bragazza, L., Grasset, L., Jassey, V.E.J., Gogo, S., and Laggoun- Défarge, F. 2015. Experimental warming differentially affects microbial structure and activity in two contrasted moisture sites in a Sphagnum-dominated peatland. Science of The Total Environment 511: 576–583. doi:10.1016/j.scitotenv.2014.12.095.
579 580 581 582	 Delarue, F., Laggoun-Défarge, F., Buttler, A., Gogo, S., Jassey, V.E.J., and Disnar, JR. 2011. Effects of short-term ecosystem experimental warming on water-extractable organic matter in an ombrotrophic Sphagnum peatland (Le Forbonnet, France). Organic Geochemistry 42: 1016–1024. doi:10.1016/j.orggeochem.2011.07.005.
583 584 585 586	D'Imperio, L., Nielsen, C.S., Westergaard-Nielsen, A., Michelsen, A., and Elberling, B. 2017. Methane oxidation in contrasting soil types: responses to experimental warming with implication for landscape-integrated CH ₄ budget. Glob Change Biol 23 : 966–976. doi:10.1111/gcb.13400.
587 588 589 590	Dollery, R., Hodkinson, I.D., and Jónsdóttir, I.S. 2006. Impact of warming and timing of snow melt on soil microarthropod assemblages associated with Dryas-dominated plant communities on Svalbard. Ecography 29: 111–119. doi:10.1111/j.2006.0906- 7590.04366.x.
591 592 593 594	Dorji, T., Totland, Ø., Moe, S.R., Hopping, K.A., Pan, J.B., and Klein, J.A. 2013. Plant functional traits mediate reproductive phenology and success in response to experimental warming and snow addition in Tibet. Global Change Biol 19 : 459–472. doi:10.1111/gcb.12059.
595 596	Dorrepaal, E. 2007. Are plant growth-form-based classifications useful in predicting northern ecosystem carbon cycling feedbacks to climate change? Journal of Ecology 95 : 1167–

597	1180. doi:10.1111/j.1365-2745.2007.01294.x.
598 599 600 601	Dorrepaal, E., Aerts, R., Cornelissen, J.H.C., Callaghan, T.V., and Van Logtestijn, R.S.P. 2004. Summer warming and increased winter snow cover affect Sphagnum fuscum growth, structure and production in a sub-arctic bog. Global Change Biology 10 : 93–104. doi:10.1111/j.1365-2486.2003.00718.x.
602 603 604	Dorrepaal, E., Aerts, R., Cornelissen, J.H.C., Van Logtestijn, R.S.P., and Callaghan, T.V. 2006. Sphagnum modifies climate-change impacts on subarctic vascular bog plants. Functional Ecology 20 : 31–41. doi:10.1111/j.1365-2435.2006.01076.x.
605 606 607 608 609 610	 Elmendorf, S.C., Henry, G.H.R., Hollister, R.D., Fosaa, A.M., Gould, W.A., Hermanutz, L., Hofgaard, A., Jónsdóttir, I.S., Jorgenson, J.C., Lévesque, E., Magnusson, B., Molau, U., Myers-Smith, I.H., Oberbauer, S.F., Rixen, C., Tweedie, C.E., and Walker, M.D. 2015. Experiment, monitoring, and gradient methods used to infer climate change effects on plant communities yield consistent patterns. Proc Natl Acad Sci USA 112: 448–452. doi:10.1073/pnas.1410088112.
611 612 613 614	Ettinger, A.K., Chuine, I., Cook, B.I., Dukes, J.S., Ellison, A.M., Johnston, M.R., Panetta, A.M., Rollinson, C.R., Vitasse, Y., and Wolkovich, E.M. 2019. How do climate change experiments alter plot-scale climate? Ecology Letters 22 : 748–763. doi:10.1111/ele.13223.
615 616 617 618	Fitzgerald, N.B., and Kirkpatrick, J.B. 2017. Wind Distortion in Alpine and Subantarctic Plants is Constant among Life Forms but does not Necessarily Reflect Prevailing Wind Direction. Arctic, Antarctic, and Alpine Research 49 : 521–535. doi:10.1657/AAAR0016- 054.
619 620 621	Gehrmann, F., Ziegler, C., and Cooper, E.J. 2022. Onset of autumn senescence in High Arctic plants shows similar patterns in natural and experimental snow depth gradients. Arctic Science 00 : 23. doi:10.1139/as-2020-0044.
622 623 624	Gillespie, M.A.K., Baggesen, N., and Cooper, E.J. 2016. High Arctic flowering phenology and plant–pollinator interactions in response to delayed snow melt and simulated warming. Environ. Res. Lett. 11 : 115006. doi:10.1088/1748-9326/11/11/115006.
625 626 627	Gillespie, M.A.K., Jónsdóttir, I.S., Hodkinson, I.D., and Cooper, E.J. 2013. Aphid–willow interactions in a high Arctic ecosystem: responses to raised temperature and goose disturbance. Global Change Biology 19 : 3698–3708. doi:10.1111/gcb.12284.
628 629 630 631	Godfree, R., Robertson, B., Bolger, T., Carnegie, M., and Young, A. 2011. An improved hexagon open-top chamber system for stable diurnal and nocturnal warming and atmospheric carbon dioxide enrichment. Global Change Biology 17 : 439–451. Wiley Online Library. doi:10.1111/j.1365-2486.2010.02276.x.
632 633 634	Hågvar, S., and Klanderud, K. 2009. Effect of simulated environmental change on alpine soil arthropods. Global Change Biology 15: 2972–2980. doi:10.1111/j.1365- 2486.2009.01926.x.
635 636 637	Hanson, P.J., and Walker, A.P. 2020. Advancing global change biology through experimental manipulations: Where have we been and where might we go? Glob Change Biol 26 : 287–299. doi:10.1111/gcb.14894.

638 639 640	Havström, M., Callaghan, T.V., Jonasson, S., and Havstrom, M. 1993. Differential growth responses of <i>Cassiope tetragona</i> , an arctic dwarf-shrub, to environmental perturbations among three contrasting high- and sub- arctic sites. Oikos 66 : 389. doi:10.2307/3544933.
641 642 643 644	 Healey, N., Oberbauer, S.F., Ahrends, H., Dierick, D., Welker, J., Leffler, A., Hollister, R.D., Vargas, S., and Tweedie, C.E. 2014. A mobile instrumented sensor platform for long-term terrestrial ecosystem analysis: An example application in an arctic tundra ecosystem. Journal of Environmental Informatics 24: 1–10. doi:10.3808/jei.201400278.
645	Healey, N.C., Oberbauer, S.F., and Hollister, R.D. 2016. Examination of surface temperature
646	modification by Open Top Chambers along moisture and latitudinal gradients in Arctic
647	Alaska using thermal infrared photography. Remote Sensing 8: 54 1–19.
648	doi:10.3390/rs8010054.
649	Hegland, S.J., Nielsen, A., Lázaro, A., Bjerknes, AL., and Totland, Ø. 2009. How does climate
650	warming affect plant-pollinator interactions? Ecology Letters 12: 184–195.
651	doi:10.1111/j.1461-0248.2008.01269.x.
652	Heijmans, M.M.P.D., Klees, H., and Berendse, F. 2002. Competition between Sphagnum
653	magellanicum and Eriophorum angustifolium as affected by raised CO2 and increased N
654	deposition. Oikos 97: 415–425. doi:10.1034/j.1600-0706.2002.970311.x.
655 656 657 658	 Henry, G.H.R., Klanderud, K., Hollister, R.D., Bjorkman, A.D., Björk, R.G., Elphinstone, C., Jonsdottir, I.S., Molau, U., Oberbauer, S.F., Petraglia, A., Rixen, C., and Wookey, P.A. 2022. The impacts of climate change on tundra ecosystems: Three Decades of Results from the International Tundra Experiment (ITEX) Network. Arctic Science Submitted.
659	Hinkel, K.M., and Nelson, F.E. 2003. Spatial and temporal patterns of active layer thickness at
660	Circumpolar Active Layer Monitoring (CALM) sites in northern Alaska, 1995–2000.
661	Journal of Geophysical Research: Atmospheres 108. doi:10.1029/2001JD000927.
662	Hollister, R.D. 1998. Response of Wet Meadow Tundra to Interannual and Manipulated
663	Temperature Variation: Implications for Climate Change Research. Master's Thesis,
664	Michigan State University.
665 666 667	Hollister, R.D., and Flaherty, K.J. 2010. Above and belowground plant biomass response to experimental warming in northern Alaska. Applied Vegetation Science 13 : 378–387. doi:10.1111/j.1654-109X.2010.01079.x.
668	Hollister, R.D., May, J.L., Kremers, K.S., Tweedie, C.E., Oberbauer, S.F., Liebig, J.A., Botting,
669	T.F., Barrett, R.T., and Gregory, J.L. 2015. Warming experiments elucidate the drivers of
670	observed directional changes in tundra vegetation. Ecology and Evolution 5: 1881–1895.
671	doi:10.1002/ece3.1499.
672 673	Hollister, R.D., and Webber, P.J. 2000. Biotic validation of small open-top chambers in a tundra ecosystem. Global Change Biol 6 : 835–842. doi:10.1046/j.1365-2486.2000.00363.x.
674	Hollister, R.D., Webber, P.J., and Bay, C. 2005a. Plant response to temperature in northern
675	Alaska: Implications for predicting vegetation change. Ecology 86: 1562–1570.
676	doi:10.1890/04-0520.
677 678	Hollister, R.D., Webber, P.J., Nelson, F.E., and Tweedie, C.E. 2006. Soil thaw and temperature response to air warming varies by plant community: Results from an open-top chamber

679 680	experiment in northern Alaska. Arct Antarct Alp Res 38 : 206–215. doi:10.1657/1523-0430(2006)38[206:STATRT]2.0.CO;2.
681 682 683	Hollister, R.D., Webber, P.J., and Tweedie, C.E. 2005b. The response of Alaskan arctic tundra to experimental warming: Differences between short- and long-term responses. Global Change Biol 11 : 525–536. doi:10.1111/j.1365-2486.2005.00926.x.
684 685	Hudson, J.M.G., and Henry, G.H.R. 2010. High Arctic plant community resists 15 years of experimental warming. J Ecol 98 : 1035–1041. doi:10.1111/j.1365-2745.2010.01690.x.
686	IPCC 2022. Climate Change 2022: Impacts, Adaptation, and Vulnerability. Contribution of
687	Working Group II to the Sixth Assessment Report of the Intergovernmental Panel on
688	Climate Change. Cambridge University Press.
689	Jassey, V.E.J., and Signarbieux, C. 2019. Effects of climate warming on Sphagnum
690	photosynthesis in peatlands depend on peat moisture and species-specific anatomical
691	traits. Global Change Biology 25: 3859–3870. doi:10.1111/gcb.14788.
692	Jassey, V.E.J., Signarbieux, C., Hättenschwiler, S., Bragazza, L., Buttler, A., Delarue, F.,
693	Fournier, B., Gilbert, D., Laggoun-Défarge, F., Lara, E., T. E. Mills, R., Mitchell, E.A.D.,
694	Payne, R.J., and Robroek, B.J.M. 2015. An unexpected role for mixotrophs in the
695	response of peatland carbon cycling to climate warming. Sci Rep 5: 16931.
696	doi:10.1038/srep16931.
697	Jeanbille, M., Clemmensen, K., Juhanson, J., Michelsen, A., Cooper, E.J., Henry, G.H.R.,
698	Hofgaard, A., Hollister, R.D., Jónsdóttir, I.S., Klanderud, K., Tolvanen, A., and Hallin, S.
699	2022. Site-specific responses of fungal and bacterial abundances to experimental
700	warming in litter and soil across Arctic and alpine tundra. Arctic Science: 1–14.
701	doi:10.1139/as-2020-0053.
702 703 704 705	Jones, M.H., Bay, C., and Nordenhall, U. 1997. Effects of experimental warming on Arctic willows (<i>Salix</i> spp.): A comparison of responses from the Canadian High Arctic, Alaskan Arctic, and Swedish Subarctic. Global Change Biol 3 : 55–60. doi:10.1111/j.1365-2486.1997.gcb135.x.
706	Jónsdóttir, I.S., Magnússon, B., Gudmundsson, J., Elmarsdóttir, Á., and Hjartarson, H. 2005.
707	Variable sensitivity of plant communities in Iceland to experimental warming. Global
708	Change Biology 11 : 553–563. doi:10.1111/j.1365-2486.2005.00928.x.
709	Junzeng, X., Qi, W., Shizhang, P., and Yanmei, Y. 2012. Error of Saturation Vapor Pressure
710	Calculated by Different Formulas and Its Effect on Calculation of Reference
711	Evapotranspiration in High Latitude Cold Region. Procedia Engineering 28: 43–48.
712	doi:10.1016/j.proeng.2012.01.680.
713	Klarenberg, I.J., Keuschnig, C., Russi Colmenares A.J., Warshan D., Jungblut A.D., Jónsdóttir
714	I.S., Vilhelmsson, O. 2022. Long-term warming effects on the microbiome and nifH gene
715	abundance of a common moss species in sub-Arctic tundra. New Phytologist 234: 2044-
716	2056. https://doi.org/10.1111/nph.17837.
717	Kennedy, A.D. 1995. Temperature Effects of Passive Greenhouse Apparatus in High-Latitude
718	Climate Change Experiments. Functional Ecology 9: 340–350. [British Ecological
719	Society, Wiley]. doi:10.2307/2390583.

720 721 722 723	 Keuper, F., Dorrepaal, E., Van Bodegom, P.M., Aerts, R., Van Logtestijn, R.S.P., Callaghan, T.V., and Cornelissen, J.H.C. 2011. A Race for Space? How Sphagnum fuscum stabilizes vegetation composition during long-term climate manipulations. Global Change Biology 17: 2162–2171. doi:10.1111/j.1365-2486.2010.02377.x.
724	Kimmel, K., Dee, L.E., Avolio, M.L., and Ferraro, P.J. 2021. Causal assumptions and causal
725	inference in ecological experiments. Trends in Ecology & Evolution 36: 1141–1152.
726	doi:10.1016/j.tree.2021.08.008.
727 728 729	Klanderud, K., and Totland, Ø. 2005. Simulated Climate Change Altered Dominance Hierarchies and Diversity of an Alpine Biodiversity Hotspot. Ecology 86 : 2047–2054. doi:10.1890/04-1563.
730	Lamb, E.G., Han, S., Lanoil, B.D., Henry, G.H.R., Brummell, M.E., Banerjee, S., and Siciliano,
731	S.D. 2011. A High Arctic soil ecosystem resists long-term environmental manipulations.
732	Global Change Biology 17: 3187–3194. doi:10.1111/j.1365-2486.2011.02431.x.
733 734 735 736	Lamentowicz, M., Słowińska, S., and Słowiński, M. 2016. Combining short-term manipulative experiments with long-term palaeoecological investigations at high resolution to assess the response of Sphagnum peatlands to drought, fire and warming. Mires and Peat: 1–17. doi:10.19189/MaP.2016.OMB.244.
737	Lett, S., Teuber, L.M., Krab, E.J., Michelsen, A., Olofsson, J., Nilsson, MC., Wardle, D.A., and
738	Dorrepaal, E. 2020. Mosses modify effects of warmer and wetter conditions on tree
739	seedlings at the alpine treeline. Global Change Biology 26: 5754–5766.
740	doi:10.1111/gcb.15256.
741	Lindwall, F., Svendsen, S.S., Nielsen, C.S., Michelsen, A., and Rinnan, R. 2016. Warming
742	increases isoprene emissions from an arctic fen. Science of The Total Environment 553:
743	297–304. doi:10.1016/j.scitotenv.2016.02.111.
744	Ma, T., Parker, T., Fetcher, N., Unger, S.L., Gewirtzman, J., Moody, M.L., and Tang, J. 2022a.
745	Leaf and root phenology and biomass of Eriophorum vaginatum in response to warming
746	in the Arctic. Journal of Plant Ecology: rtac010. doi:10.1093/jpe/rtac010.
747	Ma, T., Parker, T., Unger, S., Gewirtzman, J., Fetcher, N., Moody, M.L., and Tang, J. 2022b.
748	Responses of root phenology in ecotypes of Eriophorum vaginatum to transplantation and
749	warming in the Arctic. Science of The Total Environment 805: 149926.
750	doi:10.1016/j.scitotenv.2021.149926.
751	Marchand, F.L., Mertens, S., Kockelbergh, F., Beyens, L., and Nijs, I. 2005. Performance of
752	High Arctic tundra plants improved during but deteriorated after exposure to a simulated
753	extreme temperature event. Global Change Biology 11 : 2078–2089. doi:10.1111/j.1365-
754	2486.2005.01046.x.
755 756 757 758	 Marion, G. m., Henry, G. h. r., Freckman, D. w., Johnstone, J., Jones, G., Jones, M. h., Lévesque, E., Molau, U., Mølgaard, P., Parsons, A. n., Svoboda, J., and Virginia, R. a. 1997. Opentop designs for manipulating field temperature in high-latitude ecosystems. Global Change Biology 3: 20–32. doi:10.1111/j.1365-2486.1997.gcb136.x.
759 760 761	May, J.L., Hollister, R.D., Betway, K.R., Harris, J.A., Tweedie, C.E., Welker, J.M., Gould, W.A., and Oberbauer, S.F. 2020. NDVI changes show warming increases the length of the green season at tundra communities in northern alaska: A fine-scale analysis. Front.

762	Plant Sci. 11: 1174. doi:10.3389/fpls.2020.01174.
763 764 765	May, J.L., Oberbauer, S.F., Unger, S.L., Simon, M.J., Betway, K.R., and Hollister, R.D. 2022. Shading decreases and delays NDVI and flowering of prostrate Arctic shrubs. Arctic Science: 1–12. doi:10.1139/as-2020-0043.
766 767 768	Michelsen, A., Rinnan, R., and Jonasson, S. 2012. Two decades of experimental manipulations of heaths and forest understory in the subarctic. Ambio 41 : 218–230. doi:10.1007/s13280-012-0303-4.
769 770	Molau, U., and Mølgaard, P. 1996. International Tundra Experiment (ITEX) manual. Second edition. Danish Polar Center, Copenhagen, Denmark.
771 772 773	Molau, U., and Shaver, G.R. 1997. Controls on seed production and seed germinability in <i>Eriophorum vaginatum</i> . Global Change Biology 3 : 80–88. doi:10.1111/j.1365-2486.1997.gcb130.x.
774 775 776 777	Momberg, M., Hedding, D.W., Luoto, M., and le Roux, P.C. 2021. Species differ in their responses to wind: the underexplored link between species fine-scale occurrences and variation in wind stress. Journal of Vegetation Science 32 : e13093. doi:10.1111/jvs.13093.
778 779 780 781	Natali, S.M., Schuur, E. a. G., Trucco, C., Hicks Pries, C.E., Crummer, K.G., and Baron Lopez, A.F. 2011. Effects of experimental warming of air, soil and permafrost on carbon balance in Alaskan tundra. Global Change Biology 17 : 1394–1407. doi:10.1111/j.1365-2486.2010.02303.x.
782 783 784 785 786 787	 Oberbauer, S.F., Elmendorf, S.C., Troxler, T.G., Hollister, R.D., Rocha, A.V., Bret-Harte, M.S., Dawes, M.A., Fosaa, A.M., Henry, G.H.R., Høye, T.T., Jarrad, F.C., Jónsdóttir, I.S., Klanderud, K., Klein, J.A., Molau, U., Rixen, C., Schmidt, N.M., Shaver, G.R., Slider, R.T., Totland, Ø., Wahren, CH., and Welker, J.M. 2013. Phenological response of tundra plants to background climate variation tested using the International Tundra Experiment. Phil. Trans. R. Soc. B 368: 20120481. doi:10.1098/rstb.2012.0481.
788 789 790	Parker, T.C., Tang, J., Clark, M.B., Moody, M.M., and Fetcher, N. 2017. Ecotypic differences in the phenology of the tundra species <i>Eriophorum vaginatum</i> reflect sites of origin. Ecol Evol 7 : 9775–9786. doi:10.1002/ece3.3445.
791 792 793	Parker, T.C., Unger, S.L., Moody, M.L., Tang, J., and Fetcher, N. 2022. Intraspecific variation in phenology offers resilience to climate change for <i>Eriophorum vaginatum</i> . Arctic Science: 1–17. doi:10.1139/as-2020-0039.
794 795	Post, E. 2013. Erosion of community diversity and stability by herbivore removal under warming. Proc. R. Soc. B. 280 : 20122722. doi:10.1098/rspb.2012.2722.
796 797 798 799 800 801	 Post, E., Forchhammer, M.C., Bret-Harte, M.S., Callaghan, T.V., Christensen, T.R., Elberling, B., Fox, A.D., Gilg, O., Hik, D.S., Høye, T.T., Ims, R.A., Jeppesen, E., Klein, D.R., Madsen, J., McGuire, A.D., Rysgaard, S., Schindler, D.E., Stirling, I., Tamstorf, M.P., Tyler, N.J.C., van der Wal, R., Welker, J., Wookey, P.A., Schmidt, N.M., and Aastrup, P. 2009. Ecological dynamics across the Arctic associated with recent Climate Change. Science 325: 1355–1358. doi:10.1111/j.1365-2486.1997.gcb135.x.
802	Post, E., and Pedersen, C. 2008. Opposing plant community responses to warming with and

803 804	without herbivores. Proc. Natl. Acad. Sci. U.S.A. 105 : 12353–12358. doi:10.1073/pnas.0802421105.
805	Pugnaire, F.I., Pistón, N., Macek, P., Schöb, C., Estruch, C., and Armas, C. 2020. Warming
806	enhances growth but does not affect plant interactions in an alpine cushion species.
807	Perspectives in Plant Ecology, Evolution and Systematics 44: 125530.
808	doi:10.1016/j.ppees.2020.125530.
809	 Rixen, C., Høye, T.T., Macek, P., Aerts, R., Alatalo, J., Andeson, J., Arnold, P., Barrio, I.C.,
810	Bjerke, J., Björkman, M.P., Blok, D., Blume-Werry, G., Boike, J., Bokhorst, S.,
811	Carbognani, M., Christiansen, C., Convey, P., Cooper, E.J., Cornelissen, J.H.C., Coulson,
812	S., Dorrepaal, E., Elberling, B., Elmendorf, S., Elphinstone, C., Forte, T.G.W., Frei, E.R.,
813	Geange, S., Gehrmann, F., Gibson, C., Grogan, P., Halbritter Rechsteiner, A., Harte, J.,
814	Henry, G.H.R., Inouye, D., Irwin, R., Jespersen, G., Jónsdóttir, I.S., Jung, J.Y., Klinges,
815	D., Kudo, G., Lämsä, J., Lee, H., Lembrechts, J., Lett, S., Lynn, J.S., Mann, H.M.,
816	Mastepanov, M., Morse, J., Myers-Smith, I., Olofsson, J., Paavola, R., Petraglia, A.,
817	Phoenix, G., Semenchuk, P., Siewert, M., Slatyer, R., Spasojevic, M., Suding, K.,
818	Sullivan, P., Thompson, K., Väisänen, M., Vandvik, V., Venn, S., Walz, J., Way, R.,
819	Welker, J., Wipf, S., and Zong, S. 2022. Winters are changing: snow effects on Arctic
820	and alpine tundra ecosystems. Arctic Science: AS-2020-0058. doi:10.1139/AS-2020-
821	0058.
822 823	Robinson, S.V.J., and Henry, G.H.R. 2018. High Arctic plants show independent responses to pollination and experimental warming. Botany 96 : 385–396. doi:10.1139/cjb-2017-0200.
824	 Rozema, J., Weijers, S., Broekman, R., Blokker, P., Buizer, B., Werleman, C., El Yaqine, H.,
825	Hoogedoorn, H., Fuertes, M.M., and Cooper, E. 2009. Annual growth of Cassiope
826	tetragona as a proxy for Arctic climate: developing correlative and experimental transfer
827	functions to reconstruct past summer temperature on a millennial time scale. Global
828	Change Biology 15: 1703–1715. doi:10.1111/j.1365-2486.2009.01858.x.
829	Scharn, R., Little, C.J., Bacon, C.D., Alatalo, J.M., Antonelli, A., Björkman, M.P., Molau, U.,
830	Nilsson, R.H., and Björk, R.G. 2021. Decreased soil moisture due to warming drives
831	phylogenetic diversity and community transitions in the tundra. Environ. Res. Lett. 16:
832	064031. doi:10.1088/1748-9326/abfe8a.
833	Schedlbauer, J.L., Fetcher, N., Hood, K., Moody, M.L., and Tang, J. 2018. Effect of growth
834	temperature on photosynthetic capacity and respiration in three ecotypes of <i>Eriophorum</i>
835	<i>vaginatum</i> . Ecol Evol 8: 3711–3725. doi:10.1002/ece3.3939.
836	Schollert, M., Kivimäenpää, M., Michelsen, A., Blok, D., and Rinnan, R. 2017. Leaf anatomy,
837	BVOC emission and CO ₂ exchange of arctic plants following snow addition and summer
838	warming. Ann Bot 119 : 433–445. doi:10.1093/aob/mcw237.
839	Shaver, G.R., Canadell, J.G., Chapin, F.S., Gurevitch, J., Harte, J., Henry, G., Ineson, P.,
840	Jonasson, S., Melillo, J.M., Pitelka, L.F., and Rustad, L.E. 2000. Global warming and
841	terrestrial ecosystems: A conceptual framework for analysis. BioScience 50: 871–882.
842	doi:10.1641/0006-3568(2000)050[0871:GWATEA]2.0.CO;2.
843	Shi, F.S., Wu, Y., Wu, N., and Luo, P. 2010. Different growth and physiological responses to
844	experimental warming of two dominant plant species Elymus nutans and Potentilla
845	anserina in an alpine meadow of the eastern Tibetan Plateau. Photosynt. 48: 437–445.

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876

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880

881

doi:10.1007/s11099-010-0058-8.

- Sjögersten, S., and Wookey, P.A. 2002. Spatio-temporal variability and environmental controls of methane fluxes at the forest-tundra ecotone in the Fennoscandian mountains. Global 848 Change Biology 8: 885–894. doi:10.1046/j.1365-2486.2002.00522.x.
 - Spence, L.A., Liancourt, P., Boldgiv, B., Petraitis, P.S., and Casper, B.B. 2014. Climate change and grazing interact to alter flowering patterns in the Mongolian steppe. Oecologia 175: 251-260. doi:10.1007/s00442-014-2884-z.
- Sullivan, P.F., and Welker, J.M. 2005. Warming chambers stimulate early season growth of an arctic sedge: results of a minirhizotron field study. Oecologia 142: 616-626. 855 doi:10.1007/s00442-004-1764-3.
- 856 Sun, S.-Q., Peng, L., Wang, G.-X., Wu, Y.-H., Zhou, J., Bing, H.-J., Yu, D., and Luo, J. 2013. 857 An improved open-top chamber warming system for global change research. Silva Fenn. 858 47. doi:10.14214/sf.960.
 - Totland, Ø., and Alatalo, J.M. 2002. Effects of temperature and date of snowmelt on growth, reproduction, and flowering phenology in the arctic/alpine herb, Ranunculus glacialis. Oecologia 133: 168-175. doi:10.1007/s00442-002-1028-z.
 - Totland, Ø., and Eide, W. 1999. Environmentally-dependent pollen limitation on seed production in alpine Ranunculus acris. Ecoscience 6: 173–179. doi:10.1080/11956860.1999.11682518.
 - Wahren, C.-H.A., Walker, M.D., and Bret-Harte, M.S. 2005. Vegetation responses in Alaskan arctic tundra after 8 years of a summer warming and winter snow manipulation experiment. Global Change Biology 11: 537-552. doi:10.1111/j.1365-2486.2005.00927.x.
 - Walker, M.D., Wahren, C.H., Hollister, R.D., Henry, G.H.R., Ahlquist, L.E., Alatalo, J.M., Bret-Harte, M.S., Calef, M.P., Callaghan, T.V., Carroll, A.B., Epstein, H.E., Jónsdóttir, I.S., Klein, J.A., Magnusson, B., Molau, U., Oberbauer, S.F., Rewa, S.P., Robinson, C.H., Shaver, G.R., Suding, K.N., Thompson, C.C., Tolvanen, A., Totland, O., Turner, P.L., Tweedie, C.E., Webber, P.J., and Wookey, P.A. 2006. Plant community responses to experimental warming across the tundra biome. Proc Natl Acad Sci USA 103: 1342– 1346. doi:10.1073pnas.0503198103.
 - Wang, P., Heijmans, M.M.P.D., Mommer, L., van Ruijven, J., Maximov, T.C., and Berendse, F. 2016. Belowground plant biomass allocation in tundra ecosystems and its relationship with temperature. Environ. Res. Lett. 11: 055003. doi:10.1088/1748-9326/11/5/055003.
 - Welker, J.M., Fahnestock, J.T., Henry, G.H.R., O'Dea, K.W., and Chimner, R.A. 2004. CO₂ exchange in three Canadian High Arctic ecosystems: response to long-term experimental warming. Global Change Biol 10: 1981–1995. doi:10.1111/j.1365-2486.2004.00857.x.
- 882 Welshofer, K.B., Zarnetske, P.L., Lany, N.K., and Thompson, L.A.E. 2018. Open-top chambers 883 for temperature manipulation in taller-stature plant communities. Methods Ecol Evol 9: 884 254-259. doi:10.1111/2041-210X.12863.
- 885 Westergaard-Nielsen, A., Christiansen, C.T., and Elberling, B. 2021. Growing season leaf 886 carbon:nitrogen dynamics in Arctic tundra vegetation from ground and Sentinel-2

887 888	observations reveal reallocation timing and upscaling potential. Remote Sensing of Environment 262 : 112512. doi:10.1016/j.rse.2021.112512.
889 890 891 892 893	 Wolkovich, E.M., Cook, B.I., Allen, J.M., Crimmins, T.M., Betancourt, J.L., Travers, S.E., Pau, S., Regetz, J., Davies, T.J., Kraft, N.J.B., Ault, T.R., Bolmgren, K., Mazer, S.J., McCabe, G.J., McGill, B.J., Parmesan, C., Salamin, N., Schwartz, M.D., and Cleland, E.E. 2012. Warming experiments underpredict plant phenological responses to climate change. Nature 485: 494–497. doi:10.1038/nature11014.
894 895 896 897	 Wookey, P.A., Parsons, A.N., Welker, J.M., Potter, J.A., Callaghan, T.V., Lee, J.A., and Press, M.C. 1993. Comparative Responses of Phenology and Reproductive Development to Simulated Environmental Change in Sub-Arctic and High Arctic Plants. Oikos 67: 490–502. [Nordic Society Oikos, Wiley]. doi:10.2307/3545361.
898 899 900 901	Yang, Y., Halbritter, A.H., Klanderud, K., Telford, R.J., Wang, G., and Vandvik, V. 2018. Transplants, Open Top Chambers (OTCs) and Gradient Studies Ask Different Questions in Climate Change Effects Studies. Front. Plant Sci. 9: 1574. doi:10.3389/fpls.2018.01574.
902 903 904	Yang, Y., Wang, G., Klanderud, K., Wang, J., and Liu, G. 2015. Plant community responses to five years of simulated climate warming in an alpine fen of the Qinghai–Tibetan Plateau. Plant Ecology & Diversity 8: 211–218. doi:10.1080/17550874.2013.871654.
905 906 907 908	Yang, Y., Wang, G., Klanderud, K., and Yang, L. 2011. Responses in leaf functional traits and resource allocation of a dominant alpine sedge (Kobresia pygmaea) to climate warming in the Qinghai-Tibetan Plateau permafrost region. Plant Soil 349 : 377–387. doi:10.1007/s11104-011-0891-y.
909 910 911 912	van Zuijlen, K., Klanderud, K., Dahle, O.S., Hasvik, Å., Knutsen, M.S., Olsen, S.L., Sundsbø, S., and Asplund, J. 2022. Community-level functional traits of alpine vascular plants, bryophytes, and lichens after long-term experimental warming. Arctic Science: 1–15. doi:10.1139/as-2020-0007.
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925 Fig. 1. Photographs of open top chambers (OTCs). Images are of warming experiments at
926 Utqiaġvik, Alaska USA (upper left, photo credit Robert Hollister); Latnja, Sweden (upper right,
927 photo credit Mario Rudner); Alexandra Fjord, Ellesmere Island Canada (lower left, credit
928 Cassandra Elphinstone); and Finse, Norway (lower right, photo credit Kari Klanderud).





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Fig. 2. Range of design dimensions for most commonly implemented hexagonal open top
chambers (redrawn from Molau and Mølgaard 1996 and Hollister 1998). The size can vary, the
corners are 120° angle and can be braced with a bracket or the materials can be longer on one
side and bent to a 60° angle.





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Fig. 4. The average magnitude of OTC warming at Atqasuk, Alaska USA; Latnja, Sweden;
Finse, Norway; and three sites at Alexandra Fjord, Ellesmere Island Canada. The dotted line
represents the overall average. The daily course of warming was compiled for the summer
months (June, July, and August) (unpublished data). The smoothness of the curve is a result of
more years of observation (Atqasuk 1998-2021, Latnja 2020-2021, Finse 2019, Alexandra Fjord
Cassiope and Willow 2008-2019, Alexandra Fjord Dryas 2000-2019).

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Fig. 5. The course of temperature and relative humidity over representative days in OTCs (red
dashed) and adjacent control plots (blue solid) (redrawn from Hollister 1998). Note, these
readings are from a site with drier soils; in areas with higher moisture or standing water, relative
humidity may be higher inside the OTCs and condensation may form on the inside of chamber
walls (Bjorkman 2015).



Fig. 6. Change in the daily maximum, mean and minimum temperatures due to OTC warming.
Points show average temperature differences from 1994-2018 at Utqiaġvik, Alaska USA; lines
show the 2-week running mean for minimum (periwinkle), mean (grey) and maximum (magenta)
daily temperatures (unpublished data).



Fig. 7. Range of surface temperature observed by infrared photography of OTC (open red bars)
and control (solid blue bars) plots (redrawn from Healey et al. 2016). The histogram represents
surface temperatures observed in the Utqiaġvik dry plots near mid-day on August 4, 2014; the
spatial resolution was approximately 3 mm².



968 Fig. 8. Warming effect of the OTCs (relative to control plots) at Atgasuk, Alaska USA and three 969 sites at Alexandra Fjord, Ellesmere Island Canada. Lines represent the average daily temperature 970 difference (OTC minus control) of each year, the thick blue line is a GAM-smoothed curve for 971 the mean temperature difference across all years. Air temperatures were measured at a height of 972 10 to 15cm. The OTCs are installed for the summer only at Atgasuk and remain in place year-973 round at Alexandra Fjord (redrawn from Bjorkman 2015 for the Dryas site and unpublished data 974 compiled according to the methods in Bjorkman 2015); therefore, differences in air temperature above or within the snowpack during the winter at Atgasuk are due to differences in snow 975 976 properties which vary greatly between years. At Alexandra Fjord, OTC impacts on above ground 977 temperature greatly across the year and are greatest during the winter due to the insulative 978 properties of the changed snow regimes.



Fig. 9. Inflorescence length of *Carex aquatilis* measured at the end of the summer at Atqasuk
(triangles) and Utqiaġvik (squares) in OTCs (open red symbols) and ambient plots (closed blue
symbols) graphed against thawing degree days measured from snowmelt until August 15
(redrawn and extended from Hollister et al. 2005).