1 Environmental parasitology: stressor effects on aquatic parasites

2

| 3 | Bernd Sures ^{1,2,3*} , Milen Nachev ^{1,2} , Jessica Schwelm ^{1,2,3} , Daniel Grabner ^{1,2} , Christian Selbach ^{2,4} |
|----|---|
| 4 | ¹ Aquatic Ecology, University of Duisburg-Essen, Essen, Germany |
| 5 | ² Centre for Water and Environmental Research, University of Duisburg-Essen, Essen, |
| 6 | Germany |
| 7 | ³ Research Center One Health Ruhr, Research Alliance Ruhr, University Duisburg-Essen, |
| 8 | Essen, Germany |
| 9 | ⁴ Freshwater Ecology Group, Department of Arctic and Marine Biology, UiT – The Arctic |
| 10 | University of Norway, N-9037 Tromsø, Norway |
| 11 | * Correspondence: <u>Bernd.Sures@uni-due.de</u> (B. Sures) |
| 12 | |
| 13 | Keywords: ecosystem health, habitat alteration, global warming, pollution, climate change, |
| 14 | Anthropocene |
| 15 | |
| 16 | Abstract |
| 17 | Anthropogenic stressors are causing fundamental changes in aquatic habitats and to the |
| 18 | organisms inhabiting these ecosystems. Yet, we are still far from understanding the diverse |

stressors will affect host-parasite communities. Here, we provide an overview of the impacts of
major stressors affecting aquatic ecosystems in the Anthropocene (habitat alteration, global
warming, and pollution) and highlight their consequences for aquatic parasites at multiple levels
of organisation, from the individual to the community level. We provide directions and ideas

responses of parasites and their hosts to these environmental stressors and predicting how these

for future research to better understand responses to stressors in aquatic host-parasite systems.

25

Postprint

26 Aquatic parasites in the Anthropocene

27 Human impacts on the world are manifold, many of them being linked to negative effects on the global ecology. Such anthropogenic influences have increased greatly in recent decades, 28 which is why this era is often summarised as the Anthropocene [1]. Although it is still being 29 30 debated whether this epoch constitutes an official geological unit of time, it is characterised by the fact that humans have become the most influential species on the planet and have caused 31 32 fundamental changes in the atmosphere and the entire ecology of the Earth [1], with farreaching consequences for all levels of organisation of life, including parasites [2]. Freshwater 33 habitats are particularly affected by human impact and subsequent species loss [3,4]. Evaluation 34 35 of historical trends has revealed that freshwaters are diminishing and that these ecosystems are 36 degrading faster than their terrestrial equivalents [5]. Consequently, freshwater environments have higher rates of biodiversity loss than terrestrial ecosystems [3,6]. Accordingly, the recent 37 38 study by Wood et al. [2] for the first time clearly demonstrated a dramatic loss of aquatic parasite diversity in recent decades, which has gone largely unrecognised. In the current review, 39 we therefore focus on possible reasons for human-induced or enhanced aquatic parasite decline 40 and changes in parasite dynamics, with a focus on habitat alteration, global warming, and 41 42 pollution [4], and highlight the effects of these stressors (see Glossary) on the fate of aquatic 43 parasites and their interactions with the associated hosts at different levels of organisation (i.e. from the individual to the community level). This provides an up-to-date synthesis of the 44 advancements, novel findings, and perspectives in environmental parasitology during the last 45 46 decade, and points out key research questions and directions that will significantly advance this field of research. 47

The effects of environmental stressors on host-parasite associations can affect the life cycle of metazoan parasites at various points depending on the characteristics of host-parasite associations such as host specificity, life cycle complexity, transmission strategy, location on the host (endo- vs ectoparasites) and distribution range, to name a few. Parasites may either

directly (stressor effects on parasite stages) or indirectly (stressor effects on hosts) respond to 52 53 an increase or decrease of stressors (Box 1). Overall, the persistence of a parasite species and its population in an ecosystem is determined by the weakest link in the chain: the most sensitive 54 and specific host, or the most delicate parasite life stage. According to theory and recent 55 findings, the diversity of heteroxenous parasites with complex (multi-host) life cycles (e.g. 56 Digenea, Cestoda, Acanthocephala) is thus more likely to decrease in stressful environments 57 58 than the diversity of **monoxenous** parasites (**Box 2**; e.g. Monogenea, Microsporidia) ([2,7–10] and references therein). In contrast to heteroxenous parasites, monoxenous parasites might in 59 fact benefit from the presence of stressors, e.g. if stressors impair the host's defence against 60 61 pathogens [11]. Additionally, an increase in environmental stressors can favour populations of tolerant host species, while populations of more sensitive species disappear. The resulting 62 higher host population density increases the likelihood of transmission of monoxenous parasites 63 64 between hosts [7,12].

Furthermore, specialist parasites will rely on hosts with large and stable populations, as they 65 are not able to switch to other host species, and therefore face a higher risk of secondary 66 extinction [13]. Accordingly, it can be assumed that parasites with simple life cycles 67 (monoxenous, generalist) prevail in stressful environments, while parasites with complex life 68 69 cycles (heteroxenous, specialists) will be more successful under more stable conditions (Box 2). For heteroxenous parasites, the most sensitive host will be the limiting factor for the 70 extinction risk [14]. There is evidence that parasite diversity and overall prevalence decline 71 72 with increasing stressor intensity (e.g., [2,8,15] and that an increase in parasite richness and overall prevalence occurs with the recovery from stressors [16–18]. Due to their complex 73 interaction with stressors or environmental conditions in general, parasites reflect the state and 74 complexity of ecosystems and can therefore eventually be used as bioindicators to assess 75 environmental conditions [19]. 76

In the following sections, we will focus on selected stressors that characterise the Anthropocene, i.e. habitat alteration, global warming and pollution, and highlight their impact on the qualitative and quantitative responses of parasites (**Figure 1**). We also provide perspectives and ideas for future research, which will expand the knowledge in the field of aquatic environmental parasitology and will help to fill the missing gaps.

82

83

Habitat alteration and the occurrence of parasites

Freshwater habitats are particularly affected by degradation and subsequent species loss [3,4]. 84 Numerous human activities lead to habitat alteration and thereby jeopardise freshwater 85 86 ecosystems and their species, including habitat modification, river diversion, fragmentation and 87 flow regulation as well as agricultural and urban landscape expansion [5,20,21]. Accordingly, freshwater ecosystems are degrading faster and have higher rates of biodiversity loss than 88 89 terrestrial ones [3,5,6]. In contrast, functional and resilient ecosystems are characterised by high habitat diversity and species richness (e.g.[22]), which should also be mirrored by a high 90 parasite diversity as the association between parasite and host species richness suggests [23,24]. 91 More specifically, there is indication that parasite species richness is low and total abundance 92 93 is high in stressed ecosystems, while parasite abundance decreases and diversity increases under 94 conditions with absent or low stressor impact (see Box 2; [7]). Based on these findings, the idea of parasites as indicators of biodiversity or 'ecosystem health' was developed [12,25,26], as 95 high parasite diversity is always dependent on diverse and complex interactions among free-96 97 living species [23]. The indicative value of parasites relates to their need of one to several host species as a prerequisite of their occurrence (see a trematode life cycle in Figure 2 as an 98 example). Therefore, life cycle-related traits of parasites are key to understanding and 99 explaining stressor effects on parasite diversity [27]. Particular promising life cycle traits are 100 101 the number of required obligate host species (life cycle complexity) as well as the specialisation 102 of the parasites regarding the host range (generalists vs. specialists) [28].

In contrast to aquatic systems, terrestrial systems are rather well studied regarding the effects 103 104 of habitat alterations on host-parasite assemblages (e.g. [29,30]. For example, a recent longterm study of terrestrial host-parasite systems has shown that habitat loss and fragmentation are 105 likely to have severe impacts on parasites. Food web models predict that parasites with multiple 106 obligate hosts are at greater risk of extinction, as local extinction, or reduction in abundance of 107 one host leads to a bottleneck in the parasite's life cycle. Imbalances in the abundance of 108 109 multiple hosts over time are likely to be common following disturbance, making parasites with complex life cycles particularly vulnerable to habitat fragmentation and other disturbances [29]. 110 Although these results originate from a terrestrial study, they are most likely transferable in 111 112 their broad outlines to aquatic habitats. One of the few aquatic studies that supports this 113 hypothesis is based on the premise that low host abundance and diversity due to environmental degradation and anthropogenic habitat alterations lead to a reduction or even extirpation of 114 115 heteroxenous and specialised parasites [9]. Schwelm et al. [9] studied the trematode fauna of a protected natural reserve in Germany and found a tendency towards higher parasite diversity 116 and prevalence under near-natural conditions. Furthermore, a comparison with other similarly 117 large-scale faunal studies from Europe and North America revealed that life cycle complexity 118 119 (number of obligate hosts involved in the life cycle) is higher in near-natural areas than in those 120 that are more anthropogenically influenced [9]. Global warming might further exacerbate these dynamics [2]. In contrast, no clear pattern was found when investigating monoxenic 121 microsporidian-host associations among near-natural and restored habitats at two German river 122 123 catchments [31]. Microsporidians may have found a way to persist even in a degraded habitat despite low host abundance via resistant spores in the environment and to re-colonise it after 124 stressor release. The ability of host switching [28] and different transmission mechanisms could 125 also contribute to the persistence of members of this parasite group [31]. 126

127 The evidence to date on the relationship between parasites and habitat fragmentation is diverse128 and sometimes contradictory (see [28,29] and references therein), which is not surprising given

the complexity of possible links between habitat alteration, biodiversity loss and parasite prevalence [32,33]. But at the same time this also highlights the importance and necessity of differentiated approaches and investigation of various traits of parasites, their host associations and ecological requirements at an individual, local and global level [29].

In view of the loss of biodiversity with all its devastating consequences, long-term planning 133 and implementation of countermeasures are now urgently needed. Restoration and the 134 135 establishment and expansion of protected areas, such as the recently agreed global expansion of protected areas to a minimum of 30% (https://www.cbd.int/article/cop15-cbd-press-release-136 final-19dec2022) are just a few examples. Parasites are usually ignored in such large-scale 137 138 projects. However, if one follows the assumptions discussed in this chapter, it seems reasonable 139 to assume that protection and restoration measures may lead to an increase not only in freeliving biodiversity in restored and protected areas, but also in parasite diversity. From an 140 ecological perspective, this is of course to be welcomed, as parasites generally not only 141 contribute to biodiversity, but can also show other positive effects on ecosystems (e.g. [34,35]). 142 However, even from a disease ecology perspective, this should not be a concern in principle, as 143 it is likely that dilution effects increase parasite diversity but not abundance per se [36]. 144 145 Fortunately, issues such as parasite extinction and conservation have received more attention 146 recently (e.g. [2,35,37]) and even a global plan for their protection and inclusion in conservation programmes has been developed lately [35]. 147

148

149 Global warming

Global warming is probably the most drastic and far-reaching result of anthropogenic activity on our planet [38]. Climate change encompasses many related and associated effects beyond temperature increases, ranging from changes in the occurrence and severity of extreme weather events to modulation of stratification, water levels, flow regimes or ocean currents [39,40]. However, warming as such has been identified as one of the main drivers of the increased

frequency of parasite disease risks across terrestrial and aquatic ecosystems [41]. Temperature-155 156 driven modifications of parasite life cycles and their possible interactions with other abiotic and biotic parameters are expected to have serious ramifications for aquatic parasites and their hosts, 157 but are hard to predict due to the inherent complexity of host-parasite systems [39]. Various 158 studies have explored the possible impacts of temperature changes on host-parasite interactions, 159 and the implications for disease dynamics, ecological communities, and whole ecosystems. In 160 161 this section, we summarise recent key findings on the diverse, and sometimes unexpected, impacts of temperature changes on individual host-parasite interactions, parasite transmission 162 dynamics and life histories, and ecosystem-wide consequences for aquatic systems. 163

164 The more complex a parasite life cycle is (i.e., the more steps that are required to complete it), the more susceptible the parasite is expected to react to environmental and climate changes, 165 which might lead to neutral, positive, or negative effects on the parasite transmission and 166 167 population dynamics [13]. Among metazoan parasites, digenean trematodes have some of the most complex life histories, often requiring multiple vertebrate and invertebrate hosts, and 168 direct and indirect transmission pathways to complete their life cycle (Figure 2). The exposed 169 free-living transmission stages (miracidia and cercariae) are susceptible to direct environmental 170 influences. Moreover, the asexual production and emergence of cercariae in the first 171 172 intermediate mollusc host as well as the infectivity in the second intermediate host are strongly positively correlated with temperature [42-45]. However, while increases in temperature 173 positively affect cercarial transmission dynamics (e.g., cercarial emission and dispersal 174 activity), these impacts can vary substantially between sympatric trematode species, and 175 warming temperatures might shift parasite community structures in ecosystems rather than 176 177 favour multiple species equally [46]. It remains questionable if these temperature-induced increases in cercarial emergence can be maintained over longer periods of time or are just short-178 term peaks that level off after host acclimation (see [47,48]). 179

Various non-host organisms can interfere with the free-living parasite transmission stages, e.g., 180 181 via predation that leads to a dilution of parasites in the system (e.g., [49,50]. Recent studies have highlighted that temperature-mediated increases in parasite productivity and infectivity 182 can be counteracted by predation on free-living parasite stages, which might potentially offset 183 184 increases in disease risk under climate change [51]. However, it remains uncertain to what extent the elimination of trematode larvae by filter-feeders can compensate for increased 185 186 transmission rates, and if these processes can mitigate disease risks in ecosystems under temperature shifts [52]. Although trematode transmission and infectivity are typically expected 187 to increase under warmer temperatures [39,41,42] with concomitant decreases in host resistance 188 189 to infection [53], trematode transmission windows in coastal systems have been shown to be 190 negatively affected under higher temperatures, which can hinder the possibility for trematodes to flourish in warming oceans [45]. Accordingly, the question which parasites will be able to 191 192 profit in warming marine and freshwater systems will depend on a wide variety of biotic and abiotic factors as well as the ecology of the specific host-parasite systems. Similar varying 193 patterns have been observed for parasitic protists with direct (monoxenous) life cycles in 194 aquatic systems, where the direction and magnitude of the changes in disease occurrence is 195 196 strongly influenced by the ecological features of each disease [54]. This brings into question 197 the generality of the susceptibility of complex vs. simple life cycles to temperature changes and will require further testing in the future that takes the net effect of temperature on a parasite's 198 entire life cycle into account (see [55]). 199

Temperature significantly influences the outcome and strength of species interactions across ecosystems (see [56]). Accordingly, not only parasite transmission dynamics of free-living parasite stages are strongly temperature-dependent, but also the effects of parasitic infections on their hosts and their involvement in food webs can be drastically modulated by temperature. For instance, trematode-infected snails have been shown to change their thermal preference, which likely increases parasite proliferation and transmission [57]. Many parasites that rely on

trophic transmission at some point during their life cycle can modify the behaviour of their 206 207 hosts to facilitate such transmission processes. The plerocercoid stages of the cestode Schistocephalus solidus grow faster in their fish host at higher water temperature, enhancing 208 trophic transmission to final bird hosts as well as parasite reproduction and fecundity, while at 209 the same time shifting their host's thermal preference to warmer temperatures [58]. Different 210 and diverging temperature optima of hosts and parasites have been shown for a range of 211 212 Schistocephalus-stickelback systems, with potentially negative impacts on the hosts and drastic fitness gains for the parasites [59]. Understanding the thermal mismatches between hosts and 213 parasites and identifying the underlying mechanisms of these processes are central requirements 214 215 to improve our predictions of parasite and disease dynamics in warming ecosystems [60]. Contrastingly, field studies from the same host parasite system found little evidence that 216 temperature modified host-parasite interaction, and highlighted the possible difference between 217 218 laboratory experiments and natural settings [61]. Likewise, studies from acanthocephalan hostparasite systems have shown that temperature changes within existing and predicted 219 distribution ranges appear to have no impact on these behavioural changes in hosts and 220 subsequent transmission outcomes [62,63]. Altogether, these findings emphasise the stark 221 222 differences across various host-parasite associations. Moreover, even within the same host-223 parasite system, host genotypes from different geographical regions can show highly varied temperature responses, highlighting that disease and infection dynamics may be difficult to 224 predict within individual parasite species [64,65]. 225

Remarkably, experimental exposure of blue mussels infected with the trematode *Himasthla elongata* revealed a complex parasite-temperature interaction. While moderate parasite loads and increased temperature showed synergistic detrimental effects on mussel survival, high infection intensities appeared to nullify the negative effects of temperature stress on mussels [66]. Other studies revealed that warming and trematode infection increased HSP70 levels and changed the snail's biochemical condition but concluded that this might make them more

susceptible to thermal stress [67]. It therefore remains to be tested if, and under which 232 233 conditions, some parasite infections could provide even a beneficial effect for their hosts under thermal stress, which would contradict our common view that parasites are exclusively harmful 234 to their hosts. Besides these possible impacts of established infections on hosts, recent studies 235 have highlighted the role of non-consumptive or avoidance (i.e., pre-infection) effects that can 236 significantly alter host behaviour and performance in aquatic ecosystems and are potentially 237 238 strongly temperature dependent [68–70]. Overall, these examples underline the complex and 239 sometimes unexpected outcomes of individual host-parasite interactions in warming aquatic ecosystems. 240

241 These individual-level effects on host-parasite systems can translate into ecosystem-wide 242 impacts that structure and regulate populations and communities. Existing outdoor mesocosm approaches could demonstrate strong detrimental synergistic effects of elevated temperature 243 244 and parasitism on the diversity and structure of coastal crustacean communities with potentially negative impacts on the whole coastal habitat [56]. Likewise, recent mesocosm studies from 245 freshwater systems showed similarly complex interactions between parasites and elevated 246 temperature on invertebrate community composition and host abundance that had the potential 247 248 to significantly alter the structure and dynamics in the affected ecosystem [71]. However, such 249 community-level studies that test how parasites and temperature can shape and regulate the 250 functional structure of aquatic ecosystems under climate change pressure remain scarce. Although the examples highlight that temperature effects on parasites and their hosts can have 251 252 far-reaching ecological impacts, the actual relationship between temperature and disease is often complex and the varying underlying mechanisms are only understood for individual 253 parasites and disease agents [51] and references therein. Moreover, the first century-scale time 254 series of fish parasite abundance in North American coastal ecosystems revealed a drastic 255 decline of parasite taxa with complex life cycles (>10% decline per decade) as the result of 256 257 increasing temperature [2]. Future work must therefore continue to bring together experimental approaches investigating mechanistic interactions between host-parasite systems and environmental changes with ecosystem-level studies of climate change effects on populations and communities in aquatic systems [55]. Such approaches will allow to better integrate this knowledge with modelling approaches to more accurately anticipate, monitor and mitigate the impacts of climate-driven disease dynamics in natural ecosystems [72].

263

264 **Pollution**

Elevated pollutant concentrations in aquatic systems are mostly of anthropogenic origin, but 265 geogenic sources (e.g. various metals) might also pose a threat to organisms, including 266 267 parasites. On a global scale, the diversity of environmental contaminants has increased over the last decades in the course of industrialisation and is still increasing every year [73]. As 268 environmental contaminants, a broad spectrum of chemicals of different origin such as organic 269 270 compounds (e.g. PAHs, PCBs, pesticides, drugs, micropollutants, etc.) and their metabolites, as well as nutrients, salts, various metals, organometallic compounds and others are known (e.g. 271 [74,75]). The impact of pollutants as stressors on parasites and their associated hosts depends 272 largely on their concentration. As summarised by Sures and Nachev [10] using the example of 273 274 fish parasites, the mode of action of contaminants can be direct or indirect (see Box 1) which 275 applies not only to fish parasites and their hosts but also to other taxonomic host groups. As a result of pollution impact, parasites respond with changes in their community structure (e.g. 276 considering the ratio of heteroxenous to monoxenous parasites; for details see [15]) and 277 278 diversity [55,76,77], which can be used in environmental parasitology as a measure for assessing stressor intensity. Thus, healthy and diverse aquatic ecosystems (free from pollution 279 280 and other stressors) are generally assumed to have a greater diversity of parasites ([26], see also previous sections). 281

Aqueous pollutants can be accumulated to a high degree by parasites, making them promising
sentinels for pollution (summarised by [8,10]). Accordingly, parasites provide valuable

information on the bioavailability of pollutants and might help in their detection and 284 285 quantification when pollutants are present in very low concentrations in the environment but are accumulated to an enormous degree in the parasites. Various parasitic taxa have been 286 studied in terms of their accumulation capacity, with acanthocephalans, cestodes and nematodes 287 in particular being very sensitive sentinels for metals (see e.g.[8] and references therein). 288 Similarly, a high accumulation capacity has also been demonstrated for some organic 289 290 compounds such as PCBs [78,79]. The use of parasites such as cestodes and acanthocephalans to demonstrate the bioavailability of substances can represent a promising tool for 291 ecotoxicological studies. Detection of certain substances in the body of these taxa is proof that 292 293 these substances must be considered biologically available because they had to cross teguments 294 and membranes due to the lack of a digestive tract in cestodes and acanthocephalans. If microor nanoparticles can be detected in these gutless helminths as shown for particulate platinum-295 296 group metals (e.g. [80]), they must be bioavailable. In addition to metallic nanoparticles, this could also be a promising approach to study the uptake, accumulation, and effects of nano- and 297 microplastic particles in the aquatic environment. Additionally, the high accumulation potential 298 of parasites was found to be beneficial for the hosts by reducing the pollutant burdens within 299 300 the host body and thereby decreasing harmful effects on the physiology (reviewed by [8]; see 301 also [81,82]).

However, high pollutant levels in the environment represent also a driving evolutionary force, 302 as studies on the adaptation of fish populations to high Hg levels reveal (e.g. [83]). It therefore 303 304 seems possible that various parasite taxa might have adapted to specific pollutants and developed tolerance to contamination although evidence from the literature is lacking so far. 305 306 For example, in contrast to the expected patterns of reduced diversity, higher diversity of parasite communities of the intertidal klipfish, Clinus superciliosus, was found at marine sites 307 that had relatively high metal concentrations due to geogenic conditions [84]. In this study, 308 309 acanthocephalans, cestodes and parasitic isopods were more abundant at sites with higher element levels. It therefore can be assumed that the fish hosts involved, as well as their parasites,
have a high tolerance to geogenic background concentrations if the host-parasite systems could
evolve adaptation measures. Similarly, higher abundance and prevalence of acanthocephalans
was found at sites with poorer ecological status according to biological indices and bacterial
loads when comparing different river sections [85].

Besides individual effects due to contaminants, there may also be interactive effects of climate 315 316 change and/or habitat degradation in combination with pollutants. For example, acidification has a significant influence on bioavailability and toxicity of pollutants, both of which increase 317 for metals at lower pH levels [86]. More diverse microhabitats and a higher diversity of free-318 319 living organisms (especially plant communities) on the other hand may act as pollutant sinks 320 (e.g. [87]) and thereby reduce concentrations of available pollutants. Extreme weather events (periods of droughts or higher precipitation) can increase the pollution loads as well. For 321 322 example, higher precipitation intensifies the process of erosion within drainage areas of aquatic habitats and contaminants of anthropogenic and geogenic origin can enter the water bodies with 323 surface run-off [75,88]. Several studies on aquatic parasites showed that parasites react 324 sensitively to anthropogenic activities in the drainage area such as increased urbanisation and 325 326 pollution [89] and to extended land (habitat) destruction such as clearcutting [90]. Marcogliese 327 et al. [90] already provided evidence of the effects of precipitation events on the composition 328 of parasite communities of fish, which were associated with changes in discharge, flow velocity and overall water quality. Such qualitative and quantitative changes can be expected to affect 329 330 the composition, bioavailability and toxicity of pollutants. The latter can be similarly affected by droughts, with high evaporation rates leading to an increase of concentrations in the water 331 column of small aquatic ecosystems. In addition to this, high temperatures lead to reduced 332 aqueous oxygen concentrations and changes in the redox potential, which results in different 333 speciation of metals and an increase of their bioavailability as well [86]. In such scenarios, a 334 335 complex set of stressors (high temperatures, anoxia and elevated pollution) might impose severe effects on free-living communities and their associated parasites (see e.g. [91]). Although a large number of studies related to pollution and parasites have been published in the last decades, there are still various knowledge gaps, which represent a good starting point for future studies on the interaction between parasites and pollutants (see **Box 3**).

340

341 Future perspectives

342 Many of the stressors outlined above affect host-parasite systems on the molecular and biochemical level and the parasitized hosts often show a pattern of response different from 343 uninfected hosts. Numerous studies addressed this topic in the last decades to obtain a more 344 345 general understanding of the combined effects of environmental stressors and parasites on the hosts involved in the life cycle. The outcomes largely depend on the host-parasite system and 346 the respective stressors studied, with increasing and decreasing effects of the parasites on the 347 348 stressor response (see reviews by [8,92]). Even for the same group of parasites (trematodes) the response to a stressor (herbicide) in the same snail host species, can be opposite regarding 349 cercarial emergence and host mortality [93]. Parasites not only affect the response of their hosts 350 to stressors but vice versa, specific stressors can also change the host response to parasites. For 351 352 example, the thiol concentrations increased with the abundance of a parasitic nematode 353 (Oswaldocruzia sp.) in Northern leopard frogs if atrazine exposure was low but decreased in infected frogs under high atrazine exposure levels [94]. Even though the individual results 354 obtained are highly valuable, we are still far from a general understanding of the physiological 355 356 response of parasitised hosts that are additionally affected by other stressors and related community effects. Therefore, we highlight the suggestions for future research in this field 357 shown in **Box 3** to achieve deeper insight into host-parasite interactions under multiple stressor 358 359 exposure.

360 Concluding remarks

Although recent studies have brought major advancements in our understanding of the effects 361 362 of stressors on host-parasite systems, there are still major challenges to face. For many ecosystems, our knowledge on the parasite communities remains limited and life cycles of many 363 parasites are not fully known. Taking all research approaches into account, the overarching 364 vision should be to understand why the responses of the hosts to the combination of parasite 365 and other stressors are sometimes contradictory in different species and to be able to generalise 366 the results to build models that allow predictions of stressor impacts (see Outstanding 367 questions). Ultimately, these findings will guide us on the path to preserve biodiversity, both 368

- 369 free-living and parasitic, from further decline.
- 370

371 Acknowledgements

This paper is based on projects within the Collaborative Research Center (CRC) RESIST (A06,

A09) funded by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation)

- 374 CRC 1439/1 project number: 426547801.
- 375

376 **References**

- Höfele, P. et al. (2022) Introduction: The role of nature in the Anthropocene Defining and reacting to a new geological epoch. Anthr. Rev. 9, 129–138
- Wood, C.L. et al. (2023) A reconstruction of parasite burden reveals one century of
 climate-associated parasite decline. Proc. Natl. Acad. Sci. 120, 1–9
- 381 3. Tickner, D. et al. (2020) Bending the curve of global freshwater biodiversity loss: An
 amergency recovery plan. Bioscience 70, 330–342
- 383 4. Birk, S. et al. (2020) Impacts of multiple stressors on freshwater biota across spatial
 384 scales and ecosystems. Nat. Ecol. Evol. 4, 1060–1068
- Albert, J.S. et al. (2021) Scientists' warning to humanity on the freshwater biodiversity
 crisis. Ambio 50, 85–94
- Turak, E. et al. (2017) Essential biodiversity variables for measuring change in global
 freshwater biodiversity. Biol. Conserv. 213, 272–279
- Chapman, J.M. et al. (2015) Variation in parasite communities and health indices of juvenile *Lepomis gibbosus* across a gradient of watershed land-use and habitat quality.
 Ecol. Indic. 57, 564–572
- Sures, B. et al. (2017) Parasite responses to pollution: what we know and where we go
 in 'Environmental Parasitology.' Parasites Vectors 10, 1–19
- Schwelm, J. et al. (2021) Rare inventory of trematode diversity in a protected natural
 reserve. Sci. Rep. 11, 22066

- Sures, B. and Nachev, M. (2022) Effects of multiple stressors in fish: how parasites and contaminants interact. Parasitology 149, 1822–1828
- Coors, A. et al. (2008) Pesticide exposure strongly enhances parasite virulence in an invertebrate host model. Oikos 117, 1840–1846
- 400 12. Marcogliese, D.J. (2005) Parasites of the superorganism: Are they indicators of
 401 ecosystem health? Int. J. Parasitol. 35, 705–716
- 402 13. Cizauskas, C.A. et al. (2017) Parasite vulnerability to climate change: An evidence-based
 403 functional trait approach. R. Soc. Open Sci. 4
- Lafferty, K.D. and Kuris, A.M. (2009) Parasites reduce food web robustness because
 they are sensitive to secondary extinction as illustrated by an invasive estuarine snail.
 Philos. Trans. R. Soc. B Biol. Sci. 364, 1659–1663
- 407 15. Pérez-del Olmo, A. et al. (2007) Parasite communities in *Boops boops* (L.) (Sparidae)
 408 after the Prestige oil-spill: Detectable alterations. Mar. Pollut. Bull. 54, 266–276
- Huspeni, T.C. and Lafferty, K.D. (2004) Using larval trematodes that parasitize snails to
 evaluate a saltmarsh restoration project. Ecol. Appl. 14, 795–804
- 411 17. Santiago Bass, C. and Weis, J.S. (2008) Increased abundance of snails and trematode
 412 parasites of *Fundulus heteroclitus* (L.) in restored New Jersey wetlands. Wetl. Ecol.
 413 Manag. 16, 173–182
- Miura, O. et al. (2019) Prevalence and species richness of trematode parasites only
 partially recovers after the 2011 Tohoku, Japan, earthquake tsunami. Int. J. Parasitol. 49,
 1023–1028
- 417 19. Sures, B. et al. (2017) Parasites as drivers of key processes in aquatic ecosystems: Facts and future directions. Exp. Parasitol. 180, 141–147
- 20. Dudgeon, D. (2019) Multiple threats imperil freshwater biodiversity in the
 Anthropocene. Curr. Biol. 29, R960–R967
- 421 21. Grill, G. et al. (2019) Mapping the world's free-flowing rivers. Nature 569, 215–221
- 422 22. Rideout, N.K. et al. (2022) Environmental filtering of macroinvertebrate traits influences
 423 ecosystem functioning in a large river floodplain. Funct. Ecol. 36, 2791–2805
- 424 23. Lafferty, K.D. (2012) Biodiversity loss decreases parasite diversity: Theory and patterns.
 425 Philos. Trans. R. Soc. B Biol. Sci. 367, 2814–2827
- 426 24. Johnson, P.T.J. et al. (2016) Habitat heterogeneity drives the host-diversity-begets427 parasite-diversity relationship: evidence from experimental and field studies. Ecol. Lett.
 428 19, 752–761
- 429 25. Selbach, C. et al. (2022) Bridging the gap: aquatic parasites in the One Health concept.
 430 Trends Parasitol. 38, 109–111
- 431 26. Hudson, P.J. et al. (2006) Is a healthy ecosystem one that is rich in parasites? Trends
 432 Ecol. Evol. 21, 381–5
- 433 27. Llopis-belenguer, C. et al. (2019) Towards a unified functional trait framework for
 434 parasites. Trends Parasitol. 35, 972-982
- 28. Cable, J. et al. (2017) Global change, parasite transmission and disease control: lessons
 from ecology. Philos. Trans. R. Soc. Lond. B Biol. Sci. 372
- 437 29. Bitters, M.E. et al. (2022) Experimental habitat fragmentation disrupts host-parasite
 438 interaction over decades via life-cycle bottlenecks. Ecology 103, 1–11
- 30. Gottdenker, N.L. et al. (2014) Anthropogenic land use change and infectious diseases:
 A review of the evidence. Ecohealth 11, 619–632
- 441 31. Prati, S. et al. (2022) Generalist parasites persist in degraded environments: a lesson
 442 learned from microsporidian diversity in amphipods. Parasitology 149, 973–982
- 443 32. Faust, C.L. et al. (2017) Null expectations for disease dynamics in shrinking habitat:
 444 Dilution or amplification? Philos. Trans. R. Soc. B Biol. Sci. 372
- 33. Wood, C.L. et al. (2016) Does biodiversity protect humans against infectious disease?
 Reply. Ecology 97, 542–545

- 447 34. Wood, C.L. and Johnson, P.T.J. (2015) A world without parasites: Exploring the hidden
 448 ecology of infection. Front. Ecol. Environ. 13, 425–434
- 449 35. Carlson, C.J. et al. (2020) A global parasite conservation plan. Biol. Conserv. 250, 108596
- 451 36. Johnson, P.T.J. and Thieltges, D.W. (2010) Diversity, decoys and the dilution effect:
 452 How ecological communities affect disease risk. J. Exp. Biol. 213, 961–970
- 453 37. Kwak, M.L. et al. (2020) Methods for the assessment and conservation of threatened
 454 animal parasites. Biol. Conserv. 248
- 38. Steffen, W. et al. (2018) Trajectories of the earth system in the Anthropocene. Proc. Natl.
 Acad. Sci. U. S. A. 115, 8252–8259
- 457 39. Marcogliese, D.J. (2001) Implications of climate change for parasitism of animals in the aquatic environment. Can. J. Zool. 79, 1331–1352
- 40. Marcogliese, D.J. (2008) The impact of climate change on the parasites and infectious diseases of aquatic animals. OIE Rev. Sci. Tech. 27, 467–484
- 461 41. Harvell, C.D. et al. (2002) Climate warming and disease risks for terrestrial and marine
 biota. Science. 296, 2158–2162
- 463 42. Poulin, R. (2006) Global warming and temperature-mediated increases in cercarial
 464 emergence in trematode parasites. Parasitology 132, 143–151
- 465 43. Thieltges, D.W. and Rick, J. (2006) Effect of temperature on emergence, survival and
 466 infectivity of cercariae of the marine trematode *Renicola roscovita* (Digenea:
 467 Renicolidae). Dis. Aquat. Organ. 73, 63–68
- 468 44. Studer, A. et al. (2010) Parasites and global warming: Net effects of temperature on an intertidal host-parasite system. Mar. Ecol. Prog. Ser. 415, 11–22
- 470 45. Díaz-Morales, D.M. et al. (2022) Heat sensitivity of first host and cercariae may restrict
 471 parasite transmission in a warming sea. Sci. Rep. 12, 1174
- 46. Selbach, C. and Poulin, R. (2020) Some like it hotter: trematode transmission under changing temperature conditions. Oecologia 194, 745–755
- 474 47. Shim, K.C. et al. (2013) Variable effects of increased temperature on a trematode parasite
 475 and its intertidal hosts. J. Exp. Mar. Bio. Ecol. 439, 61–68
- 476 48. Paull, S.H. et al. (2015) How temperature shifts affect parasite production: Testing the
 477 roles of thermal stress and acclimation. Funct. Ecol. 29, 941–950
- 478 49. Gopko, M. et al. (2017) Freshwater mussels (*Anodonta anatina*) reduce transmission of
 479 a common fish trematode (eye fluke, *Diplostomum pseudospathaceum*). Parasitology
 480 144, 1971–1979
- 481 50. Selbach, C. et al. (2019) Cercarial behavior determines risk of predation. J. Parasitol.
 482 105, 330
- 483 51. Goedknegt, M.A. et al. (2015) Climate change and parasite transmission: How temperature affects parasite infectivity via predation on infective stages. Ecosphere 6
- 485 52. Gopko, M. et al. (2020) Parasite transmission in aquatic ecosystems under temperature
 486 change: effects of host activity and elimination of parasite larvae by filter-feeders. Oikos
 487 129, 1531–1540
- 488 53. Leicht, K. and Seppälä, O. (2014) Infection success of *Echinoparyphium aconiatum*489 (Trematoda) in its snail host under high temperature: Role of host resistance. Parasites
 490 Vectors 7, 192
- 491 54. Louhi, K.R. et al. (2010) Is the population genetic structure of complex life cycle parasites determined by the geographic range of the most motile host? Infect. Genet.
 493 Evol. 10, 1271–1277
- 494 55. Marcogliese, D.J. (2016) The distribution and abundance of parasites in aquatic
 495 ecosystems in a changing climate: More than just temperature. Integr. Comp. Biol. 56,
 496 611–619
- 497 56. Mouritsen, K.N. et al. (2018) Coastal ecosystems on a tipping point: Global warming

498 and parasitism combine to alter community structure and function. Glob. Chang. Biol. 499 24, 4340–4356 57. Wang, S.Y.S. et al. (2019) Trematode parasite infection affects temperature selection in 500 aquatic host snails. Physiol. Biochem. Zool. 92, 71-79 501 Macnab, V. and Barber, I. (2012) Some (worms) like it hot: Fish parasites grow faster in 58. 502 warmer water, and alter host thermal preferences. Glob. Chang. Biol. 18, 1540-1548 503 59. Franke, F. et al. (2019) Consequences of divergent temperature optima in a host-parasite 504 system. Oikos 128, 869-880 505 Rohr, J.R. and Cohen, J.M. (2020) Understanding how temperature shifts could impact 506 60. infectious disease. PLoS Biol. 18 507 61. Granroth-Wilding, H.M. V and Candolin, U. (2022) No strong associations between 508 temperature and the host-parasite interaction in wild stickleback. J. Fish Biol. 101, 453-509 510 463 62. Labaude, S. et al. (2020) Increased temperature has no consequence for behavioral 511 manipulation despite effects on both partners in the interaction between a crustacean host 512 and a manipulative parasite. Sci. Rep. 10, 11670 513 Fanton, H. et al. (2021) Effects of temperature and a manipulative parasite on the 514 63. swimming behaviour of Gammarus pulex in flowing water. Hydrobiologia 848, 4467-515 516 4476 64. Leicht, K. et al. (2017) Potential for adaptation to climate change: Family-level variation 517 518 in fitness-related traits and their responses to heat waves in a snail population. BMC 519 Evol. Biol. 17 Santos, J.L. and Ebert, D. (2022) The effects of temperature and host-parasite 520 65. interactions on parasite persistence in a planktonic crustacean. J. Freshw. Ecol. 37, 555-521 568 522 Selbach, C. et al. (2020) Temperature-parasite interaction: do trematode infections 523 66. protect against heat stress? Int. J. Parasitol. 50, 1189-1194 524 Díaz-Morales, D.M. et al. (2023) Parasitism enhances gastropod feeding on invasive and 67. 525 native algae while altering essential energy reserves for organismal homeostasis upon 526 warming. Sci. Total Environ. 863 527 Behringer, D.C. et al. (2018) Parasite avoidance behaviours in aquatic environments. 528 68. 529 Philos. Trans. R. Soc. B Biol. Sci. 373 69. Buck, J.C. et al. (2018) Ecological and evolutionary consequences of parasite avoidance. 530 Trends Ecol. Evol. 33, 619-632 531 532 70. Selbach, C. and Mouritsen, K.N. (2020) Mussel shutdown: Does the fear of trematodes regulate the functioning of filter feeders in coastal ecosystems? Front. Ecol. Evol. 8 533 71. Friesen, O. et al. (2021) Temperature and multiple parasites combine to alter host 534 535 community structure. Oikos 130, 1500-1511 Altizer, S. et al. (2013) Climate change and infectious diseases: From evidence to a 72. 536 predictive framework. Science. 341, 514-519 537 Zimmermann, S. and Sures, B. (2023) Environmental Toxicology. In Drug Discovery 538 73. and Evaluation: Safety and Pharmacokinetic Assays (Franz J. Hock, Michael R. 539 Gralinski, M. K. P., ed), Springer-Verlag GmbH 540 541 74. Rothe, L.E. et al. (2021) Effects of conventionally-treated and ozonated wastewater on mortality, physiology, body length, and behavior of embryonic and larval zebrafish 542 (Danio rerio). Environ. Pollut. 286 543 75. Kontchou, J.A. et al. (2023) Pollutant load and ecotoxicological effects of sediment from 544 stormwater retention basins to receiving surface water on Lumbriculus variegatus. Sci. 545 Total Environ. 859 546 Nachev, M. and Sures, B. (2009) The endohelminth fauna of barbel (Barbus barbus) 547 76. correlates with water quality of the Danube River in Bulgaria. Parasitology 136, 545-548

- 549 552
- 550 77. Krause, R.J. et al. (2010) Parasite fauna of *Etheostoma nigrum* (Percidae:
 551 Etheostomatinae) in localities of varying pollution stress in the St. Lawrence River,
 552 Quebec, Canada. Parasitol. Res. 107, 285–294
- 553 78. Brázová, T. et al. (2012) Bioaccumulation of six PCB indicator congeners in a heavily
 554 polluted water reservoir in Eastern Slovakia: Tissue-specific distribution in fish and their
 555 parasites. Parasitol. Res. 111, 779–786
- 556 79. Le, Y.T.T. et al. (2014) Accumulation of persistent organic pollutants in parasites.
 557 Chemosphere 108, 145–151
- Sures, B. et al. (2005) The intestinal parasite *Pomphorhynchus laevis* as a sensitive accumulation indicator for the platinum group metals Pt, Pd, and Rh. Environ. Res. 98, 83–88
- 81. Molbert, N. et al. (2020) Potential benefits of acanthocephalan parasites for chub hosts
 in polluted environments. Environ. Sci. Technol. 54, 5540–5549
- Molbert, N. et al. (2021) Parasitism reduces oxidative stress of fish host experimentally
 exposed to PAHs. Ecotoxicol. Environ. Saf. 219
- 83. Weis, J.S. and Weis, P. (1989) Tolerance and stress in a polluted environment.
 Bioscience 39, 89–95
- 567 84. Erasmus, A. et al. (2022) High element concentrations are not always equivalent to a
 568 stressful environment: differential responses of parasite taxa to natural and
 569 anthropogenic stressors. Mar. Pollut. Bull. 184
- 570 85. Fanton, H. et al. (2022) Acanthocephalan parasites reflect ecological status of freshwater
 571 ecosystem. Sci. Total Environ. 838
- 572 86. Merian, E. (2004) *Elements and Their Compounds in the Environment: Occurrence,*573 *Analysis and Biological Relevance*, Wiley-VCH Verlag GmbH & Co. KGaA
- Abid Ali Ansari, Sarvajeet Singh Gill, Ritu Gill, Guy R. Lanza, L.N. (2016) *Phytoremediation. Management of Environmental Contaminants, Volume 3*, Springer
 Cham
- 577 88. Schertzinger, G. et al. (2018) Metal accumulation in sediments and amphipods
 578 downstream of combined sewer overflows. Sci. Total Environ. 616–617, 1199–1207
- 579 89. Taglioretti, V. et al. (2018) Fish-trematode systems as indicators of anthropogenic disturbance: Effects of urbanization on a small stream. Ecol. Indic. 93, 759–770
- Marcogliese, D.J. et al. (2016) Variation in parasite communities in spottail shiners
 (*Notropis hudsonius*) linked with precipitation. J. Parasitol. 102, 27–36
- 583 91. Tracy, A.M. et al. (2020) Warming and pollutants interact to modulate octocoral immunity and shape disease outcomes. Ecol. Appl. 30
- 585 92. Grabner, D. and Sures, B. (2019) Amphipod parasites may bias results of 586 ecotoxicological research. Dis. Aquat. Organ. 136, 121–132
- 587 93. Koprivnikar, J. and Walker, P.A. (2011) Effects of the herbicide atrazine's metabolites
 588 on host snail mortality and production of trematode cercariae. J. Parasitol. 97, 822–827
- 589 94. Marcogliese, D.J. et al. (2021) Effects of multiple stressors on northern leopard frogs in agricultural wetlands. Parasitology 148, 827–834
- 591 95. Nadler, S.A. and De Len, G.P.P. (2011) Integrating molecular and morphological approaches for characterizing parasite cryptic species: Implications for parasitology.
 593 Parasitology 138, 1688–1709
- 594 96. Nachev, M. et al. (2017) Understanding trophic interactions in host-parasite associations
 595 using stable isotopes of carbon and nitrogen. Parasites Vectors 10, 90
- 596 97. Gilbert, B.M. et al. (2020) You are how you eat: differences in trophic position of two
 597 parasite species infecting a single host according to stable isotopes. Parasitol. Res. 119,
 598 1393–1400
- 599 98. Cowell, A.N. and Winzeler, E.A. (2019) Advances in omics-based methods to identify

- novel targets for malaria and other parasitic protozoan infections. Genome Med. 11
- 601 99. Cwiklinski, K. and Dalton, J.P. (2022) Omics tools enabling vaccine discovery against fasciolosis. Trends Parasitol. 38, 1068–1079
- 100. Defo, M.A. et al. (2019) Cumulative effects of cadmium and natural stressors
 (temperature and parasite infection) on molecular and biochemical responses of juvenile
 rainbow trout. Aquat. Toxicol. 217
- Lacaze, É. et al. (2019) Cumulative effects of municipal effluent and parasite infection
 in yellow perch: A field study using high-throughput RNA-sequencing. Sci. Total
 Environ. 665, 797–809
- Scharsack, J.P. et al. (2021) Climate change facilitates a parasite's host exploitation via
 temperature-mediated immunometabolic processes. Glob. Chang. Biol. 27, 94–107
- 103. Noreikiene, K. et al. (2020) Humic-acid-driven escape from eye parasites revealed by
 RNA-seq and target-specific metabarcoding. Parasites Vectors 13, 433
- 613 104. Beng, K.C. et al. (2021) Temporal dynamics of freshwater planktonic parasites inferred
 614 using a DNA metabarcoding time-series. Parasitology 148, 1602–1611
- 105. Douchet, P. et al. (2022) Make visible the invisible: Optimized development of an
 environmental DNA metabarcoding tool for the characterization of trematode parasitic
 communities. Environ. DNA 4, 627–641
- 618 106. Thomas, L.J. et al. (2022) Lurking in the water: testing eDNA metabarcoding as a tool
 619 for ecosystem-wide parasite detection. Parasitology 149, 261–269
- 107. Dheilly, N.M. et al. (2019) Parasite microbiome project: Grand challenges. PLoS Pathog.
 15
- Poulin, R. et al. (2022) Inter-individual variation in parasite manipulation of host
 phenotype: A role for parasite microbiomes? J. Anim. Ecol. DOI: 10.1111/13652656.13764

627 Glossary

628 Aqueous pollutants: Pollutants solved in surface or groundwaters.

Bioavailability: The bioavailability describes which proportion of a pollutant present in theenvironment can be taken up by organisms.

Bioindicator: Organisms that show a measurable reaction to environmental impact (stressors)

at the individual (molecular, cellular, tissue) or population level (e.g. population decline).

633 Dilution effect: Removal of free living parasite stages from the environment by non-target-

hosts that can be infected but where the parasite cannot be propagated further, or via predation

of parasite stages. Leading to a decrease of infection pressure on target hosts.

636 **Geogenic sources:** Source of pollutants (e.g. metals) originating from weathering of bedrock.

637 Heteroxenous life cycle: Parasite life cycle involving two or more host species (indirect life638 cycle).

639 Intermediate/final (definitive) hosts: In the intermediate host, the parasite can grow and 640 develop and it can multiply asexually. In the final (definitive) host the parasite reaches the 641 mature adult stage and reproduces sexually.

Microcosm, Mesocosm: Experimental systems that mimics natural conditions in a controlled
setup by using multiple species. Link between field study and highly controlled laboratory
experiment.

645 Monoxenous life cycle: Parasite life cycle involving a single host species (direct life cycle).

646 Stressor: Any impact that leads to environmental variables, individuals, populations,
647 communities, or ecosystem functions exceeding the range of normal variation relative to
648 undisturbed reference conditions.

Box 1. Effects of environmental stressors on host-parasite associations

If generalist host-parasite systems are exposed to deleterious stressors, alternative hosts can take over the role of intermediate or **final hosts** when individual host species fail (indirect effect on parasite) (**Figure IA**), while for specialised parasites the harmful effects of stressors always disrupt the life cycle (**Figure IB**). Besides stressor effects on hosts, the survival and presence of parasites in ecosystems may be determined by direct stressor effects on the parasitic stages (e.g. temperature effects on parasite stages in poikilothermic hosts) or on the free-living larval stages of parasites species (e.g. coracidia, miracidia, cercariae or other larval stages; see **Figure IC**).

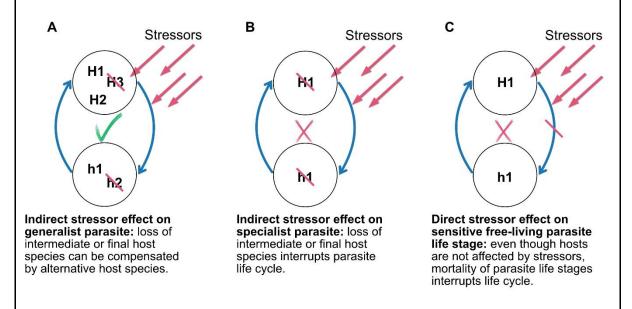


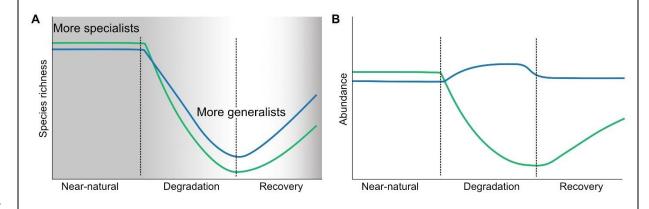
Figure I (in Box 1). Hypothetical life cycles of parasites using final (Hx) and intermediate hosts (hx). The different scenarios (A-C) show possible consequences of severe stressor impacts. Red lines: host lost from system, \checkmark life-cycle maintained, X: life cycle interrupted.

650

651

653 **Box 2.** Assumed development of parasite dynamics over time

In a scenario of environmental degradation (e.g. increasing pollutant concentration in the 654 655 water), species numbers, but also community composition, will change with varying 656 environmental conditions from proportionally more complex life cycles and higher numbers of 657 host specialists under near-natural conditions to lower life cycle complexity and more host 658 generalists under degraded conditions (Figure IA). After stressor release (phase of recovery), 659 the composition of the parasite community is still different from the near-natural situation with 660 a dominance of few species with less complex life cycles, whereas specialists with complex life 661 cycles can only establish once the habitats allow for recolonization with the required host species (Figure IA). The relative abundance of mono- and heteroxenous parasites will also 662 change. Monoxenous parasite communities will become species poor, with few but abundant 663 species that parasitise the remaining hosts during stressor impact (degradation phase) (Figure 664 **IB**). Relative abundance of heteroxenous parasites will decrease in degraded environments. The 665 666 relative abundance of both mono- and heteroxenous parasites will return to the pre-degradationstate, but with delay for heteroxenous species (Figure IB). The observed development of the 667 parasite communities can vary for some stressors (e.g. nutrients) that might, for example, lead 668 669 to an increase of the abundance of heteroxenous parasites due to increasing numbers of intermediate hosts. 670



671 672

Figure I (in Box 2). Hypothetical development of parasite communities before (Nearnatural), during (Degradation) and after the occurrence of a stressor (Recovery) in an

| 674 | aquatic system. A: Changes in species richness. B: Changes in parasite abundance. Green line: |
|-----|--|
| 675 | heteroxenous parasites. Blue line: monoxenous parasites. Grey areas indicate higher number of |
| 676 | host specialist parasites, while more light areas indicate higher numbers of generalists. |
| 677 | The graphs show a hypothetical situation of a stressor intensity that is strong enough to affect |
| 678 | communities but without complete eradication of all parasite species. |
| 679 | |

Box 3. Directions for future research

Research foci on model host-parasite systems

Studies applying different stressors to the same host-parasite system or the same stressor to a different host-parasite system with an identical experimental setup will allow the comparison of results and will reveal general trends. This would require more cooperative research projects that allow the development of coordinated experimental protocols between different working groups. A prerequisite is the reliable identification of the studied parasites by appropriate taxonomy specialists including molecular vouchers that prevent the use of morphologically similar, but genetically distinguishable cryptic species in subsequent work [95]. Methodologically, microcosm and mesocosm studies might bridge the gap between field and mere laboratory studies as they provide controlled and replicated conditions, but at the same time allow to study biotic interactions of different species. To our knowledge there are to date only few studies addressing stressor (temperature) effects in a multi-host and multi-parasite setting in experiments [45,56,71]. The responses of different host species were complex and contradictory due to interactions of the species among each other and presence or absences of the parasite in the system had a fundamental effect on the free-living community. Similar studies are required to assess ecosystem effects of parasites that cannot be elucidated when studying single host species.

Clearly defined parasite life strategies

There is a huge variety of life strategies among parasites that differ greatly in their effect on the host. For example, there is a major difference between parasites that are adsorptive feeders (e.g. adult cestodes, acanthocephalans) and those that actively feed on host tissue (e.g. nematodes, monogeneans) which is also reflected by the trophic position of these parasites [96,97]. Similarly, effects on the hosts are also largely dependent on the developmental stage of a given parasite species. Usually, larval parasites inside their intermediate hosts are more detrimental to their hosts than adult parasites within their **definitive hosts** in order to facilitate transmission to the final host (e.g., via trophic transmission, host manipulation, or by producing large numbers for parasite transmission stages). Such differences exist not only between the major groups of parasites, but may also be important for more closely related species (e.g. trematodes forming sporocysts or rediae in the snail intermediate host). Therefore, studies should be designed using parasites with different life strategies under similar conditions (ideally same host, same stressor exposure) with precise descriptions of the relevant life cycle traits (e.g. life cycle complexity, specialisation of the parasites regarding the host range).

Field studies on re-emergence and community formation of parasites after restoration

Theoretical considerations would suggest that host diversity and abundance will be reduced with increasing stressor presence and effects. As a consequence, parasite richness will decrease ([2]; ecosystem degradation, **Box 2**). If stressors reach a threshold level, parasite life cycles will collapse and consequently, parasites will disappear. Upon stressor release and recovery of the system, the number and diversity of generalist parasites with less complex life cycles can be expected to increase, while specialists with complex multi-host life cycles are assumed to re-establish later, after conditions and host populations have recovered (**Box 2**). These theoretical predictions should be tested under "real world conditions" by including analyses of parasite abundance and prevalence data at field sites which had been subjected to ecological restoration.

Description of processes involved in pollutant uptake by parasites

Basic aspects regarding the uptake, storage and detoxification of contaminants by parasites as well as a deeper mechanistic understanding of how pollution and parasites interact require further elucidation. Additionally, there is still a limited number of studies on marine host parasite systems as well as on pollutant accumulation potential of parasites of bird and mammalian hosts. Apart from metal accumulation, studies on uptake of micropollutants (organic compounds and their metabolites) and various micro- and nanoparticles (e.g. plastic, metals) by parasites still remain scarce. Likewise, the role of parasites as sinks for organic pollutants needs to be further investigated, as few initial studies demonstrated the beneficial effects of parasites on concentrations in infected fish [78] and on host physiology [81,82].

Study host transcriptomes to detect the target genes and metabolic pathways affected by stressors and parasites

"Omics" based applications, mainly transcriptomics, have been used successfully to study parasites of medical or veterinarian significance (e.g. [98,99]). However, transcriptome sequencing to detect differential expression of genes in infected or stressor exposed hosts is also suitable for ecological questions. The major advantage to measuring a preselected set of biomarkers via qPCR or using biochemical assays is that genes or pathways regulated during parasite infection or exposure to stressors can be identified. This is particularly relevant if there is no prior knowledge on the effects of the parasite or stressor applied. To date, only few studies have applied transcriptomics to study stressor effects (temperature, pollution or combinations of both) on parasitized hosts in aquatic ecosystems [100–102]. Further research efforts are needed to increase the number of sequenced genomes of relevant hosts and parasites, to improve interpretation and functional annotation of the transcripts. Besides transcriptomes, next generation sequencing can provide further valuable tools e.g. for the assessment of parasite biodiversity using environmental DNA (eDNA) or the identification of (micro-)parasite communities by metabarcoding (e.g. [103–106]).

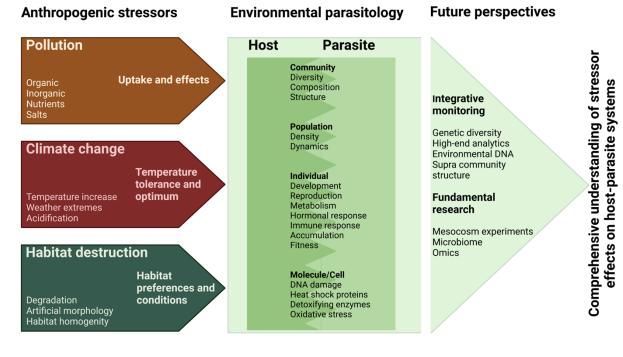
Considering host and parasite microbiomes

Part of the current research is focusing on the study of host and parasite microbiomes [107], which will add another layer of complexity to the research related to environmental parasitology. Differences in the microbiome of host and/or parasite individuals changes their phenotype and might also alter their response to stressors [108]. Therefore, microbiomes might be a promising approach to understand cases of seemingly contradictory results when studying the effects of stressors on hosts and parasites.

681

682

Postprint

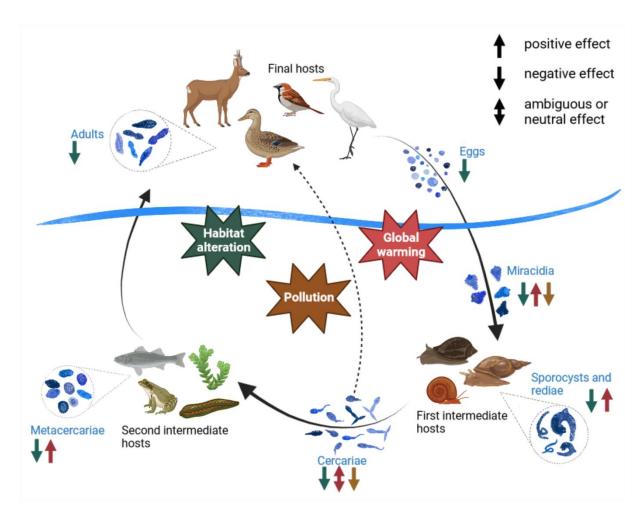


685 Figure 1. Relevance of `Environmental Parasitology` for ecosystem research in times of

686 increasing anthropogenic pressure (Anthropocene). Figure created with BioRender.com

687

688



690

Figure 2. Generalised life cycle of digenean trematodes with final vertebrate hosts, 691 invertebrate and vertebrate intermediate hosts incorporating environmental stressors 692 (habitat alteration, global warming, pollution) and their effects on the parasite stages of 693 the life cycle. Environmental stressors can directly and indirectly (via their hosts) affect parasite 694 reproduction, transmission, abundance and pathogenicity (see examples in text). These stressors 695 can have negative, neutral or positive effects (see arrows, colours represent respective stressors) 696 on the individual steps in the parasite's complex life cycle, highlighting our need to understand 697 both the mechanistic processes and the net effects of environmental stressors on parasite life 698 cycles. Figure created with BioRender.com 699