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Host-associated probiotics, a key factor in sustainable aquaculture?

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Abstract

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The aquaculture industry has dramatically developed during the last two decades. However, this development has, in some cases, resulted in environmental degradation, emergence of diseases and low productivity. The need for improving disease resistance, growth performance, feed efficiency, and safe aquatic production for human consumption has stimulated development and applications of probiotics in aquaculture. Probiotics used in aquaculture include genera of Bacillus, Lactobacillus, Enterococcus, and Carnobacterium, and yeast. However, most of these probiotics are derived from terrestrial sources and not from the environment in which the aquatic animals live or the host animal. The use of "host-associated probiotics" has recently gained attention, as they offer an alternative strategy within aquaculture, which per se is dependent on the use of terrestrial microorganisms. The benefits of host-associated probiotics include improved growth performance, feed value, enzymatic contribution to digestion, inhibit adherence and colonization of pathogenic microorganisms in the gastrointestinal tract, increase hematological parameters, and immune response. The present review addressed insight into the application of host-associated probiotics within aquaculture, with special focus on their immunomodulatory and growth enhancing effects. Furthermore, the current review discusses research gaps and issues that merit further investigations.

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Keywords: Host-associated probiotics, Sustainable aquaculture, Immunomodulation, Growth

promoter, Terrestrial microorganisms; Disease resistance

1. Introduction

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Aquaculture is the fastest growing animal protein sectors to meet the demands of today's growing worldwide population. For the satisfaction of the increased human consumption, intensive aquaculture systems at high densities have been developed (Tal et al., 2009). This intensification has however, resulted in significant damage to the aquaculture environment, due to organic wastes production which consume dissolved oxygen in ponds, and increase toxic metabolites; responsible for high mortality (Martinez Cruz et al., 2012). For long time, chemotherapies and antibiotics were applied as an effective remedy for disease outbreaks in aquatic animals (Serrano, 2005). However, due to their excessive use, aquatic pathogens have become resistant to such treatments, along with the suppression of the host's immune system (Capkin et al., 2015, 2017; Dawood & Koshio, 2016). Furthermore, the application of antibiotics may extend to human consumption, which could adversely affect human health and weaken the immune systems (Cheng et al., 2014; Hoseinifar et al., 2015; Langdon et al., 2016). As a result, strict regulations against antibiotic administration in livestock-, poultry-, and the aquaculture sector have been established in several countries. Alternative strategies such as; vaccines, pro -, pre -, synbiotics, and medicinal herbs must further be investigated to improve health of aquatic animals (Martínez Cruz et al., 2012; Altinok et al., 2015; Adel et al., 2016; Abdelkhalek et al., 2017; Van Doan et al., 2017; Dawood et al., 2018). Host aquatic microorganisms play a crucial role in the health maintenance by boosting the immune system, and increasing the resistance against opportunistic pathogens via the production of antimicrobial substances (e.g. Llewellyn et al., 2014; Ibrahem, 2015; Yan et al., 2017). When discussing the functionality of gut microbiota, it depends on the ability of microorganisms to interact within the gastrointestinal (GI) tract, which benefit the host through influence upon its biological functions (Ramírez & Romero, 2017).

For nearly half a century the word probiotic has been used in several different ways, since Lilly & Stillwell (1965) first used it. Gatesoupe (1999) defined probiotics as "live microbial feed supplements that improve the health of both humans and terrestrial livestock", and according to Dawood et al. (2016), probiotics is required to maintain a healthy environment for aquatic animals and to increase their performance, without any negative impacts upon the consumers. Supplementation of aquatic animal diets with probiotics positively influences growth performance, feed utilization, physiological conditions, optimize cell proliferation under stressful conditions, promote immune cell stimulation, modulate the gut microbiota and improve disease resistance (e.g. Irianto & Austin, 2003; Merrifield et al., 2010; De et al., 2014; Ringø et al., 2014; Hai, 2015a). When discussing disease resistance, the importance of a stable microbiota is worth to mention. The best evidence for this protective effect of the gut microbiota stems from an early study revealing that germfree animals are more susceptible to diseases compared to corresponding conventional animals with a "complete gut microbiota" (Collins & Carter, 1978). Germfree mouse can be killed with 10 cells of Salmonella enteritidis whereas 10⁶ cells are needed to kill conventional mouse with a conventional gut microbiota. Therefore, it is crucial to increase our knowledge on probiotics adhering and colonising the GI tract of endothermic animals as well as aquatic animals, in the context of improved growth performance and health. When discussing the importance of probiotics in aquaculture, the mode of action and their effects on the intestinal microbiota, immunological, physiological responses, as well as growth performance must be continually investigated. Recently, there has been increasing attention towards administration of host-associated probiotics in aquaculture (Lazado et al., 2015; Li et al., 2018). However, as no clear definition per se exist, we defined host-associated probiotics as; bacteria originally isolated from the rearing water or the GI tract of the host to improve growth and health of the host. In aquaculture, per se it is not clear, whether host-associated

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probiotics are more effective than probiotics from other origins, even though there is some evidence demonstrating beneficial effects of host-associated probiotics *vs.* probiotics isolated from other sources (Lazado et al., 2015). One possible reason for their superior function may be because they perform better within their own original environment. Therefore, the present review addressed on the health benefits of host-associated probiotics in aquaculture, and their potential immunomodulatory and growth promoting effects.

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2. Sustainable aquaculture: the role of probiotics

Outbreaks of infectious diseases are a major obstacle for the development of aquaculture, which adversely affect the sustainability of this industry (Mardones et al., 2018). Throughout the last decades, the aquaculture industry was heavily dependent on the use of antibiotics and chemotherapeutics for diseases control and prevention (Romero et al., 2012; Fečkaninová et al., 2017). However, the abusive use of these substances caused many adverse effects upon the host and its environment (e.g. Romero et al., 2012; Miller et al., 2016; Capkin et al., 2017; Suzuki et al., 2017), and therefore development of alternative strategies to antibiotics and chemotherapeutics are needed (e.g. Defoirdt et al., 2011; Altinok et al., 2016; Founou et al., 2016). Sustainable aquaculture has been created to preserve and develop the industry (Bostock et al., 2010). In this context, several criteria have been put forward and include; vaccines, immunostimulants, pro -, pre - and synbiotics (e.g. Ringø et al., 2014; Lazado et al., 2015; Hoseinifar et al., 2016). Several types of vaccines have been developed for aquaculture, in which the most successful effects are reported in salmonids and cyprinids (e.g. Dhar et al., 2014; Ringø et al., 2014; Lazado et al., 2015; Hoare et al., 2017). Recent findings have revealed that vaccines are effective for disease control in aquaculture (e.g. Johnson et al., 2008; Brudeseth et al., 2013; Assefa & Abunna, 2018). However, individual vaccines are known to be effective against only one type of pathogen; which limiting their use in aquaculture (Ardó et al., 2008; Feodorova et al., 2014). Immunostimulants with a wide range of applications and functions have therefore, been considered as an alternative strategy to overcome the "drawback" of vaccines (Barman et al., 2013; Hoseinifar et al., 2017). These bioactive compounds boost the immune system of the host, and protect the host against wide range of pathogens (e.g. Bricknell & Dalmo 2005; Ringø et al., 2012; Hoseinifar et al., 2015; Nawaz et al., 2018). Probiotics have many beneficial properties to the host and its environment, and they are now widely used in aquaculture as growth promoters, immunostimulants, and for disease protection (Pérez-Sánchez et al., 2014; Hai, 2015b). The word probiotic stems from the Greek roots pro and bios, or "profile" (Schrezenmeir & de Vrese, 2001). As a concept, the use of probiotics to control disease is referred to as "the use microorganisms against microorganisms" (Lazado et al., 2015). During the last two decades, numerous comprehensive reviews have addressed on probiotics and their impacts in aquaculture (e.g. Gatesoupe 1999; Verschuere et al., 2000; Irianto & Austin, 2003; Balcázar et al., 2006; Merrifield et al., 2010; Pérez-Sánchez et al., 2014; Ringø et al., 2014; Hai, 2015a,b; Hoseinifar et al., 2016) with multidimensional actions. Probiotics offer numerous beneficial effects, including immunomodulatory, nutritional, and environmental capacity, and therefore have a great competitive advantage as prophylactic against disease (Ohashi & Ushida, 2009; Lazado et al., 2015). While there is no concrete evidence to conclude that probiotics are better than immunostimulants or vaccines, the beneficial effects upon the host and their environment ensure that probiotics will remain one of the most promising approaches used to control diseases and the subsequent environmental modifiers (Newaj-Fyzul & Austin, 2015). Several probiotics species are used in aquaculture; e.g. Bacillus sp., Lactobacillus sp., Enterococcus sp., Streptomyces, Carnobacterium sp., and yeast (e.g. Gatesoupe, 2007; Martínez Cruz et al., 2012; Tan et al., 2016; Elshaghabee et al.,

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2017). Table 1, 2 and 3 summarize the results of studies from 2005 to 2018, focusing on the effects of host-associated probiotics as growth promoters and immunostimulants in aquaculture.

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3. Host-associated vs. terrestrial probiotics

To our knowledge, commercial probiotics have been used in aquaculture since the early 80ties (Boyed et al., 1984) and until today (e.g. Zheng et al., 2017; Zhou et al., 2017; Interaminense et al., 2018; Pereira et al., 2018). However, recently, attention has focused on using hostmicrobiota as a probiotics source (e.g. Lazado et al., 2015; Interaminense et al., 2018; Li et al., 2018; Wang et al., 2018). They are naturally established within the host defense system (Gomez et al., 2013; Llewellyn et al., 2014), and revealed a large number of beneficial characteristics; such as intestinal enzymes and novel bioactive lipid sebastenoic acid (Lazado et al., 2012; Sanchez et al., 2012). This is important for fish farming, as fish are a rich source of probiotic strains that provide additional mechanisms against numerous infectious diseases (e.g. Spanggaard et al., 2000; Pandiyan et al., 2013; Carnevali et al., 2017). Host-related microbiota can be an alternative probiotic source within aquaculture, as terrestrial sources per se are mostly used (Lakshmi et al., 2013; Lazado et al., 2015). Today, most probiotic candidates are derived from the mucosal layers, the autochthonous bacteria, of aquatic animals (Balcázar et al., 2007; Newaj-Fyzul et al., 2007; Caipang et al., 2010; Lazado et al., 2010; Maeda et al., 2014; Lazado & Caipang 2014a; Tzuc et al., 2014). For example, probiotics isolated from the GI tract of channel catfish (Ictalurus punctatus) (Larsen et al., 2014), rainbow trout (Oncorhynchus mykiss) (Spanggaard et al., 2000; Araújo et al., 2015), Atlantic cod (Gadus morhua) (Dhanasiri et al., 2011; Fjellheim et al., 2007), and Atlantic salmon (Salmo salar) (Jöborn et al., 1999). Two basic principles are generally applied as a guide for the use of host-associated bacteria as a probiotic (Lazado & Caipang, 2014a). The differences in physiological peculiarities and discrepancies of each host, as well as the considerable influences of various environmental factors make it hard to establish a probiotic candidate for global application (Lazado et al.,

2015). For example, growth and bacteriocin production of Leuconostoc mesenteroides and Lactobacillus curvatus are affected by pH, temperature, and aquatic animal species (Mataragas et al., 2003). These parameters were reported to affect the adhesion ability of lactic acid bacteria (LAB) (Tuomola et al., 2001). Physiological variances between marine and terrestrial microorganisms exist, and impose differential responses to diverse environment (Morgan-Kiss et al., 2006; Lazado et al., 2015). Furthermore, a discrepancy exists in acquisition mechanism between marine and terrestrial microorganisms involved in inhibition of pathogen growth (Sandy & Butler, 2009), which is controlled by siderophore production; a mechanism of probiotics (Lazado et al., 2010; Ahmed & Holmström, 2014). The mode of action of bacterial iron acquisition includes siderophore-mediated transport, direct import through divalent metal transporters, or direct piracy from ironbound host proteins (Caza & Kronstad, 2013; Pokorzynski et al., 2017). Siderophores are low molecular weight substances with a high affinity for ferric iron, which are synthesized and secreted by bacteria in order to scavenge iron when the availability is limited. In Gram-negative bacteria, siderophore-bound iron is transported through cognate outer membrane receptors that require energy transduction via the TonB-ExbB-ExbD protein complex (Andrews et al., 2003; Ellermann & Arthur, 2017). The second mechanism depends on specific enzymatic hydrolysis of the siderophore, which serves to weaken its interactions with iron, and enable its liberation. In both Gram-negative and Grampositive bacteria, ferrous iron is directly transported through cytoplasmic membrane permeases or ABC transporters (Kammler et al., 1993; Sabri et al., 2006; Shapiro & Wencewicz, 2016). Some bacterial pathogens belonging to family Neisseriaceae, Pasteurellaceae and Moraxellaceae are also capable of utilizing host sources of iron by expressing outer membrane receptors that directly bind to host iron-binding proteins such as transferrin (Cornelissen, 2003; Parrow et al., 2013). The bottlenecks caused by the physio-chemical parameters on probiotic properties may be eliminated by using host-associated microorganisms as probiotics if these

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microorganisms are originally isolated from the environment where they were applied (Verschuere et al., 2000; Vadstein et al., 2013). Because the highest physiological activities of microorganisms are obtained in their natural habitats, we may presume that the ultimate benefits are achieved when they are applied in those conditions (Ibrahem, 2015). Because existing knowledge is fragmentary, it is difficult to draw a solid scientific conclusion to whether host-associated microbiota are superior to terrestrial sources (Lazado et al., 2015). However, it is believed that the survival rate and function of host-associated probiotics are optimal when they are applied in the "natural" environment (Murall et al., 2017). The application of both host and non-host candidate probiotics in aquaculture represent promising future alternatives (Lazado et al., 2015). Additional beneficial effects are obtained when combining two or more microorganisms, providing that they do not inhibit each other. Previous findings have indicated that the functionality and efficacy of their administration are improved with the use of multi-strain probiotics (Timmerman et al., 2004; Mohapatra et al., 2014). In probiotics-mediated disease prevention, the best level of protection for a broad spectrum of fish pathogens can only be achieved through the use of multiple strains in dietary preparations (Merrifield et al., 2010). Interestingly, the use of host-associated probiotics addresses the above concerns, but also shows that host-microbiota have functions other than being a part of natural defense system, such as immunomodulation and nutrient conversion (Lazado et al., 2015; Montalban-Arques et al., 2015). This is bioprospecting in a lesser scale, with a sustainable core concept of improving the health and welfare of aquaculture animals, through the utilization of their own commensal microbes (Llewellyn et al., 2014; Lazado et al., 2015). Recent findings have revealed that the microbial community of fish intestines are a new source of probiotics, and offer a biosynthetic diversity for natural products discovery (Sanchez et al., 2012).

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4. Host specificity

When discussing host-associated *vs.* terrestrial probiotics, host specificity is an important subject to mention. Adhesion of probiotic LAB was previously reported to be host specific, and Fuller (1989) stated, "The attachment to epithelial cells is very host specific which means in practical terms that a strain which is suitable as pig probiotic may not be active in chick and other animals". However, later studies have indicated that LAB originated from one host adhere to mucus of other species (e.g. Tuomola et al., 1999; Rinkinen et al., 2000, 2003; Nikoskelainen et al., 2001a,b). In a recent review by Li et al. (2018), adherence and colonization of microorganisms in the GI tract of fish was discussed. As in vitro and ex vivo studies suggested host specificity of LAB colonization in sturgeon (Askarian et al., 2011; Salma et al., 2011), Li et al. (2018) put forward the controversial hypothesis that host specificity of LAB does not occur in "younger" fish species, e.g. Atlantic cod, Atlantic salmon and turbot, while it occurs in older species, sturgeons; "living fossil". However, adhesion ability of LAB may be related to; adhesive and non-adhesive ability (Zhou et al., 2012), variation of mucin adhesion and cell surface characteristics depending on their isolation habitats indicated by Buntin et al. (2017), and hydrophobic properties (Grajek et al., 2016).

5. Mode of actions of probiotics

Probiotic microorganisms influence the immune responses of the host, as well as the interaction between these responses and their intestinal bacterial communities (Derrien & van Hylckama Vlieg, 2015; Fischbach & Segre, 2016). During the last decades, intensive studies on probiotics have shed new light on the importance of probiotics and their modes of action (Papadimitriou et al., 2015), and several modes of action have been proposed.

(I) Probiotics enhance feed utilization and weight gain promoters of aquatic animals (Hai, 2015b; Zorriehzahra et al., 2016). Probiotics improve the host's appetite and feed

digestion by breaking down indigestible components, increase production of vitamins, and detoxify compounds in the diet (Giraffa et al., 2010; Zorriehzahra et al., 2016).

(II) Competitive exclusion of pathogens in the GI tract (Ringø et al., 2010a), by production of peroxide, bacteriocin, siderophore, and lysozyme enzymes (Yan et al., 2002). The physiological and immunological effects (Mohapatra et al., 2014; De et al., 2014) are considered to be two of the most important beneficial mechanisms of probiotic bacteria (Collado et al., 2010; Zorriehzahra et al., 2016).

(III) Improve the resistance of farmed aquatic animals against stress caused by several environmental hazards throughout the aquaculture activity (Nimrat et al., 2012; Zorriehzahra et al., 2016).

These modes of actions present the beneficial effects of probiotics in cultured aquatic animals.

Nonetheless, future researches on the interaction between probiotics and the host, involving transcriptomic and proteomic analyses, are needed to understand of probiotic activity.

6. Host-associated probiotics and their applications in aquaculture

6.1 Bacillus sp.

Bacteria belonging to genus *Bacillus* has simple nutritional requirements, fast metabolic rate, ease to isolate and preserve, and secrete multiple bacteriocins make them good probiotic candidates (Moriarty, 1998; Ziaei-Nejad et al., 2006). Genus *Bacillus* is the most studied host-associated probiotic, among a wide variety of aquatic animals. To our knowledge, Yanbo and Zirong (2006) conducted the first study using host-associated probiotics in aquaculture. Their findings showed that *Bacillus* sp. supplementation diets significantly improved growth performance, survival rate, and feed conversion ratio; as well as protease and lipase activities in common carp (*Cyprinus carpio*) (Yanbo & Zirong, 2006). The authors suggested that these improvements were due to enhanced enzyme activities, resulting in improved diet digestibility; including proteins, starches, and lipid.

In channel catfish, dietary inclusion of *Bacillus* spp. isolated from oil or intestine, significantly increased channel catfish resistance towards Edwardsiella ictaluri and Aeromonas hydrophila (Ran et al., 2012). The improved diseases resistance was suggested to be attributable to a biologically meaningful discrepancy in the interactions between Bacillus strains and their respective host. Additionally, interactions may exist between host, pathogen, and probiotic bacteria that are influenced by environmental factors. Ran et al. (2012) suggested that more research is needed to clarify the complex interactions between the host, pathogen and the probiotic Bacillus strains, as well as how to improve the environment in order to obtain optimal biological control of disease. A study with *Bacillus* sp. isolated from shrimp intestine, Cha et al. (2013) reported significant increase in growth performance, innate immune response, and disease resistance in olive flounder (Paralichthys olivaceus), to Streptococcus iniae. The authors speculated that the increase in growth performance of fish fed dietary Bacillus sp. was a result of improved diet utilization. The GI tract of fish are known to be very sensitive to nutrients, presenting immediate changes in digestive enzyme activity, leading to enhanced growth and well-being (Bolasina et al., 2006; Shan et al., 2008). Zhang et al. (2010) indicated that the growth improvement in sea cucumber (Apostichopus japonicus) fed a Bacillus subtilis supplemented diet was due to the increase in digestive activity via enhanced enzymatic activity. Previous probiotic studies suggested that supplementation increases the favorable microorganism in the host's gut, which in turn release exoenzymes that can improve feed utilization and health conditions (Vine et al., 2006; Yanbo & Zirong, 2006). These beneficial organisms secrete numerous proteases, which may play an important role in breakdown of peptide bonds of proteins, into monomers and free amino acids, which positively affect host's nutritional status (Macfarlane & Macfarlane, 2012). Similarly, a significant increase in growth performance, immune response, and disease resistance were observed in orange-spotted grouper (Epinephelus coioides) larvae (Sun et al.,

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2013), rohu (Labeo rohita) (Ramesh et al., 2015), and striped catfish (Pangasianodon hypophthalmus) (Truong Thy et al., 2017) fed host-associated Bacillus sp. The authors suggested that further investigations are needed to evaluate the effect of probiotics on the immune function in fish larvae, and to estimate the duration of persistence of the *Bacillus* after discontinuation of probiotic supplementation. There is limited information available regarding the effects of host-associated probiotics on the health and well-being of crustacean and mollusk. Nimrat et al. (2011) reported that giant tiger prawn (*Penaeus monodon*) fed *Bacillus* spp. isolated from shrimp intestines and pond sediment, presented significantly improved growth performance, and survival rate; as well as increasing the number of additional beneficial microbial probiotics. In mud crab (Scylla paramamosain), dietary administration of *Bacillus* spp. isolated from the crab's intestine significantly enhanced survival rate, respiratory burst activity, immune related genes expression, and resistance to Vibrio parahaemolyticus (Wu et al., 2014). A significant decrease in hemocytes mortality and increase resistance toward Vibrio harveyi were observed in Pacific oyster (Crassostrea gigas) fed Bacillus sp. isolated from Tunisian hypersaline (Fdhila et al., 2017). The significant improvement in growth performance within these findings may be due to the ability of Bacillus spp. to adhere to the intestinal mucosa of the host, allowing them to secrete a wide range of digestive enzymes; such as amylase, lipase, and protease; all of which facilitate feed utilization (Marzouk et al., 2008; Latorre et al., 2016). Moreover, Bacillus spp. has been reported to detoxify potentially harmful food components, and produce many essential vitamins in the B-complex group (particular vitamin B₁₂ and biotin), which results in higher feed utilization and digestibility of feed components (Blain Kennedy et al., 1998; El-Haroun et al., 2006). Regarding the immunomodulatory effects of probiotics, it should be noted that adhesion and colonization of probiotics in the intestines of the host are necessary to enhance the immune response (Ausubel, 2005; Govender et al., 2014; Nishiyama et al., 2016; Li et al., 2018).

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Probiotics interact with immune system via microbe associated molecular patterns (MAMPs), consisting of cell wall polysaccharides, peptidoglycans, lipoprotein anchors, and lipoteichoic acids (Hosoi et al., 2003). Cells or components within the immune system are then able to interact with MAMPs through pattern recognition receptor; such as Toll like receptors, C-type receptor, and nucleotide oligomerigation domain like receptors (Kleerebezem et al., 2010; Lebeer et al., 2010; Bron et al., 2012). Another critical feature of probiotics is their presence inside the host, in which they are potentially colonizing and replicating, which further serves to improve the host's health (Pandiyan et al., 2013). The colonization ability of probiotics within the GI tract of animals provides a necessary competition to indigenous microbiota (Hill, 1993). The ability to attach the gut surface is an important aspect of probiotics, which in turn is associated with mucous formation of the bacteria. These abilities are useful for protection against pathogen (Olsson et al., 1992), or immune stimulation (Salminen et al., 2007). Other species within genus Bacillus; Bacillus OJ, Bacillus amyloliquifaciens, and Bacillus aerophilus, isolated from GI tracts of Pacific white shrimp (Litopenaeus vannamei), yellow fin bream (Acanthopagrus latus), and rohu significantly enhanced growth performance, immune response, and disease resistance of Pacific white shrimp (Li et al., 2009), Nile tilapia (Oreochromis niloticus) (Ridha & Azad, 2012), and rohu (Ramesh et al., 2017). More recently, Meidong et al. (2018) reported that B. aerius isolated from pla-mong (Pangasius bocourti) improved growth performance, immune responses, as well as resistance against A. hydrophila. **6.2** *Bacillus subtilis* B. subtilis is one of the most studied species within genus Bacillus. Dietary administration of B. subtilis isolated from mrigal (Cirrhinus mrigala) significantly improved total serum protein, globulin content, antibody level, hematology and immune response, as well as disease resistance against E. tarda and A. hydrophila in rohu (Nayak et al., 2007; Kumar et al., 2008). In a study with grass carp (Ctenopharyngodon idella), Wu et al. (2012) revealed that dietary

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administration of B. subtilis isolated from grass carp significantly improved specific growth rate, feed conversion ratio, enzyme activity, and total bacteria counts in the GI tract of the fish. In a study using rainbow trout, reported Newaj-Fyzul et al. (2007) that dietary supplementation of B. subtilis isolated from rainbow trout intestines significantly enhanced respiratory burst, serum and gut lysozyme, peroxidase, phagocytic killing, and all α1-antiprotease activities. Furthermore, the study revealed a significant increase in lymphocyte population, as well as the resistance towards A. hydrophila. Likewise, the supplementation of B. subtilis isolated from intestines of sea cucumber, Japanese eel (Anguilla japonica), and pond water significantly enhanced the growth performance, immune response, enzyme activity, and disease resistance of sea cucumber, Japanese eel and Nile tilapia (Zhao et al., 2012; Lee et al., 2017; Liu et al., 2017). In Pacific white shrimp dietary inclusion of B. subtilis significantly increased survival rate, immune response, and resistance to V. harveyi (Liu et al., 2014). However, body crude lipid content significantly decreased in the shrimp fed B. subtilis, vs. the control diet. More recently, a Bacillus spp. revealing exoenzymes activities improved the host's growth performance, including weight gain and feed efficiency (Liu et al., 2017). Bacillus exoenzymes are very efficient at metabolizing a large variety of carbohydrates, lipids, and proteins (Liu et al., 2009), and dietary supplementation of bacilli may improve digestive enzyme activities (Han et al., 2015). It is widely accepted, that the level of digestive enzyme activity is a useful comparative indicator of the host's food utilization rate, digestive capacity, and growth performance (Ueberschär, 1995; Suzer et al., 2008). Regarding immunomodulation and disease improvement, several mechanisms have been suggested, antimicrobial synthesis, pathogen competition, prevention of intestinal inflammation, and improvement of the GI tract microbiota (Suva et al., 2016). B. subtilis spores trigger specific humoral and cell-mediated immune responses (Amuguni & Tzipori, 2012). The interaction between B. subtilis spores and macrophages plays an important role in the

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development of both innate and adaptive immune responses of the host (Guo et al., 2016). Two studies have demonstrated that the inclusion of *B. subtilis* leads to successful macrophage activation. Suva et al. (2016) revealed that *B. subtilis* B10, *B. subtilis* BS02, and *B. subtilis* (natto) B4 spores might possess immunomodulatory activities, obtained through the induction of pro-inflammatory cytokines, which exert probiotic activities through activated macrophage functions. Commensal bacteria play an important role in the development of the gut-associated lymphoid tissue (GALT), and are important for both innate and adaptive immunity. *B. subtilis* promotes active lymphocyte proliferation within GI tract, due to their high metabolic activity. *Bacillus* activity is determined mainly by their ability to produce antibiotics; of which, *B. subtilis* is the most productive species. *Bacillus subtilis* devotes 4%–5% of its genome to antibiotic synthesis, and produces as many as 66 antibiotics. Each *Bacillus* antibiotic contains a different structure and spectrum of antimicrobial activity (Sorokulova, 2013).

6.3 Bacillus cereus

Few studies have used *B. cereus* as potential probiotic in fish and shrimp. Navin et al. (2014) indicated that the giant tiger prawn (*Penaeus monodon*) fed dietary inclusion of *B. cereus* isolated from shrimp's intestines; significantly increase growth performance, feed conversion ratio, and immune response. Rengpipat et al. (2000) reported a similar improvement in immune response in shrimp, which they attributed to the presence of the surface antigens of *Bacillus* S11 and their metabolites, which serve as immunogens in the shrimp's immune defense mechanisms. They also noted that the additional of *Bacillus* S11 cell wall peptidoglycan may trigger immune functions in shrimp. In a study using sea cucumber, Zhao et al. (2016) observed significant increases in phagocytosis, respiratory burst, and total nitric oxide synthase activities; as well as disease resistance to *Vibrio splendidus*. The dietary *B. cereus* isolated from the mud of sea cucumber culturing water bodies, while effective in the above ways, showed no changes in growth performance, total coelomocytes counts, acid phosphatase, or superoxide dismutase

activities. This difference could be attributed to the specific *Bacillus* strains, the size and source of sea cucumbers, as well as experimental period and conditions (Zhao et al., 2016).

6.4 Bacillus pumilus

Bacillus pumilus is a host-associated probiotic of recently interest in the field of aquaculture. Dietary administration of *B. pumilus* or *B. pumilus* in combination with *B. clausii* isolated from fish hosts significant improved growth performance, immune response, and disease resistance of Nile tilapia and orange-spotted grouper (Sun et al., 2010; Srisapoome & Areechon, 2017). They concluded that the significant improvement in growth performance via *B. pumilus* diet was attributed to the absence of an antigenic stimulus provided by the pathogenic bacteria could reduce a number of immune cells. This condition may create the efficient absorption and utilization of nutrients in the intestines (Sun et al., 2010). Significant enhancement of fish health and disease resistance may be credited to the increase in none-specific immunity by enhancing phagocytic activity, the number of vital leukocytes, and superoxide anion production. In theory, *B. pumilus* provides high anti-spore IgG titers, pro-inflammatory tumor necrosis factor, and a bacteriocin-like activity against other *Bacillus* (Duc et al., 2004); as well as complement activity (Sun et al., 2010). These abilities may effectively elevate the survival rate of experimental fish fed *B. pumilus* (Aly et al., 2008).

6.5 Bacillus licheniformis

B. licheniformis is another efficient host-associated probiotic in aquaculture. Grass carp fed dietary B. licheniformis and B. subtilis isolated from the grass carp pond, significantly increased globulin level, IgM, lysozyme, and complement activities; as well as myeloperoxidase content, superoxide anion production, total antioxidant activity, glutathione, and anti-superoxide anion free radical (Weifen et al., 2012). The improvement in fish immunity may be due to supplementation of Bacillus, which improves water quality, resulting in better living condition for fish (Lalloo et al., 2007; Borges et al., 2008; Wang et al., 2008). The addition of

probiotic bacteria to the water environment can decrease harmful microorganisms, act as a food source for zooplanktons, and stimulate remineralization of macronutrients and micronutrients that boost phytoplankton growth, and thereby enhance the oxygen level (Verschuere et al., 2000). Bacillus species reduces the abundance of pathogenic Vibrio bacteria in in vitro test and improves water quality through the reduction of toxic NH₄⁺ and NO₂-, maintaining a neutral pH (Nimrat et al., 2012). Additionally, Bacillus produce antimicrobial compounds (Rengpipat et al., 1998; Ringø et al., 2007; Nayak, 2010; Nandi et al., 2018). Bacillus preparations, added to the water, 108 CFU m³, increased serum immunoglobulin levels, non-specific immune parameters and the antioxidant ability of grass carp, but by adding *Bacillus* preparations into the water and feed, 10⁸ CFU g⁻¹, significantly improved immune effects were revealed (Weifen et al., 2012). Adding probiotics in both water and feed may provide better opportunities for the colonization of probiotics in the fish intestine, thereby enabling them to compete with pathogens for nutrients, and/or to inhibit the growth of pathogens; thus altering the prophylactic use of chemicals, antibiotics, and biocides (Decamp et al., 2008; Ma et al., 2009). The approach by adding probiotics to water and feed is better to use in saltwater as the environment is saltier, the fish loses water passively, and the fish has to drink. The noticeable increase in antioxidant enzyme activity may be due to that Bacillus act as antigen, which can stimulate the body's antioxidant enzyme system and increase the amount of antioxidants (Weifen et al., 2012; Wang et al., 2017). These antioxidant enzymes effectively remove excessive free radicals produced by a high metabolism and adverse environmental stress; as well as regulate of the body's free radical balance, and repair damages to tissues and organs (e.g. Harris, 1992; Gill et al., 2010). Recently, Gobi et al. (2018) displayed that dietary administration of B. licheniformis significantly improved growth performance, mucus and serum immune parameters, antioxidant enzyme activity, and resistance to A. hydrophila in tilapia. It is known that increase in fish body weight gain by

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feeding fish probiotic supplemented diets, could be contributed to the increase in digestive enzyme activity, increase in appetite, increase in the production of vitamin, breakdown of indigestible components, as well as possible improvement of intestinal morphology (Irianto & Austin, 2002).

There have been numerous investigations on the immunomodulatory effect of probiotics on different immune cells to enhance immune responses (e.g. Nayak et al., 2010; Das et al., 2013; Sangma & Kamilya, 2015; Gobi et al., 2018). Mucus and serum protein were significantly higher in rohu fed a *B. subtilis* supplemented diet, catla (*Catla catla*) fed a diet added *B. amyloliquifaciens*, and catla fed *B. subtilis* (Nayak et al., 2010; Das et al., 2013; Sangma & Kamilya, 2015). Similarly, Gobi et al. (2018) revealed that serum protein and mucus protein were significantly higher in tilapia (*Oreochromis mossambicus*) fed *B. licheniformis* Dahb1 supplemented diets. Serum protein and mucus protein associated with agglutinins, lectins, lysozyme, immunoglobulins are considered as important proteins involved in the defense mechanism (Alexander & Ingram, 1992; Esteban, 2012). Gobi et al. (2018) suggested that tilapia fed diets containing *B. licheniformis* Dahb1 or other probiotics increased the level of

6.6 Lactic acid bacteria

The host-associated lactobacilli have gained much attention in aquaculture, for review see the comprehensive review of Ringø et al. (2018). Harikrishnan et al. (2010) reported that kelp grouper (*Epinephelus bruneus*) fed dietary *Lactobacillus sakei* isolated from olive flounder intestine significantly increased macrophage phagocytic, peroxidase, serum lysozyme activities, and total protein levels; as well as a resistance to *S. iniae* and *S. parauberis*. In addition, Pacific red snapper (*Lutjanus peru*) fed *Lb. sakei* isolated from the mucus of spotted sand bass (*Paralabrax maculatofaciatus*) significantly promoted haemoglobin, myeloperoxidase, lysozyme, and total antiproteases activities; as well as IgM, serum

these proteins, and subsequently increased serum and mucus protein concentration.

antioxidant capabilities, and vacuoles inside the enterocytes (Reyes-Becerril et al., 2014). Enhanced fish growth following feeding of Lb. sakei may be due to the improvement of nutrient digestibility and/or to the alteration of the intestinal microbiota and immunostimulation (Reyes-Becerril et al., 2012). The chemical composition of microorganisms includes different polysaccharides that may have potential prebiotics in the host's intestine (Ringø et al., 2010b). Both bifidobacteria and lactobacilli are capable of utilizing these oligosaccharides, and increase the number of beneficial bacteria in host's gut. Further benefits of dietary supplemented oligosaccharides; include improved feed efficiency, enhanced growth, and increased digestive enzyme activities. Furthermore, they seem able to modulate immune responses and preserve the integrity of the intestinal wall (Xu et al., 2009; Dimitroglou et al., 2010). Lactobacillus plantarum has also gained popularity in fish and shellfish farming. Kongnum & Hongpattarakere (2012) revealed that Lactobacillus plantarum isolated from the shrimp intestines significantly improved relative growth rate, feed conversion ratio, survival rate, and hemocytes count as well as disease resistance to V. harveyi in Pacific white shrimp. Similarly, significant increases in growth performance, feed utilization efficiency, survival rate, enzyme activity, immune response, and disease resistance to A. hydrophila were observed in rohu (Giri et al., 2013); and swimming crab (Portunus pelagicus) (Talpur et al., 2013). More recently, the dietary supplementation of Lb. plantarum, Weissella confuse, Lactococcus lactic, and Enterococcus faecalis were investigated (Li et al., 2018b). The bacteria were isolated from the intestines of flathead grey mullet (Mugil cephalus), rockfish (Sebastes schlegeli), fat greenling (Hexagrammos otakii), and söhachi (Cleisthenes herzensteini) and significantly stimulated growth performance, disease resistance against V. splendidus, alkaline phosphatase, acid phosphatase, super oxide dismutase, lysozyme activity, and up-regulated heat shock proteins genes of juvenile sea cucumber. The elevated of growth rate of aquatic animals may be related to the effects of LAB actions on competitive exclusion of pathogenic bacteria, supplying

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hydrolytic enzymes, and/or by supplying fatty acid and vitamins (Huang et al., 2014; Hai, 2015b). Regarding the immunostimulatory effects of Lb. plantarum, it was reported that lactobacilli could elicit innate and adaptive immune responses in the host via binding to the pattern recognition receptors (PRR) expressed on immune cells and many other tissues, including the intestinal epithelium. PRR recognize the conserved molecular structures known as microbe-associated molecular patterns, which induce the production of cytokines, chemokines, and other innate effectors (Abreu, 2010; Kawai & Akira, 2010; Wells et al., 2010). In addition to lactobacilli, are several other genera of LAB used as probiotics in aquaculture, Enterococcus, Lactococcus, and Weissella. Enterococcus faecium isolated from the intestine of flathead grey mullet significantly boosted nitroblue tetrazolium assay and common carp resistance to A. hydrophila (Gopalakannan & Arul, 2011). The authors suggested that the reason of the noticed results might be the extracellular products of probiotic bacteria, such as bacteriocin- and nisin-like compounds, which induce the immune system of common carp. Additionally, there are several mechanisms through which probiotic bacteria can induce bacterial antagonism, by producing antimicrobial agents, like antibiotics, antimicrobial peptides (bacteriocins), siderophore substances, or hydrogen peroxide (e.g. Sugita et al., 1998; Ringø et al., 2018). Like many probiotics, E. faecium produce bacteriocin, which creates a pore in the cell membrane of pathogens, which leads to the efflux of K⁺ ions, resulting in cell death (Satish & Arul, 2009). In a study with grouper, Sun et al. (2012) reported that dietary supplementation of E. faecium and Lactococcus lactic, originating from the gut of orangespotted grouper, significantly increased feed utilization, hepatopancreatic protease activities, complement component-3, and serum lysozyme activities; however, no change in phagocytic, serum superoxide dismutase activities, IgM, or complement component-4 were observed. Similarly, significant improvements in growth performance, innate immune response, upregulation of lysozyme genes, and disease resistance to Streptococcosis were observed in

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kurumar shrimp (*Marsupenaeus japonicus*) and olive flounder, fed host-associated *Lactococcus* lactic as a probiotic (Maeda et al., 2014; Nguyen et al., 2017). Allameh et al. (2014) isolated an *E. faecalis* from intestine of snakehead fish (*Channa striatus*) and revealed significantly improved resistance of snakehead fish against *A. hydrophila* when the *E. faecalis* was included in the diet. More recently, Mouriño et al. (2016) reported that hybrid surubim catfish (*Pseudoplatystoma reticulatum* $\mathcal{P} \times P$. *corruscans* \mathcal{P}), fed *Weissella cibaria* isolated from its foregut intestine, showed a significant increased red blood cell count. However, no significant difference occurred in white blood cells, total thrombocyte count, haematocrit percentage, basophils and other leukocytes; as well as glucose levels between supplemented diets and control.

6.7 Other host-associated probiotics

Aeromonas sobria has been used as a potential probiotic in aquaculture (Brunt & Austin, 2005). The authors suggested that dietary administration of *A. sobria* isolated from GI tract of rainbow trout significantly enhanced appetite, leukocytes count, phagocytic and respiratory burst activities; as well as resistance to *Lactococcus garvieae* and *S. iniae*. The mode of action of *A. sobria* involves the stimulation of cellular immunity, which directly increases the number of leucocytes, and enhances phagocytic and respiratory burst activities. It remains speculative, exactly how *A. sobria* stimulates this activity when used as a feed supplement (Brunt & Austin, 2005).

Senegalese sole (*Solea senegalensis*) fed diet supplemented with two *Shewanella*, Pdp11 and Pdp13 isolated from gilthead sea bream skin, significantly increased growth and nutrient utilization, proximate composition, and alkaline phosphatase of juvenile sole (Sáenz et al., 2009). Sharifuzzaman & Austin (2009) conducted an experiment with the use of *Kocuria* SM1 solated from the digestive tract of rainbow trout. Fish fed dietary *Kocuria* SM1 at concentration of 10⁸ cells g⁻¹ for four weeks showed significantly enhanced phagocytic,

peroxidase, and serum lysozyme activities. In a study with rainbow trout, LaPatra et al. (2014) revealed that feeding host-associated *Enterobacter* sp. significantly increased survival rate, antibody titers, and resistance to Flavobacterium psychrophilum. Similarly, significant increase were noticed on growth performance, tolerance to crowding stress, microbiota, enzyme activities, innate immune response, relative immune gene expressions, and disease resistance shellfish fed Shewanella putrefaciens, Psychrobacter namhaensis, Pseudoalteromonas, and Clostridium butyricum host- associated probiotics (Tapia-Paniagua et al., 2014; Makled et al., 2017; Offret et al., 2018; Sumon et al., 2018). The significant increase in growth performance and health status of fish, as well as disease resistance within above studies are possibly due to the increase in digestive enzyme activities, absorptive capacities of the intestine, innate immune, relative immune gene expressions, and modulate GI tract microbiota. In the pioneer review devoted to "Probiotics in man and animals", Fuller (1989) wrote, "Probiotic preparations may consist of single strains or may contain any number up to eight strains". However, most probiotic studies carried out in aquaculture since the early 1990ties and until today have used single administration, but during the last decade, supplementation of multiple probiotics in the diets to aquatic animals has gained interest (e.g. Mohapatra et al., 2011; Ibrahem, 2015; Zorriehzahra et al., 2016). The advantage of multiple-strain preparations is; they are active against wider range of conditions and species. Dietary inclusion of Enterobacter cloacae and Bacillus mojavensis, singular or combined, significantly improved growth performance, cellular immunity, and disease resistance of rainbow trout (Capkin & Altinok, 2009). These improvements may be due to the probiotic effects of these two microorganisms. Probiotics stimulate the digestive processes through the proliferation of a favorable microbiota population, enhancing microbial enzyme activity, improving the intestinal microbial balance; and, consequently, improving the digestibility,

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absorption of food, and feed utilization (El-Haroun et al., 2006; Capkin & Altinok, 2009). The authors indicated that the putative probiotic bacteria isolated from intestines of rainbow trout possess antibacterial abilities. Presence of such intestinal bacteria in the GI tract can protect fish against infections caused by pathogenic bacteria (Capkin & Altinok 2009). The mode of action of Enterobacter and B. mojavensis is to stimulate cellular immunity, specifically due to an increased number of leucocytes. E. cloacae are listed under the same family as Y. ruckeri, Enterobacteriaceae. This strain may not only competitively exclude Y. ruckeri, but also present an appearance likely to trigger higher immune reactions than that of an unrelated Bacillus (Capkin & Altinok, 2009). Similarly, the combination of three probiotics; *Bacillus coagulans*, Rhodoseudomonas palustris, and Lactobacillus acidophilus isolated from intestines of common carp and grass carp significantly increased weight gain, final weight, and digestive enzyme activity of grass carp (Wang, 2011). Chi et al. (2014) revealed that the dietary supplementation of Aeromonas veronii, Vibrio lentus, and Flavobacterium sasangense derived from GI tract of common carp significantly boosted total serum protein, albumin and globulin levels, innate immune response, and expression of 1L-1b and TNF-α in common carp. Similar results improving growth performance, digestive enzyme activities, stress indicators, immune response, relative immune gene expressions, modulation of gut microbiota composition, and disease resistance were observed in rainbow trout fed Kocuria SM1 and Rhodococcus SM2 (Sharifuzzaman et al., 2014), in grass carp fed Shewanella xiamenensis and A. veronii (Wu et al., 2015), in sea cucumber fed Bacillus cereus, B. cereus, and Paracoccus marcusii (Yang et al., 2015), in Malaysian mahseer (Tor tambroides) fed Bacillus sp. and Alcaligenes sp. or Bacillus sp., Alcaligenes sp. and Shewanella sp. (Asaduzzaman et al., 2018a, b). In a study using Pacific white shrimp, dietary administration of host-associated Shewanella haliotis, B. cereus, and Aeromonas bivalvium significantly stimulated respiratory burst, superoxide dismutase, respiratory burst activity, acid phosphatase activities, and up-regulated

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prophenoloxidase and β-glucan-binding protein genes, as well as resistance to V. harveyi (Hao et al., 2014). In recent studies with Pacific white shrimp fed host-associated probiotics Rhodobacter sphaeroides and Afifella marina or B. subtilis and S. algae isolated from ponds or its GI tract; significantly improvements in water quality, growth performance, resistance to acute hepatopancreatic necrosis disease (AHPND), and control Vibrio in shrimp hepatopancreas were observed (Chumpol et al., 2017; Interaminense et al., 2018). As with other host-associated probiotics, significant improvements in growth performance, immune response, and disease resistance of fish may be due to the increase in digestive enzyme activities, absorptive capacities of the intestine, innate immune and relative immune gene expressions, modulation of the GI tract microbiota, and production of antibacterial substances (Van Doan et al., 2018). Additionally, a recent study reported that growth performance improvement of Malaysian mahseer was governed primary by muscle fibres hypertrophy and up-regulated growth related (GH and IGF1) gene expression (Asaduzzaman et al., 2018b). In contrast, Allameh et al. (2016) revealed that inclusion of individual LAB strains in the diet resulted in significantly improved growth performances of Javanese carps compared to fish fed a diet supplemented a LAB mixture. Dietary inclusion E. faecalis at 10⁷ cfu g⁻¹ revealed higher growth performance compared to other treatments. Additionally, the population levels of LAB in the fish intestine significantly increased, while the levels of Gram-negative bacteria significantly decreased vs. the control.

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7. Conclusions and future applications of probiotic in aquaculture

The present review addressed whether or not host-associated probiotics *vs.* probiotics obtained from other sources and commercial probiotics are more suitable to use in aquaculture. Although some studies have revealed beneficial effects of host-associated probiotics on performance, immune responses and disease resistance of aquatic organisms, the observed effects were species-specific. Therefore, it is necessary to carry out additional studies where both host-

associated probiotics and probiotics obtained from other sources are used in the same study, and to determine optimum inclusion levels for host-associated probiotics. In order, to evaluate in vivo adherence and colonization of the host-associated probiotic strains within the complex microbial ecosystem of the intestine, detection of green fluorescence protein (GFP) tagged strains or fluorescence in situ hybridization (FISH) targeting 16S rRNA to identify the probiotics on the mucus surface must be carried out. Furthermore, mucus associated (autochthonous) microbiome must be investigated by next-generation sequencing (NGS), transcriptomic or proteomic profiling, and not the allochthonous microbiome; mostly investigated per sc. In addition, we recommend that gnotobiotic approaches are used in future studies, as the gnotobiotic approaches have been reported to have important roles to understand the function of gut microbiota on numerous biological processes of the host. Moreover, data is needed to understand the mechanisms by which the immune system of the intestinal mucosa discriminates between pathogenic, probiotics and commensal microorganisms. In summary, the present study revealed higher effectiveness of host-associated probiotics compared probiotics obtained from other sources, but increased attention towards isolation, optimum inclusion level and administration of host-associated probiotics in aquaculture is needed.

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References

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654 Abdelkhalek NKM, Eissa IAM, Ahmed E, Kilany OE, El-Adl M, Dawood MAO, et al., (2017) Protective role of dietary Spirulina platensis against diazinon-induced Oxidative 655 damage in Nile tilapia; Oreochromis niloticus. Environmental Toxicology and 656 *Pharmacology* **54:** 99-104. 657 Abreu MT (2010) Toll-like receptor signaling in the intestinal epithelium: how bacterial 658 recognition shapes intestinal function. *Nature Reviews Immunology* **10:** 131. 659 Adel M, Yeganeh S, Dadar M, Sakai M, Dawood MAO (2016) Effects of dietary Spirulina 660 platensis on growth performance, humoral and mucosal immune responses and disease 661 662 resistance in juvenile great sturgeon (Huso huso Linnaeus, 1754). Fish & Shellfish *Immunology* **56:** 436-444. 663 Ahmed E, Holmström SJM (2014) Siderophores in environmental research: roles and 664 665 applications. *Microbial Biotechnology* **7:** 196-208. Alexander, J.B. & Ingram, G.A. (1992) Noncellular nonspecific defence mechanisms of fish. 666 Annual Review of Fish Diseases 2: 249-279. 667 Allameh SK, Ringø E, Yusoff FM, Daud HM, Ideris A (2014) Properties of Enterococcus 668 faecalis, a probiotic bacterium isolated from intestine of snakehead (Channa striatus). 669 670 *African Journal of Microbiology Research* **8:** 2215-2222. Allameh SK, Yusoff FM, Ringø E, Daud HM, Saad CR, Ideris A (2016) Effects of dietary 671 mono-, and multiprobiotic strains on growth performance, gut bacteria and body 672 composition of Javanese carp (Puntius gonionotus, Bleeker 1850). Aquaculture 673 Nutrition 22: 367-373. 674 Altinok I, Capkin E, Karsi A (2015) Succinate dehydrogenase mutant of Listonella anguillarum 675 protects rainbow trout against vibriosis. *Vaccine* **33:** 5572-5577. 676

| 677 | Altinok I, Ozturk RC, Kahraman UC, Capkin E (2016) Protection of rainbow trout against |
|-----|---|
| 678 | yersiniosis by lpxD mutant Yersinia ruckeri. Fish & Shellfish Immunology 55: 21-27. |
| 679 | Aly SM, Abd-El-Rahman AM, John G, Mohamed MF (2008) Characterization of some bacteria |
| 680 | isolated from Oreochromis niloticus and their potential use as probiotics. Aquaculture |
| 681 | 277: 1-6. |
| 682 | Amuguni H, Tzipori S (2012) Bacillus subtilis: A temperature resistant and needle free delivery |
| 683 | system of immunogens. Human Vaccines & Immunotherapeutics 8: 979-986. |
| 684 | Andrews SC, Robinson AK, Rodriguez-Quinones F (2003) Bacterial iron homeostasis. FEMS |
| 685 | Microbiology Reviews 27: 215-237. |
| 686 | Araújo C, Muñoz-Atienza E, Nahuelquín Y, Poeta P, Igrejas G, Hernández PE, et al., (2015) |
| 687 | Inhibition of fish pathogens by the microbiota from rainbow trout (Oncorhynchus |
| 688 | mykiss, Walbaum) and rearing environment. Anaerobe 32: 7-14. |
| 689 | Ardó L, Yin G, Xu P, Váradi L, Szigeti G, Jeney Z, et al., (2008) Chinese herbs (Astragalus |
| 690 | membranaceus and Lonicera japonica) and boron enhance the non-specific immune |
| 691 | response of Nile tilapia (Oreochromis niloticus) and resistance against Aeromonas |
| 692 | hydrophila. Aquaculture 275 : 26-33. |
| 693 | Asaduzzaman M, Iehata S, Akter S, Kader MA, Ghosh SK, Khan MNA, et al., (2018a) Effects |
| 694 | of host gut-derived probiotic bacteria on gut morphology, microbiota composition and |
| 695 | volatile short chain fatty acids production of Malaysian Mahseer Tor tambroides. |
| 696 | Aquaculture Reports 9: 53-61. |
| 697 | Asaduzzaman M, Sofia E, Shakil A, Haque NF, Khan MNA, Ikeda D, et al., AB (2018b) Host |
| 698 | gut-derived probiotic bacteria promote hypertrophic muscle progression and upregulate |
| 699 | growth-related gene expression of slow-growing Malaysian mahseer Tor tambroides. |
| 700 | Aquaculture Reports 9: 37-45. |

Askarian F, Kousha A, Salma W, Ringø E (2011) The effect of lactic acid bacteria 701 administration on growth, digestive enzyme activity and gut microbiota in Persian 702 sturgeon (Acipenser persicus) and beluga (Huso huso) fry. Aquaculture Nutrition 17: 703 704 488-497. Assefa A, Abunna F (2018) Maintenance of fish health in aquaculture: Review of 705 epidemiological approaches for prevention and control of infectious disease of fish. 706 707 Veterinary Medicine International **2018**: 10. Ausubel FM (2005) Are innate immune signaling pathways in plants and animals conserved? 708 Nature Immunology **6**: 973. 709 710 Balcázar JL, Blas Id, Ruiz-Zarzuela I, Cunningham D, Vendrell D, Múzquiz JL (2006) The role of probiotics in aquaculture. Veterinary Microbiology 114:173-186. 711 Balcazar JL, de Blas I, Ruiz-Zarzuela I, Vendrell D, Girones O, Muzquiz JL (2007) 712 713 Enhancement of the immune response and protection induced by probiotic lactic acid bacteria against furunculosis in rainbow trout (Oncorhynchus mykiss). FEMS 714 715 immunology and medical microbiology **51**: 185-193. Barman D, Nen P, Mandal SC, Kumar V (2013) Immunostimulants for Aquaculture Health 716 Management. Journal of Marine Scicence: Research and Development 3: 11. 717 Blain Kennedy S, Tucker Jr JW, Neidig CL, Vermeer GK, Cooper VR, Jarrell JL, et al., (1998) 718 Bacterial management strategies for stock enhancement of warmwater marine fish: a 719 case study with common snook (Centropomus undecimalis). Bulletin of Marine Science 720 721 **62**: 573-588. Bolasina S, Pérez A, Yamashita Y (2006) Digestive enzymes activity during ontogenetic 722 development and effect of starvation in Japanese flounder, Paralichthys olivaceus. 723

Aquaculture **252**: 503-515.

Borges M-T, Sousa A, De Marco P, Matos A, Hönigová P, Castro PML (2008) Aerobic and 725 726 Anoxic Growth and Nitrate Removal Capacity of a Marine Denitrifying Bacterium Isolated from a Recirculation Aquaculture System. *Microbial Ecology* **55**: 107-118. 727 Bostock J, McAndrew B, Richards R, Jauncey K, Telfer T, Lorenzen K, et al., (2010) 728 Aquaculture: global status and trends. Philosophical Transactions of the Royal Society 729 *B: Biological Sciences* **365**: 2897-2912. 730 731 Boyd CE, Hollerman WD, Plumb JA, Saeed M (1984) Effect of treatment with a commercial bacterial suspension on water quality in channel catfish ponds. Progressive Fish 732 Culturist 46: 37-40. 733 734 Bricknell I, Dalmo RA (2005) The use of immunostimulants in fish larval aquaculture. Fish & Shellfish Immunology 19: 457-472. 735 Bron PA, van Baarlen P, Kleerebezem M (2012) Emerging molecular insights into the 736 737 interaction between probiotics and the host intestinal mucosa. Nature Reviews *Microbiology* **10**: 66-78. 738 739 Brudeseth BE, Wiulsrød R, Fredriksen BN, Lindmo K, Løkling K-E, Bordevik M, et al., K (2013) Status and future perspectives of vaccines for industrialised fin-fish farming. 740 Fish & Shellfish Immunology 35: 1759-1768. 741 742 Brunt J, Austin B (2005) Use of a probiotic to control lactococcosis and streptococcosis in rainbow trout, Oncorhynchus mykiss (Walbaum). Journal of Fish Diseases 28: 693-701. 743 Buntin N, de Vos WM, Hongpattarakere T (2017) Variation of mucin adhesion, cell surface 744 characteristics, and molecular mechanisms among Lactobacillus plantarum isolated 745 from different habitats. *Applied Microbiology and Biotechnology* **101:** 7663-7674. 746 Caipang CMA, Brinchmann MF, Kiron V (2010) Antagonistic activity of bacterial isolates from 747 intestinal microbiota of Atlantic cod, Gadus morhua, and an investigation of their 748 immunomodulatory capabilities. Aquaculture Research 41: 249-256. 749

- 750 Capkin E, Altinok I (2009) Effects of dietary probiotic supplementations on
- prevention/treatment of yersiniosis disease. Journal of Applied Microbiology 106:
- 752 1147-1153.
- 753 Capkin E, Ozcelep T, Kayis S, Altinok I (2017) Antimicrobial agents, triclosan, chloroxylenol,
- methylisothiazolinone and borax, used in cleaning had genotoxic and histopathologic
- effects on rainbow trout. *Chemosphere* **182**: 720-729.
- 756 Capkin E, Ozdemir S, Ozturk RC, Altinok I (2017) Determination and transferability of
- 757 plasmid-mediated antibiotic resistance genes of the bacteria isolated from rainbow trout.
- 758 *Aquaculture Research* **48**: 5561-5575.
- 759 Capkin E, Terzi E, Altinok I (2015) Occurrence of antibiotic resistance genes in culturable
- bacteria isolated from Turkish trout farms and their local aquatic environment. *Diseases*
- 761 *of Aquatic Organisms* **114**: 127-137.
- 762 Carnevali, O., Maradonna, F. & Gioacchini, G. (2017) Integrated control of fish metabolism,
- wellbeing and reproduction: The role of probiotic. *Aquaculture* **472**: 144-155.
- Caza M, Kronstad JW (2013) Shared and distinct mechanisms of iron acquisition by bacterial
- and fungal pathogens of humans. Frontiers in Cellular and Infection Microbiology 3:
- 766 80.
- 767 Cha J-H, Rahimnejad S, Yang S-Y, Kim K-W, Lee K-J (2013) Evaluations of *Bacillus* spp. as
- dietary additives on growth performance, innate immunity and disease resistance of
- olive flounder (Paralichthys olivaceus) against Streptococcus iniae and as water
- 770 additives. *Aquaculture* **402-403**: 50-57.
- 771 Cheng G, Hao H, Xie S, Wang X, Dai M, Huang L, Yuan Z (2014) Antibiotic alternatives: the
- substitution of antibiotics in animal husbandry? Frontiers in Microbiology 5: 217.
- 773 Chi C, Jiang B, Yu X-B, Liu T-Q, Xia L, Wang G-X (2014) Effects of three strains of intestinal
- autochthonous bacteria and their extracellular products on the immune response and

- disease resistance of common carp (*Cyprinus carpio*). Fish & Shellfish Immunology **36**:
- 776 9-18.
- 777 Chumpol S, Kantachote D, Nitoda T, Kanzaki H (2017) The roles of probiotic purple nonsulfur
- bacteria to control water quality and prevent acute hepatopancreatic necrosis disease
- 779 (AHPND) for enhancement growth with higher survival in white shrimp (*Litopenaeus*
- 780 *vannamei*) during cultivation. *Aquaculture* **473**: 327-336.
- 781 Collado MC, Gueimonde M, Salminen S (2010) Chapter 23 Probiotics in adhesion of
- pathogens: Mechanisms of action A2 Watson, Ronald Ross. In: Bioactive Foods in
- 783 Promoting Health (ed by Preedy VR). Academic Press, Boston, pp. 353-370.
- 784 Collins FM, Carter PB (1978) Growth of Salmonellae in orally infected germfree mice.
- 785 *Infection and Immunity* **21:** 41-47.
- 786 Cornelissen CN (2003) Transferrin-iron uptake by Gram-negative bacteria. Frontiers in
- 787 *Bioscience* **8**: d836-847.
- Das, A., Nakhro, K., Chowdhury, S. & Kamilya, D. (2013) Effects of potential probiotic
- 789 Bacillus amyloliquifaciens FPTB16 on systemic and cutaneous mucosal immune
- responses and disease resistance of catla (*Catla catla*). Fish & Shellfish Immunology **35**:
- 791 1547-1553.
- Dawood MAO, Koshio S (2016) Recent advances in the role of probiotics and prebiotics in
- carp aquaculture: A review. *Aquaculture* **454**: 243-251.
- 794 Dawood MAO, Koshio S, Ishikawa M, El-Sabagh M, Esteban MA, Zaineldin AI (2016)
- Probiotics as an environment-friendly approach to enhance red sea bream, *Pagrus major*
- growth, immune response and oxidative status. Fish & Shellfish Immunology 57: 170-
- 797 178.
- A.O., D.M., Shunsuke, K., M., A.-D.M. & Hien, V.D. Probiotic application for sustainable
- aquaculture. *Reviews in Aquaculture* (In press): https://doi.org/10.1111/raq.12272

| 800 | De BC, Meena DK, Behera BK, Das P, Das Mohapatra PK, Sharma AP (2014) Probiotics in |
|-----|--|
| 801 | fish and shellfish culture: immunomodulatory and ecophysiological responses. Fish |
| 802 | Physiology and Biochemistry 40: 921-971. |
| 803 | Decamp O, Moriarty DJW, Lavens P (2008) Probiotics for shrimp larviculture: review of field |
| 804 | data from Asia and Latin America. Aquaculture Research 39: 334-338. |
| 805 | Defoirdt T, Sorgeloos P, Bossier P (2011) Alternatives to antibiotics for the control of bacterial |
| 806 | disease in aquaculture. Current Opinion in Microbiology 14: 251-258. |
| 807 | Derrien M, van Hylckama Vlieg JET (2015) Fate, activity, and impact of ingested bacteria |
| 808 | within the human gut microbiota. Trends in Microbiology 23: 354-366. |
| 809 | Dhanasiri AKS, Brunvold L, Brinchmann MF, Korsnes K, Bergh Ø, Kiron V (2011) Changes |
| 810 | in the intestinal microbiota of wild Atlantic cod Gadus morhua L. Upon captive rearing. |
| 811 | Microbial Ecology 61: 20-30. |
| 812 | Dhar AK, Manna SK, Thomas Allnutt FC (2014) Viral vaccines for farmed finfish. Virus |
| 813 | Disease 25 : 1-17. |
| 814 | Dimitroglou A, Merrifield DL, Spring P, Sweetman J, Moate R, Davies SJ (2010) Effects of |
| 815 | mannan oligosaccharide (MOS) supplementation on growth performance, feed |
| 816 | utilisation, intestinal histology and gut microbiota of gilthead sea bream (Sparus |
| 817 | aurata). Aquaculture 300 : 182-188. |
| 818 | Duc LH, Hong HA, Barbosa TM, Henriques AO, Cutting SM (2004) Characterization of |
| 819 | Bacillus probiotics available for human Use. Applied and Environmental Microbiology |
| 820 | 70 : 2161-2171. |
| 821 | El-Haroun E, Goda AS, Chowdhury K (2006) Effect of dietary probiotic Biogen® |
| 822 | supplementation as a growth promoter on growth performance and feed utilization of |
| 823 | Nile tilapia Oreochromis niloticus (L.). Aquaculture Research 37: 1473-1480. |

| 824 | Ellermann M, Arthur JC (2017) Siderophore-mediated iron acquisition and modulation of host- |
|-----|---|
| 825 | bacterial interactions. Free Radical Biology and Medicine 105: 68-78. |
| 826 | Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H (2017) Bacillus as potential |
| 827 | probiotics: Status, concerns, and future perspectives. Frontiers in Microbiology 8. |
| 828 | Esteban, M.A. (2012) An Overview of the Immunological Defenses in Fish Skin. ISRN |
| 829 | Immunology 2012: 29. |
| 830 | Fdhila K, Haddaji N, Chakroun I, Dhiaf A, Macherki MEE, Khouildi B, et al., (2017) Culture |
| 831 | conditions improvement of Crassostrea gigas using a potential probiotic Bacillus sp |
| 832 | strain. Microbial Pathogenesis 110: 654-658. |
| 833 | Fečkaninová A, Koščová J, Mudroňová D, Popelka P, Toropilová J (2017) The use of probiotic |
| 834 | bacteria against Aeromonas infections in salmonid aquaculture. Aquaculture 469: 1-8. |
| 835 | Feodorova VA, Sayapina LV, Corbel MJ, Motin VL (2014) Russian vaccines against especially |
| 836 | dangerous bacterial pathogens. Emerging Microbes & Infections 3: e86. |
| 837 | Fischbach MA, Segre JA (2016) Signaling in host-associated microbial communities. Cell |
| 838 | 164 :1288-1300. |
| 839 | Fjellheim AJ, Playfoot KJ, Skjermo J, Vadstein O (2007) Vibrionaceae dominates the |
| 840 | microflora antagonistic towards Listonella anguillarum in the intestine of cultured |
| 841 | Atlantic cod (Gadus morhua L.) larvae. Aquaculture 269: 98-106. |
| 842 | Founou LL, Founou RC, Essack SY (2016) Antibiotic resistance in the food chain: A |
| 843 | developing country-perspective. Frontiers in Microbiology 7: 1881. |
| 844 | Fuller R (1989) Probiotics in man and animals. <i>Journal of Applied Bacteriology</i> 66: 365-378. |
| 845 | Gatesoupe FJ (1999) The use of probiotics in aquaculture. Aquaculture 180: 147-165. |
| 846 | Gatesoupe FJ (2007) Live yeasts in the gut: Natural occurrence, dietary introduction, and their |
| 847 | effects on fish health and development. Aquaculture 267: 20-30. |

| 848 | Gill R, Tsung A, Billiar T (2010) Linking oxidative stress to inflammation: Toll-like receptors. |
|-----|--|
| 849 | Free Radical Biology & Medicine 48: 1121-1132. |
| 850 | Giraffa G, Chanishvili N, Widyastuti Y (2010) Importance of lactobacilli in food and feed |
| 851 | biotechnology. Research in Microbiology 161: 480-487. |
| 852 | Giri SS, Sukumaran V, Oviya M (2013) Potential probiotic Lactobacillus plantarum VSG3 |
| 853 | improves the growth, immunity, and disease resistance of tropical freshwater fish, |
| 854 | Labeo rohita. Fish & Shellfish Immunology 34: 660-666. |
| 855 | Gobi N, Vaseeharan B, Chen J-C, Rekha R, Vijayakumar S, Anjugam M, et al., (2018) Dietary |
| 856 | supplementation of probiotic Bacillus licheniformis Dahb1 improves growth |
| 857 | performance, mucus and serum immune parameters, antioxidant enzyme activity as well |
| 858 | as resistance against Aeromonas hydrophila in tilapia Oreochromis mossambicus. Fish |
| 859 | & Shellfish Immunology 74 : 501-508. |
| 860 | Gomez D, Sunyer JO, Salinas I (2013) The mucosal immune system of fish: the evolution of |
| 861 | tolerating commensals while fighting pathogens. Fish & shellfish immunology 35: 1729- |
| 862 | 1739. |
| 863 | Gopalakannan A, Arul V (2011) Inhibitory activity of probiotic Enterococcus faecium MC13 |
| 864 | against Aeromonas hydrophila confers protection against hemorrhagic septicemia in |
| 865 | common carp Cyprinus carpio. Aquaculture International 19: 973-985. |
| 866 | Govender, M., Choonara, Y.E., Kumar, P., du Toit, L.C., van Vuuren, S. & Pillay, V. (2014) A |
| 867 | review of the advancements in probiotic delivery: Conventional vs. non-conventional |
| 868 | formulations for intestinal flora supplementation. AAPS PharmSciTech 15: 29-43. |
| 869 | Grajek K, Sip A, Foksowicz-Flaczyk J, Dobrowolska A, Wita A 2016) Adhesive and |
| 870 | hydrophobic properties of the selected LAB isolated from the gastrointestinal tract of |
| 871 | farming animals. Acta Biochimica Polonica 63: 311-314. |

Guo M, Hao G, Wang B, Li N, Li R, Wei L, et al., (2016) Dietary administration of Bacillus 872 subtilis enhances growth performance, immune response and disease resistance in 873 cherry valley ducks. Frontiers in Microbiology 7: 1975. 874 875 Hai NV (2015a) Research findings from the use of probiotics in tilapia aquaculture: A review. Fish & Shellfish Immunology 45: 592-597. 876 Hai NV (2015b) The use of probiotics in aquaculture. Journal of Applied Microbiology 119: 877 917-935. 878 Han B, Long W-q, He J-y, Liu Y-j, Si Y-q, Tian L-x (2015) Effects of dietary Bacillus 879 licheniformis on growth performance, immunological parameters, intestinal 880 morphology and resistance of juvenile Nile tilapia (Oreochromis niloticus) to challenge 881 infections. Fish & Shellfish Immunology 46: 225-231. 882 Hao K, Liu J-Y, Ling F, Liu X-L, Lu L, Xia L, et al., (2014) Effects of dietary administration 883 884 of Shewanella haliotis D4, Bacillus cereus D7 and Aeromonas bivalvium D15, single or combined, on the growth, innate immunity and disease resistance of shrimp, 885 Litopenaeus vannamei. Aquaculture 428-429: 141-149. 886 Harikrishnan R, Balasundaram C, Heo M-S (2010) Lactobacillus sakei BK19 enriched diet 887 enhances the immunity status and disease resistance to streptococcosis infection in kelp 888 grouper, Epinephelus bruneus. Fish & Shellfish Immunology 29: 1037-1043. 889 Harris ED (1992) Regulation of antioxidant enzymes. FASEB Journal 6: 2675-2683. 890 Hill M (1993) Probiotics: the scientific basis. Gut 34: 863. 891 Hoare R, Ngo TPH, Bartie KL, Adams A (2017) Efficacy of a polyvalent immersion vaccine 892

against Flavobacterium psychrophilum and evaluation of immune response to

vaccination in rainbow trout fry (Oncorhynchus mykiss L.). Veterinary Research 48: 43.

893

| 895 | Hoseinifar SH, Esteban MÁ, Cuesta A, Sun Y-Z (2015) Prebiotics and fish immune response: |
|-----|--|
| 896 | A review of current knowledge and future perspectives. Reviews in Fisheries Science & |
| 897 | Aquaculture 23: 315-328. |
| 898 | Hoseinifar SH, Ringø E, Shenavar Masouleh A, Esteban MA (2016) Probiotic, prebiotic and |
| 899 | synbiotic supplements in sturgeon aquaculture: a review. Reviews in Aquaculture 8: 89- |
| 900 | 102, |
| 901 | Hoseinifar, S.H., Sun, Y.Z. & Caipang, C.M. (2017) Short-chain fatty acids as feed supplements |
| 902 | for sustainable aquaculture: an updated view. Aquaculture Research 48: 1380-1391. |
| 903 | Hosoi T, Hirose R, Saegusa S, Ametani A, Kiuchi K, Kaminogawa S (2003) Cytokine responses |
| 904 | of human intestinal epithelial-like Caco-2 cells to the nonpathogenic bacterium Bacillus |
| 905 | subtilis (natto). International Journal of Food Microbiology 82: 255-264. |
| 906 | Huang J-B, Wu Y-C, Chi S-C (2014) Dietary supplementation of <i>Pediococcus pentosaceus</i> |
| 907 | enhances innate immunity, physiological health and resistance to Vibrio anguillarum in |
| 908 | orange-spotted grouper (Epinephelus coioides). Fish & Shellfish Immunology 39: 196- |
| 909 | 205. |
| 910 | Ibrahem MD (2015) Evolution of probiotics in aquatic world: Potential effects, the current |
| 911 | status in Egypt and recent prospectives. Journal of Advanced Research 6: 765-791. |
| 912 | Interaminense JA, Vogeley JL, Gouveia CK, Portela RWS, Oliveira JP, Andrade HA, et al., |
| 913 | (2018) In vitro and in vivo potential probiotic activity of Bacillus subtilis and |
| 914 | Shewanella algae for use in Litopenaeus vannamei rearing. Aquaculture 488: 114-122. |
| 915 | Irianto A, Austin B (2002) Use of probiotics to control furunculosis in rainbow trout, |
| 916 | Oncorhynchus mykiss (Walbaum). Journal of Fish Diseases 25: 333-342. |
| 917 | Irianto A, Austin B (2003) Use of dead probiotic cells to control furunculosis in rainbow trout, |
| 918 | Oncorhynchus mykiss (Walbaum). Journal of Fish Diseases 26: 59-62. |

| 919 | Jöborn A, Dorsch M, Olsson JC, Westerdahl A, Kjelleberg S (1999) Carnobacterium inhibens |
|-----|--|
| 920 | sp. nov., isolated from the intestine of Atlantic salmon (Salmo salar). International |
| 921 | Journal of Systematic Bacteriology 49 Pt 4: 1891-1898. |
| 922 | Johnson KN, van Hulten MCW, Barnes AC (2008) "Vaccination" of shrimp against viral |
| 923 | pathogens: Phenomenology and underlying mechanisms. Vaccine 26: 4885-4892. |
| 924 | Kammler M, Schön C, Hantke K (1993) Characterization of the ferrous iron uptake system of |
| 925 | Escherichia coli. Journal of Bacteriology 175: 6212-6219. |
| 926 | Kawai T, Akira S (2010) The role of pattern-recognition receptors in innate immunity: update |
| 927 | on Toll-like receptors. <i>Nature Immunology</i> 11 : 373. |
| 928 | Kennedy S, Tucker J, Thomersen M, Sennett D (1998) Current methodology for the use of |
| 929 | probiotic bacteria in the culture of marine fish larvae. Aquaculture'98 Book of |
| 930 | Abstracts. World Aquaculture Society, Las Vegas, USA, 286. |
| 931 | Kleerebezem M, Hols P, Bernard E, Rolain T, Zhou M, Siezen R, et al., (2010) The extracellular |
| 932 | biology of the lactobacilli. FEMS Microbiological Reviews 34: 199 - 230. |
| 933 | Kongnum K, Hongpattarakere T (2012) Effect of Lactobacillus plantarum isolated from |
| 934 | digestive tract of wild shrimp on growth and survival of white shrimp (Litopenaeus |
| 935 | vannamei) challenged with Vibrio harveyi. Fish & Shellfish Immunology 32: 170-177. |
| 936 | Kumar R, Mukherjee SC, Ranjan R, Nayak SK (2008) Enhanced innate immune parameters in |
| 937 | Labeo rohita (Ham.) following oral administration of Bacillus subtilis. Fish & Shellfish |
| 938 | Immunology 24 : 168-172. |
| 939 | Lakshmi B, Viswanath B, Sai Gopal DVR (2013) Probiotics as antiviral agents in shrimp |
| 940 | aquaculture. Journal of Pathogens 2013: 13. |
| 941 | Lalloo R, Ramchuran S, Ramduth D, Görgens J, Gardiner N (2007) Isolation and selection of |
| 942 | Bacillus spp. as potential biological agents for enhancement of water quality in culture |
| 943 | of ornamental fish. Journal of Applied Microbiology 103: 1471-1479. |

| 944 | Langdon A, Crook N, Dantas G (2016) The effects of antibiotics on the microbiome throughout |
|-----|---|
| 945 | development and alternative approaches for therapeutic modulation. Genome Medicine |
| 946 | 8: 39. |
| 947 | LaPatra SE, Fehringer TR, Cain KD (2014) A probiotic Enterobacter sp. provides significant |
| 948 | protection against Flavobacterium psychrophilum in rainbow trout (Oncorhynchus |
| 949 | mykiss) after injection by two different routes. Aquaculture 433: 361-366. |
| 950 | Larsen AM, Mohammed HH, Arias CR (2014) Characterization of the gut microbiota of three |
| 951 | commercially valuable warm water fish species. Journal of Applied Microbiology |
| 952 | 116 :1396-1404. |
| 953 | Latorre JD, Hernandez-Velasco X, Wolfenden RE, Vicente JL, Wolfenden AD, Menconi A, et |
| 954 | al., (2016) Evaluation and selection of Bacillus species based on enzyme production, |
| 955 | antimicrobial activity, and biofilm synthesis as direct-fed microbial candidates for |
| 956 | poultry. Frontiers in Veterinary Science 3: 95. |
| 957 | Lazado CC, Caipang CMA (2014a) Atlantic cod in the dynamic probiotics research in |
| 958 | aquaculture. Aquaculture 424-425: 53-62. |
| 959 | Lazado CC, Caipang CMA (2014b) Mucosal immunity and probiotics in fish. Fish & Shellfish |
| 960 | Immunology 39 : 78-89. |
| 961 | Lazado CC, Caipang CMA, Estante EG (2015) Prospects of host-associated microorganisms in |
| 962 | fish and penaeids as probiotics with immunomodulatory functions. Fish & Shellfish |
| 963 | <i>Immunology</i> 45 : 2-12. |
| 964 | Lazado CC, Caipang CMA, Kiron V (2012) Enzymes from the gut bacteria of Atlantic cod, |
| 965 | Gadus morhua and their influence on intestinal enzyme activity. Aquaculture Nutrition |
| 966 | 18 : 423-431. |

| 967 | Lazado CC, Caipang CMA, Rajan B, Brinchmann MF, Kiron V (2010) Characterization of |
|-----|---|
| 968 | GP21 and GP12: two potential probiotic bacteria isolated from the gastrointestinal tract |
| 969 | of Atlantic cod. Probiotics and Antimicrobial Proteins 2: 126-134. |
| 970 | Lebeer S, Vanderleyden J, De Keersmaecker SCJ (2010) Host interactions of probiotic bacterial |
| 971 | surface molecules: comparison with commensals and pathogens. Nature Reviews |
| 972 | Microbiology 8: 171. |
| 973 | Lee S, Katya K, Park Y, Won S, Seong M, Hamidoghli A, et al., (2017) Comparative evaluation |
| 974 | of dietary probiotics Bacillus subtilis WB60 and Lactobacillus plantarum KCTC3928 |
| 975 | on the growth performance, immunological parameters, gut morphology and disease |
| 976 | resistance in Japanese eel, Anguilla japonica. Fish & Shellfish Immunology 61: 201- |
| 977 | 210. |
| 978 | Li J, Tan B, Mai K (2009) Dietary probiotic <i>Bacillus</i> OJ and isomaltooligosaccharides influence |
| 979 | the intestine microbial populations, immune responses and resistance to white spot |
| 980 | syndrome virus in shrimp (Litopenaeus vannamei). Aquaculture 291: 35-40. |
| 981 | Li X, Ringø E, Hoseinifar SH, Lauzon H, Birkbeck H, Yang D (2018a) Adherence and |
| 982 | colonisation of microorganisms in the fish gastrointestinal tract. Reviews in Aquaculture |
| 983 | [Epub ahead of print], doi: 10.1111/raq.12248. |
| 984 | Li C, Ren Y, Jiang S, Zhou S, Zhao J, Wang R, Li Y (2018b) Effects of dietary supplementation |
| 985 | of four strains of lactic acid bacteria on growth, immune-related response and genes |
| 986 | expression of the juvenile sea cucumber Apostichopus japonicus Selenka. Fish & |
| 987 | Shellfish Immunology 74 : 69-75. |
| 988 | Lilly DM, Stillwell RH (1965) Probiotics: Growth promoting factors produced by |
| 989 | microorganisms. Science 147: 747-748. |

| 990 | Liu CH, Chiu CS, Ho PL, Wang SW (2009) Improvement in the growth performance of white |
|------|---|
| 991 | shrimp, Litopenaeus vannamei, by a protease-producing probiotic, Bacillus subtilis E20, |
| 992 | from natto. Journal of Applied Microbiology 107: 1031-1041. |
| 993 | Liu H, Li Z, Tan B, Lao Y, Duan Z, Sun W, et al., (2014) Isolation of a putative probiotic strain |
| 994 | S12 and its effect on growth performance, non-specific immunity and disease-resistance |
| 995 | of white shrimp, Litopenaeus vannamei. Fish & Shellfish Immunology 41: 300-307. |
| 996 | Liu H, Wang S, Cai Y, Guo X, Cao Z, Zhang Y, et al., (2017) Dietary administration of Bacillus |
| 997 | subtilis HAINUP40 enhances growth, digestive enzyme activities, innate immune |
| 998 | responses and disease resistance of tilapia, Oreochromis niloticus. Fish & Shellfish |
| 999 | Immunology 60 : 326-333. |
| 1000 | Llewellyn MS, Boutin S, Hoseinifar SH, Derome N (2014) Teleost microbiomes: the state of |
| 1001 | the art in their characterization, manipulation and importance in aquaculture and |
| 1002 | fisheries. Frontiers in Microbiology 5. |
| 1003 | Luis Balcázar J, Decamp O, Vendrell D, De Blas I, Ruiz-Zarzuela I (2006) Health and |
| 1004 | nutritional properties of probiotics in fish and shellfish. Microbial Ecology in Health |
| 1005 | and Disease 18 : 65-70. |
| 1006 | Ma C-W, Cho Y-S, Oh K-H (2009) Removal of pathogenic bacteria and nitrogens by |
| 1007 | Lactobacillus spp. JK-8 and JK-11. Aquaculture 287: 266-270. |
| 1008 | Macfarlane GT, Macfarlane S (2012) Bacteria, colonic fermentation, and gastrointestinal |
| 1009 | health. Journal of AOAC International 95: 50-60. |
| 1010 | Maeda M, Shibata A, Biswas G, Korenaga H, Kono T, Itami T, et al., (2014) Isolation of lactic |
| 1011 | acid bacteria from kuruma shrimp (Marsupenaeus japonicus) intestine and assessment |
| 1012 | of immunomodulatory role of a selected strain as probiotic. Marine Biotechnology 16: |
| 1013 | 181-192. |

| L014 | Makled SO, Hamdan AM, El-Sayed A-FM, Hafez EE (2017) Evaluation of marine |
|------|---|
| 1015 | psychrophile, Psychrobacter namhaensis SO89, as a probiotic in Nile tilapia |
| 1016 | (Oreochromis niloticus) diets. Fish & Shellfish Immunology 61: 194-200. |
| 1017 | Mardones FO, Paredes F, Medina M, Tello A, Valdivia V, Ibarra R, et al., (2018) Identification |
| 1018 | of research gaps for highly infectious diseases in aquaculture: The case of the endemic |
| 1019 | Piscirickettsia salmonis in the Chilean salmon farming industry. Aquaculture 482: 211- |
| 1020 | 220. |
| 1021 | Martínez Cruz P, Ibáñez AL, Monroy Hermosillo OA, Ramírez Saad HC (2012) Use of |
| 1022 | probiotics in aquaculture. ISRN microbiology 2012: 916845. |
| 1023 | Marzouk M, Moustafa M, Mohamed N (2008) The influence of some probiotics on the growth |
| 1024 | performance and intestinal microbial flora of O. niloticus. In: Proceeding of the 8th |
| 1025 | International Symposium on Tilapia in Aquaculture, pp. 1059-1071. |
| 1026 | Mataragas M, Metaxopoulos J, Galiotou M, Drosinos EH (2003) Influence of pH and |
| 1027 | temperature on growth and bacteriocin production by Leuconostoc mesenteroides L124 |
| 1028 | and Lactobacillus curvatus L442. Meat Science 64: 265-271. |
| 1029 | Meidong R, Khotchanalekha K, Doolgindachbaporn S, Nagasawa T, Nakao M, Sakai K, et al., |
| 1030 | (2018) Evaluation of probiotic <i>Bacillus aerius</i> B81e isolated from healthy hybrid catfish |
| 1031 | on growth, disease resistance and innate immunity of Pla-mong Pangasius bocourti. |
| 1032 | Fish & Shellfish Immunology 73: 1-10. |
| 1033 | Merrifield DL, Dimitroglou A, Foey A, Davies SJ, Baker RTM, Bøgwald J, et al., (2010) The |
| 1034 | current status and future focus of probiotic and prebiotic applications for salmonids. |
| 1035 | Aquaculture 302 : 1-18. |
| 1036 | Miller JH, Novak JT, Knocke WR, Pruden A (2016) Survival of antibiotic resistant bacteria |
| L037 | and horizontal gene transfer control antibiotic resistance gene content in anaerobic |
| 1038 | digesters Frontiers in Microbiology 7 |

| 1039 | Mohapatra, S., Chakraborty, T., Prusty, A.K., Das, P., Paniprasad, K. & Mohanta, K.N. (2011) |
|------|--|
| 1040 | Use of different microbial probiotics in the diet of rohu, Labeo rohita fingerlings: effects |
| 1041 | on growth, nutrient digestibility and retention, digestive enzyme activities and intestinal |
| 1042 | microflora. Aquaculture Nutrition 18: 1-11. |
| 1043 | Mohapatra S, Chakraborty T, Prusty AK, PaniPrasad K, Mohanta KN (2014) Beneficial effects |
| 1044 | of dietary probiotics mixture on hemato-immunology and cell apoptosis of Labeo rohita |
| 1045 | fingerlings reared at higher water temperatures. PLOS ONE 9: e100929. |
| 1046 | Montalban-Arques A, De Schryver P, Bossier P, Gorkiewicz G, Mulero V, Gatlin DM, et al., |
| 1047 | (2015) Selective manipulation of the gut microbiota improves immune status in |
| 1048 | vertebrates. Frontiers in Immunology 6: 512. |
| 1049 | Morgan-Kiss RM, Priscu JC, Pocock T, Gudynaite-Savitch L, Huner NPA (2006) Adaptation |
| 1050 | and acclimation of photosynthetic microorganisms to permanently cold environments. |
| 1051 | Microbiology and Molecular Biology Reviews: MMBR 70: 222-252. |
| 1052 | Moriarty DJW (1998) Control of luminous Vibrio species in penaeid aquaculture ponds. |
| 1053 | Aquaculture 164 : 351-358. |
| 1054 | Mouriño JLP, Pereira GDV, Vieira FDN, Jatobá AB, Ushizima TT, Silva BCD, et al., Seiffert |
| 1055 | WQ, Jesus GFA, Martins ML (2016) Isolation of probiotic bacteria from the hybrid |
| 1056 | South American catfish Pseudoplatystoma reticulatum×Pseudoplatystoma corruscans |
| 1057 | (Siluriformes: Pimelodidae): A haematological approach. Aquaculture Reports 3: 166- |
| 1058 | 171. |
| 1059 | Murall CL, Abbate JL, Puelma Touzel M, Allen-Vercoe E, Alizon S, Froissart R, et al., (2017) |
| 1060 | Chapter five - invasions of host-associated microbiome networks. In: Advances in |
| 1061 | Ecological Research (ed. by Bohan DA, Dumbrell AJ, Massol F). Academic Press, pp. |
| 1062 | 201-281. |

Nandi, A., Banerjee, G., Dan, S.K., Ghosh, K. & Ray, A.K. (2018) Evaluation of in vivo 1063 1064 probiotic efficiency of Bacillus amyloliquifaciens in Labeo rohita challenged by pathogenic strain of Aeromonas hydrophila MTCC 1739. Probiotics and Antimicrobial 1065 1066 Proteins 10: 391-398. Nawaz, A., Bakhsh javaid, A., Irshad, S., Hoseinifar, S.H. & Xiong, H. (2018) The functionality 1067 1068 of prebiotics as immunostimulant: Evidences from trials on terrestrial and aquatic 1069 animals. Fish & Shellfish Immunology 76: 272-278. Navin Chandran M, Iyapparaj P, Moovendhan S, Ramasubburayan R, Prakash S, Immanuel G, 1070 et al., A (2014) Influence of probiotic bacterium Bacillus cereus isolated from the gut 1071 1072 of wild shrimp *Penaeus monodon* in turn as a potent growth promoter and immune enhancer in P. monodon. Fish & Shellfish Immunology **36**: 38-45. 1073 Nayak SK (2010) Probiotics and immunity: A fish perspective. Fish & Shellfish Immunology 1074 1075 **29**: 2-14. Nayak SK, Swain P, Mukherjee SC (2007) Effect of dietary supplementation of probiotic and 1076 1077 vitamin C on the immune response of Indian major carp, Labeo rohita (Ham.). Fish & Shellfish Immunology 23: 892-896. 1078 1079 Newaj-Fyzul A, Adesiyun AA, Mutani A, Ramsubhag A, Brunt J, Austin B (2007) Bacillus 1080 subtilis AB1 controls Aeromonas infection in rainbow trout (Oncorhynchus mykiss, Walbaum). Journal of Applied Microbiology 103: 1699-1706. 1081 Newaj-Fyzul A, Austin B (2015) Probiotics, immunostimulants, plant products and oral 1082 vaccines, and their role as feed supplements in the control of bacterial fish diseases. 1083 Journal of Fish Diseases 38: 937-955. 1084 1085 Nguyen TL, Park C-I, Kim D-H (2017) Improved growth rate and disease resistance in olive flounder, Paralichthys olivaceus, by probiotic Lactococcus lactis WFLU12 isolated 1086 from wild marine fish. Aquaculture 471: 113-120. 1087

| 1088 | Nikoskelainen S, Salminen S, Bylund G, Ouwehand AC (2001a) Characterization and the |
|------|---|
| 1089 | properties of human- and dairy-derived probiotics for prevention of infectious diseases |
| 1090 | in fish. Applied and Environmental Microbiology 67: 2430-2435. |
| 1091 | Nikoskelainen S, Ouwehand AC, Salminen S, Bylund G (2001b) Protection of rainbow trout |
| 1092 | (Oncorhynchus mykiss) from furunculosis by Lactobacillus rhamnosus. Aquaculture |
| 1093 | 198: 229-236. |
| 1094 | Nimrat S, Boonthai T, Vuthiphandchai V (2011) Effects of probiotic forms, compositions of |
| 1095 | and mode of probiotic administration on rearing of Pacific white shrimp (Litopenaeus |
| 1096 | vannamei) larvae and postlarvae. Animal Feed Science and Technology 169: 244-258. |
| 1097 | Nimrat S, Suksawat S, Boonthai T, Vuthiphandchai V (2012) Potential Bacillus probiotics |
| 1098 | enhance bacterial numbers, water quality and growth during early development of white |
| 1099 | shrimp (Litopenaeus vannamei). Veterinary Microbiology 159: 443-450. |
| 1100 | Nishiyama, K., Sugiyama, M. & Mukai, T. (2016) Adhesion Properties of Lactic Acid Bacteria |
| 1101 | on Intestinal Mucin. Microorganisms 4: 34. |
| 1102 | Offret C, Rochard V, Laguerre H, Mounier J, Huchette S, Brillet B, et al., (2018) Protective |
| 1103 | Efficacy of a Pseudoalteromonas Strain in European Abalone, Haliotis tuberculata, |
| 1104 | Infected with Vibrio harveyi ORM4. Probiotics and Antimicrobial Proteins doi: |
| 1105 | 10.1007/s12602-018-9389-8. [Epub ahead of print] |
| 1106 | Ohashi Y, Ushida K (2009) Health-beneficial effects of probiotics: Its mode of action. Animal |
| 1107 | Science Journal 80 : 361-371. |
| 1108 | Olsson JC, Westerdahl A, Conway PL, Kjelleberg S (1992) Intestinal colonization potential of |
| 1109 | turbot (Scophthalmus maximus) - and dab (Limanda limanda)-associated bacteria with |
| 1110 | inhibitory effects against Vibrio anguillarum. Applied and Environmental Microbiology |
| 1111 | 58 : 551-556. |

| 1112 | Pandiyan P, Balaraman D, Thirunavukkarasu R, George EGJ, Subaramaniyan K, Manikkam S, |
|------|---|
| 1113 | et al., (2013) Probiotics in aquaculture. Drug Invention Today 5: 55-59. |
| 1114 | Papadimitriou K, Zoumpopoulou G, Foligné B, Alexandraki V, Kazou M, Pot B, et al., (2015) |
| 1115 | Discovering probiotic microorganisms: in vitro, in vivo, genetic and omics approaches. |
| 1116 | Frontiers in Microbiology 6. |
| 1117 | Parrow NL, Fleming RE, Minnick MF (2013) Sequestration and scavenging of iron in infection. |
| 1118 | Infection and Immunity 81: 3503-3514. |
| 1119 | Pereira LF, Peixoto MJ, Carvalho P, Sansuwan K, Santos GA, Goncalves JFM, et al., (2018) |
| 1120 | Cross-effects of dietary probiotic supplementation and rearing temperature on growth |
| 1121 | performance, digestive enzyme activities, cumulative mortality and innate immune |
| 1122 | response in seabass (Dicentrarchus labrax). Aquaculture Nutrition 24: 453-460. |
| 1123 | Pérez-Sánchez T, Ruiz-Zarzuela I, de Blas I, Balcázar JL (2014) Probiotics in aquaculture: a |
| 1124 | current assessment. Reviews in Aquaculture 6: 133-146. |
| 1125 | Pokorzynski ND, Thompson CC, Carabeo RA (2017) Ironing out the unconventional |
| 1126 | mechanisms of iron acquisition and gene regulation in chlamydia. Frontiers in Cellular |
| 1127 | and Infection Microbiology 7. |
| 1128 | Ramesh D, Souissi S, Ahamed TS (2017) Effects of the potential probiotics Bacillus aerophilus |
| 1129 | KADR3 in inducing immunity and disease resistance in Labeo rohita. Fish & Shellfish |
| 1130 | Immunology 70 : 408-415. |
| 1131 | Ramesh D, Vinothkanna A, Rai AK, Vignesh VS (2015) Isolation of potential probiotic |
| 1132 | Bacillus spp. and assessment of their subcellular components to induce immune |
| 1133 | responses in Labeo rohita against Aeromonas hydrophila. Fish & Shellfish Immunology |
| 1134 | 45 : 268-276. |
| 1135 | Ramírez C, Romero J (2017) The microbiome of Seriola lalandi of wild and aquaculture origin |
| 1126 | reveals differences in composition and notantial function. Frantiers in Microbiology 8 |

| 1137 | Ran C, Carrias A, Williams MA, Capps N, Dan BCT, Newton JC, et al., (2012) Identification |
|------|---|
| 1138 | of Bacillus strains for biological control of catfish pathogens. PLoS ONE 7: e45793. |
| 1139 | Rengpipat S, Phianphak W, Piyatiratitivorakul S, Menasveta P (1998) Effects of a probiotic |
| 1140 | bacterium on black tiger shrimp Penaeus monodon survival and growth. Aquaculture |
| 1141 | 167 : 301-313. |
| 1142 | Rengpipat S, Rukpratanporn S, Piyatiratitivorakul S, Menasaveta P (2000) Immunity |
| 1143 | enhancement in black tiger shrimp (Penaeus monodon) by a probiont bacterium |
| 1144 | (Bacillus S11). Aquaculture 191 : 271-288. |
| 1145 | Reyes-Becerril M, Angulo C, Estrada N, Murillo Y, Ascencio-Valle F (2014) Dietary |
| 1146 | administration of microalgae alone or supplemented with Lactobacillus sakei affects |
| 1147 | immune response and intestinal morphology of Pacific red snapper (Lutjanus peru). |
| 1148 | Fish & Shellfish Immunology 40: 208-216. |
| 1149 | Reyes-Becerril M, Ascencio-Valle F, Macias ME, Maldonado M, Rojas M, Esteban MÁ (2012) |
| 1150 | Effects of marine silages enriched with Lactobacillus sakei 5-4 on haemato- |
| 1151 | immunological and growth response in Pacific red snapper (Lutjanus peru) exposed to |
| 1152 | Aeromonas veronii. Fish & Shellfish Immunology 33: 984-992. |
| 1153 | Ridha MT, Azad IS (2012) Preliminary evaluation of growth performance and immune |
| 1154 | response of Nile tilapia Oreochromis niloticus supplemented with two putative |
| 1155 | probiotic bacteria. Aquaculture Research 43: 843-852. |
| 1156 | Ringø E, Myklebust R, Mayhew TM, Olsen RE (2007) Bacterial translocation and pathogenesis |
| 1157 | in the digestive tract of larvae and fry. Aquaculture 268: 251-264. |
| 1158 | Ringø E, Løvmo L, Kristiansen M, Salinas I, Myklebust R, Olsen RE et al., (2010a) Lactic acid |
| 1159 | bacteria vs. pathogens in the gastrointestinal tract of fish: A review. Aquaculture |
| 1160 | Research 41: 451-467. |

Ringø E, Olsen RE, Gifstad TØ, Dalmo RA, Amlund H, Hemre GI, et al., (2010b) Prebiotics 1161 1162 in aquaculture: a review. Aquaculture Nutrition 16: 117-136. Ringø E, Olsen RE, Gonzales Vecino JL, Wadsworth S, Song SK (2012) Use of 1163 immunostimulants and nucleotides in aquaculture: A review. Journal of Marine 1164 Science: Research & Development 2:1. 1165 Ringø E, Olsen RE, Jensen I, Romero J, Lauzon HL (2014) Application of vaccines and dietary 1166 1167 supplements in aquaculture: possibilities and challenges. Reviews in Fish Biology and Fisheries **24:** 1005-1032. 1168 Ringø E, Hoseinifar SH, Ghosh K, Van Doan H, Beck BR, Song SK (2018) Lactic acid bacteria 1169 1170 – an update. Frontiers in Microbiology **9:** 1818. Rinkinen M, Mättö J, Salminen S, Westermarck E, Ouwehand AC (2000) In vitro adhesion of 1171 lactic acid bacteria to canine small intestinal mucus. Journal of Animal Physiology and 1172 *Animal Nutrition* **84:** 43-47. 1173 Rinkinen M, Westermarck E, Salminen S, Ouwehand AC (2003) Absence of host specificity 1174 1175 for in vitro adhesion of probiotic lactic acid bacteria to intestinal mucus. Veterinary *Microbiology* **97:** 55-61. 1176 Romero J, Feijoo' CG, Navarrete P (2012) Antibiotics in aquaculture - use, abuse and 1177 alternatives. In: Carvalho E (ed) *Health and Environment in Aquaculture*, pp. 159–198. 1178 In-Tech, ISBN 978-953-51-0497-1. 1179 Sabri M, Léveillé S, Dozois CM (2006) A SitABCD homologue from an avian pathogenic 1180 Escherichia coli strain mediates transport of iron and manganese and resistance to 1181 hydrogen peroxide. *Microbiology* **152**: 745-758. 1182 Sáenz de Rodrigáñez MA, DÍAz-Rosales P, ChabrillÓN M, Smidt H, Arijo S, LeÓN-Rubio 1183 JM, et al., (2009) Effect of dietary administration of probiotics on growth and intestine 1184

| 1185 | functionality of juvenile Senegalese sole (Solea senegalensis, Kaup 1858). Aquaculture |
|------|---|
| 1186 | Nutrition 15 : 177-185. |
| 1187 | Salma W, Zhou Z, Wang W, Askarian F, Kousha A, Ebrahimi MT, et al., (2011) Histological |
| 1188 | and bacteriological changes in intestine of beluga (Huso huso) following ex vivo |
| 1189 | exposure to bacterial strains. Aquaculture 314: 24-33. |
| 1190 | Salminen S, Bouley C, Boutron MC, Cummings JH, Franck A, Gibson GR, et al., (2007) |
| 1191 | Functional food science and gastrointestinal physiology and function. The British |
| 1192 | journal of nutrition 80 : S147-S171. |
| 1193 | Sanchez LM, Wong WR, Riener RM, Schulze CJ, Linington RG (2012) Examining the fish |
| 1194 | microbiome: Vertebrate-derived bacteria as an environmental niche for the discovery of |
| 1195 | unique marine natural products. PLOS ONE 7: e35398. |
| 1196 | Sandy M, Butler A (2009) Microbial iron acquisition: Marine and terrestrial siderophores. |
| 1197 | Chemical reviews 109 : 4580-4595. |
| 1198 | Sangma, T. & Kamilya, D. (2015) <i>In vitro</i> and dietary effects of chitin on cellular and humoral |
| 1199 | immune parameters of Catla, Catla catla (Hamilton). Journal of the World Aquaculture |
| 1200 | Society 46 : 617-623. |
| 1201 | Satish RK, Arul V (2009) Purification and characterization of phocaecin PI80: an anti-listerial |
| 1202 | bacteriocin produced by Streptococcus phocae PI80 Isolated from the gut of Peneaus |
| 1203 | indicus (Indian white shrimp). Journal of microbiology and biotechnology 19: 1393- |
| 1204 | 1400. |
| 1205 | Schrezenmeir J, de Vrese M (2001) Probiotics, prebiotics, and synbiotics—approaching a |
| 1206 | definition 1–3. The American Journal of Clinical Nutrition 73: 361s-364s. |
| 1207 | Serrano PH (2005) Responsible use of antibiotics in aquaculture, Food & Agriculture Org. |
| 1208 | Shan X, Xiao Z, Huang W, Dou S (2008) Effects of photoperiod on growth, mortality and |
| 1209 | digestive enzymes in mijuy croaker larvae and juveniles. <i>Aquaculture</i> 281 : 70-76. |

| 1210 | Shapiro JA, Wencewicz TA (2016) Acinetobactin isomerization enables adaptive iron |
|------|---|
| 1211 | acquisition in acinetobacter baumannii through pH-triggered siderophore swapping. |
| 1212 | ACS Infectious Diseases 2: 157-168. |
| 1213 | Sharifuzzaman SM, Al-Harbi AH, Austin B (2014) Characteristics of growth, digestive system |
| 1214 | functionality, and stress factors of rainbow trout fed probiotics Kocuria SM1 and |
| 1215 | Rhodococcus SM2. Aquaculture 418–419: 55-61. |
| 1216 | Sharifuzzaman SM, Austin B (2009) Influence of probiotic feeding duration on disease |
| 1217 | resistance and immune parameters in rainbow trout. Fish & Shellfish Immunology 27: |
| 1218 | 440-445. |
| 1219 | Sorokulova I (2013) Modern Status and Perspectives of Bacillus Bacteria as Probiotics. |
| 1220 | Probiotics & Health 2013: 5. |
| 1221 | Spanggaard B, Huber I, Nielsen J, Nielsen T, Appel KF, Gram L (2000) The microflora of |
| 1222 | rainbow trout intestine: a comparison of traditional and molecular identification. |
| 1223 | Aquaculture 182 : 1-15. |
| 1224 | Srisapoome P, Areechon N (2017) Efficacy of viable Bacillus pumilus isolated from farmed |
| 1225 | fish on immune responses and increased disease resistance in Nile tilapia (Oreochromis |
| 1226 | niloticus): Laboratory and on-farm trials. Fish & Shellfish Immunology 67: 199-210. |
| 1227 | Sugita H, Hirose Y, Matsuo N, Deguchi Y (1998) Production of the antibacterial substance by |
| 1228 | Bacillus sp. strain NM 12, an intestinal bacterium of Japanese coastal fish. Aquaculture |
| 1229 | 165 : 269-280. |
| 1230 | Sumon MS, Ahmmed F, Khushi SS, Ahmmed MK, Rouf MA, Chisty MAH, et al., (2018) |
| 1231 | Growth performance, digestive enzyme activity and immune response of |
| 1232 | Macrobrachium rosenbergii fed with probiotic Clostridium butyricum incorporated |
| 1233 | diets. Journal of King Saud University - Science 30: 21-28. |

Sun Y-Z, Yang H-L, Huang K-P, Ye J-D, Zhang C-X (2013) Application of autochthonous 1234 1235 Bacillus bioencapsulated in copepod to grouper Epinephelus coioides larvae. *Aquaculture* **392**: 44-50. 1236 1237 Sun Y-Z, Yang H-L, Ma R-L, Lin W-Y (2010) Probiotic applications of two dominant gut Bacillus strains with antagonistic activity improved the growth performance and 1238 immune responses of grouper Epinephelus coioides. Fish & Shellfish Immunology 29: 1239 1240 803-809. Sun YZ, Yang HL, Ma RL, Song K, Li JS (2012) Effect of Lactococcus lactis and Enterococcus 1241 faecium on growth performance, digestive enzymes and immune response of grouper 1242 1243 *Epinephelus coioides. Aquaculture Nutrition* **18**: 281-289. Suva M, Sureja V, Kheni D (2016) Novel insight on probiotic *Bacillus subtilis*: Mechanism of 1244 action and clinical applications. Journal of Current Research in Scientific Medicine 2: 1245 1246 65-72. Suzer C, Çoban D, Kamaci HO, Saka Ş, Firat K, Otgucuoğlu Ö, et al., (2008) Lactobacillus 1247 1248 spp. bacteria as probiotics in gilthead sea bream (Sparus aurata, L.) larvae: Effects on growth performance and digestive enzyme activities. Aquaculture 280: 140-145. 1249 Suzuki S, Pruden A, Virta M, Zhang T (2017) Editorial: Antibiotic resistance in aquatic 1250 1251 systems. Frontiers in Microbiology 8. Tal Y, Schreier HJ, Sowers KR, Stubblefield JD, Place AR, Zohar Y (2009) Environmentally 1252 sustainable land-based marine aquaculture. *Aquaculture* **286**: 28-35. 1253 Talpur AD, Ikhwanuddin M, Abdullah MDD, Ambok Bolong A-M (2013) Indigenous 1254 Lactobacillus plantarum as probiotic for larviculture of blue swimming crab, Portunus 1255 pelagicus (Linnaeus, 1758): Effects on survival, digestive enzyme activities and water 1256 quality. *Aquaculture* **416**: 173-178. 1257

| 1258 | Tan LT-H, Chan K-G, Lee L-H, Goh B-H (2016) Streptomyces Bacteria as Potential Probiotics |
|------|---|
| 1259 | in Aquaculture. Frontiers in Microbiology 7. |
| 1260 | Tapia-Paniagua ST, Vidal S, Lobo C, Prieto-Álamo MJ, Jurado J, Cordero H, et al., (2014) The |
| 1261 | treatment with the probiotic Shewanella putrefaciens Pdp11 of specimens of Solean |
| 1262 | senegalensis exposed to high stocking densities to enhance their resistance to disease. |
| 1263 | Fish & Shellfish Immunology 41: 209-221. |
| 1264 | Timmerman HM, Koning CJM, Mulder L, Rombouts FM, Beynen AC (2004) Monostrain, |
| 1265 | multi strain and multispecies probiotics—A comparison of functionality and efficacy. |
| 1266 | International Journal of Food Microbiology 96 : 219-233. |
| 1267 | Truong Thy HT, Tri NN, Quy OM, Fotedar R, Kannika K, Unajak S, et al., (2017) Effects of |
| 1268 | the dietary supplementation of mixed probiotic spores of Bacillus amyloliquifaciens |
| 1269 | 54A, and Bacillus pumilus 47B on growth, innate immunity and stress responses of |
| 1270 | striped catfish (Pangasianodon hypophthalmus). Fish & Shellfish Immunology 60: 391- |
| 1271 | 399. |
| 1272 | Tuomola EM, Ouwehand AC, Salminen SJ (1999) Human ileostomy glycoproteins as a model |
| 1273 | for small intestinal mucus to investigate adhesion of probiotics. Letters in Applied |
| L274 | Microbiology 28: 159-163. |
| 1275 | Tuomola E, Crittenden R, Playne M, Isolauri E, Salminen S (2001) Quality assurance criteria |
| 1276 | for probiotic bacteria. The American Journal of Clinical Nutrition 73: 393s-398s. |
| 1277 | Tzuc JT, Escalante DR, Rojas Herrera R, Gaxiola Cortés G, Ortiz MLA (2014) Microbiota from |
| 1278 | Litopenaeus vannamei: digestive tract microbial community of Pacific white shrimp |
| 1279 | (Litopenaeus vannamei). SpringerPlus 3: 280. |
| 1280 | Ueberschär B (1995) The use of tryptic enzyme activity measurement as a nutritional condition |
| 1281 | index: laboratory calibration data and field application. In: ICES Marine Science |

| 1282 | Symposia. Copenhagen, Denmark: International Council for the Exploration of the Sea, |
|------|--|
| 1283 | 1991, pp. 119-129. |
| 1284 | Vadstein O, Bergh Ø, Gatesoupe F-J, Galindo-Villegas J, Mulero V, Picchietti S, et al., (2013) |
| 1285 | Microbiology and immunology of fish larvae. Reviews in Aquaculture 5: S1-S25. |
| 1286 | Van Doan H, Hoseinifar SH, Dawood MAO, Chitmanat C, Tayyamath K (2017) Effects of |
| 1287 | Cordyceps militaris spent mushroom substrate and Lactobacillus plantarum on |
| 1288 | mucosal, serum immunology and growth performance of Nile tilapia (Oreochromis |
| 1289 | niloticus). Fish & Shellfish Immunology 70 : 87-94. |
| 1290 | Van Doan H, Hoseinifar SH, Khanongnuch C, Kanpiengjai A, Unban K, Srichaiyo S (2018) |
| 1291 | Host-associated probiotics boosted mucosal and serum immunity, disease resistance and |
| 1292 | growth performance of Nile tilapia (Oreochromis niloticus). Aquaculture 491: 94-100. |
| 1293 | Van Doan H, Hoseinifar SH, Khanongnuch C, Kanpiengjai A, Unban K, Van Kim V, et al., |
| 1294 | (2018) Host-associated probiotics boosted mucosal and serum immunity, disease |
| 1295 | resistance and growth performance of Nile tilapia (Oreochromis niloticus). Aquaculture |
| 1296 | 491 : 94-100. |
| 1297 | Verschuere L, Rombaut G, Sorgeloos P, Verstraete W (2000) Probiotic bacteria as biological |
| 1298 | control agents in aquaculture. Microbiology and Molecular Biology Reviews: MMBR |
| 1299 | 64 : 655-671. |
| 1300 | Vine NG, Leukes WD, Kaiser H (2006) Probiotics in marine larviculture. FEMS Microbiology |
| 1301 | Reviews 30 : 404-427. |
| 1302 | Wang L, Ge C, Wang J, Dai J, Zhang P, Li Y (2017) Effects of different combinations of |
| 1303 | Bacillus on immunity and antioxidant activities in common carp. Aquaculture |
| 1304 | International 25 : 2091-2099. |
| 1305 | Wang Y-B, Li J-R, Lin J (2008) Probiotics in aquaculture: Challenges and outlook. Aquaculture |
| 1306 | 281 : 1-4. |

| 1307 | wang Y (2011) Use of problotics Bacillus coagulans, Rhodopseudomonas palustris and |
|------|---|
| 1308 | Lactobacillus acidophilus as growth promoters in grass carp (Ctenopharyngodon idella) |
| 1309 | fingerlings. Aquaculture Nutrition 17: e372-e378. |
| 1310 | Wang H, Wang C, Tang Y, Sun B, Huang J, Song X. (2018) Pseudoalteromonas probiotics as |
| 1311 | potential biocontrol agents improve survival of Penaeus vannamei challenged with |
| 1312 | acute hepatopancreatic necrosis (AHPND)-causing Vibrio parahaemolyticus. |
| 1313 | Aquaculture 497: 30-36. |
| 1314 | Weifen L, Xiaoping Z, Wenhui S, Bin D, Quan L, Luoqin F, et al., (2012) Effects of Bacillus |
| 1315 | preparations on immunity and antioxidant activities in grass carp (Ctenopharyngodon |
| 1316 | idellus). Fish Physiology and Biochemistry 38: 1585-1592. |
| 1317 | Wells JM, Loonen LMP, Karczewski JM (2010) The role of innate signaling in the homeostasis |
| 1318 | of tolerance and immunity in the intestine. International Journal of Medical |
| 1319 | Microbiology 300 : 41-48. |
| 1320 | Wu H-J, Sun L-B, Li C-B, Li Z-Z, Zhang Z, Wen X-B, et al., (2014) Enhancement of the |
| 1321 | immune response and protection against Vibrio parahaemolyticus by indigenous |
| 1322 | probiotic Bacillus strains in mud crab (Scylla paramamosain). Fish & Shellfish |
| 1323 | Immunology 41 : 156-162. |
| 1324 | Wu Z-Q, Jiang C, Ling F, Wang G-X (2015) Effects of dietary supplementation of intestinal |
| 1325 | autochthonous bacteria on the innate immunity and disease resistance of grass carp |
| 1326 | (Ctenopharyngodon idellus). Aquaculture 438: 105-114. |
| 1327 | Wu ZX, Feng X, Xie LL, Peng XY, Yuan J, Chen XX (2012) Effect of probiotic Bacillus |
| 1328 | subtilis Ch9 for grass carp, Ctenopharyngodon idella (Valenciennes, 1844), on growth |
| 1329 | performance, digestive enzyme activities and intestinal microflora. Journal of Applied |
| 1330 | Ichthyology 28 : 721-727. |

- 1331 Xu B, Wang Y, Li J, Lin Q (2009) Effect of prebiotic xylooligosaccharides on growth
- performances and digestive enzyme activities of allogynogenetic crucian carp
- 1333 (*Carassius auratus gibelio*). Fish Physiology and Biochemistry **35**: 351-357.
- 1334 Yan J, Guo C, Dawood MAO, Gao J (2017) Effects of dietary chitosan on growth, lipid
- metabolism, immune response and antioxidant-related gene expression in *Misgurnus*
- anguillicaudatus. Beneficial Microbes 8: 439-449.
- 1337 Yan L, Boyd KG, Grant Burgess J (2002) Surface Attachment induced production of
- antimicrobial compounds by marine epiphytic bacteria using modified roller bottle
- cultivation. *Marine Biotechnology* **4**: 356-366.
- Yanbo W, Zirong X (2006) Effect of probiotics for common carp (*Cyprinus carpio*) based on
- growth performance and digestive enzyme activities. Animal Feed Science and
- 1342 *Technology* **127**: 283-292.
- Yang G, Tian X, Dong S, Peng M, Wang D (2015) Effects of dietary Bacillus cereus G19,
- B. cereus BC-01, and Paracoccus marcusii DB11 supplementation on the growth,
- immune response, and expression of immune-related genes in coelomocytes and
- intestine of the sea cucumber (Apostichopus japonicus Selenka). Fish & Shellfish
- 1347 *Immunology* **45**: 800-807.
- 21348 Zhang Q, Ma H, Mai K, Zhang W, Liufu Z, Xu W (2010) Interaction of dietary *Bacillus subtilis*
- and fructooligosaccharide on the growth performance, non-specific immunity of sea
- cucumber, *Apostichopus japonicus*. Fish & Shellfish Immunology **29**: 204-211.
- Zhao Y, Yuan L, Wan J, Sun Z, Wang Y, Sun H (2016) Effects of potential probiotic *Bacillus*
- cereus EN25 on growth, immunity and disease resistance of juvenile sea cucumber
- 1353 Apostichopus japonicus. Fish & Shellfish Immunology **49**: 237-242.
- Zhao Y, Zhang W, Xu W, Mai K, Zhang Y, Liufu Z (2012) Effects of potential probiotic
- Bacillus subtilis T13 on growth, immunity and disease resistance against Vibrio

| 1356 | splendidus infection in juvenile sea cucumber Apostichopus japonicus. Fish & Shellfish |
|------|--|
| 1357 | Immunology 32 : 750-755. |
| 1358 | Zheng X, Tang J, Ren G, Wang Y (2017) The effect of four microbial products on production |
| 1359 | performance and water quality in integrated culture of freshwater pearl mussel and |
| 1360 | fishes. Aquaculture Research 48: 4897-4909. |
| 1361 | Zhou Z, Wang W, Liu W, Gatlin DM, Zhang Y, Yao B, et al., (2012) Identification of highly- |
| 1362 | adhesive gut Lactobacillus strains in zebrafish (Danio rerio) by partial rpoB gene |
| 1363 | sequence analysis. Aquaculture 370-371: 150-157. |
| 1364 | Zhou T, Zheng XF, Tang JY, Qin JG, Wang Y (2017) Effect of three commercial microbial |
| 1365 | products on bacterial community in a freshwater polyculture system. Aquaculture |
| 1366 | Research 48: 4449-4460. |
| 1367 | Ziaei-Nejad S, Rezaei MH, Takami GA, Lovett DL, Mirvaghefi A-R, Shakouri M (2006) The |
| 1368 | effect of Bacillus spp. bacteria used as probiotics on digestive enzyme activity, survival |
| 1369 | and growth in the Indian white shrimp Fenneropenaeus indicus. Aquaculture 252: 516- |
| L370 | 524. |
| L371 | Zorriehzahra MJ, Delshad ST, Adel M, Tiwari R, Karthik K, Dhama K, Lazado CC (2016) |
| 1372 | Probiotics as beneficial microbes in aquaculture: an update on their multiple modes of |
| 1373 | action: a review. Veterinary Quarterly 36: 228-241. |
| 1374 | |
| 1375 | |
| 1276 | |