Adaptive fluctuation imaging captures rapid subcellular dynamics

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ABSTRACT

In this work we have explored the live-cell friendly nanoscopy method Multiple Signal Classification Algorithm (MUSICAL) for multi-colour imaging of various organelles and sub-cellular structures in the cardiomyoblast cell line H2C9. We have tested MUSICAL for fast (up to 230Hz), multicolour time-lapse sequences of various sub-cellular structures (mitochondria, endoplasmic reticulum, microtubules, endosomes and nuclei) in living cells using low excitation-light dose. Challenges and opportunities with applying MUSICAL for studying rapid sub-cellular dynamics are discussed.

Keywords: fluorescence nanoscopy, fluctuation imaging, live cells, cardiomyoblasts

1. INTRODUCTION

Structural components of living cells are challenging to study because of their size, sensitivity, and highly dynamic nature. Still, understanding the different cellular features and their interactions is crucial for comprehending basic cellular functions, and for finding potential treatments for cellular malfunction. Overcoming the challenges in monitoring the position and morphology of sub-cellular structures in living cells is hence of great interest.

Fluorescence nanoscopy[1] offers excellent contrast and specificity, but time and photon budget are limiting factors when resolution beyond the Abbe limit is required in live-cell imaging. Temporal resolution is limited and determined by the imaging system, technique and implementation, while the photon budget depends on the selected fluorophores, illumination mode and cellular system with varying label densities and chemical environments.

Fluctuation-based nanoscopy techniques like Multiple Signal Classification Algorithm (MUSICAL)[2] extracts super-resolved information from image sequences and are highly feasible for multicolour experiments, contrasting localization-based nanoscopy in these regards. A caveat of time-accumulated live-cell images created from image sequences is, however, that (intra-)cellular dynamics smears structural details, compromising achivable resolution and the reliability of reconstructed feature. The resulting artifacts can be difficult to distinguish from stationary biological components and therefore, imaging speed is critical for reliable image reconstruction.

In this work, we have acquired fluctuation data using a commercial widefield system and sCMOS cameras in fast acquisition mode and tested the abilities and challenges of the computational nanoscopy method MUSICAL in capturing cellular details and dynamics in the rapidly changing and microenvironment of living cardiomyoblasts.

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2. METHODS

The cardiomyoblast cell line H9C2 was cultivated in glass bottom dishes and labelled with live-cell compatible fluorescent labels, taking care to reduce label-induced toxicity. The cells were imaged at room temperature, but in heated media and for less than 30-60 min.

Wide-field, time-lapse data was acquired as fast as the system allowed while maintaining signal-background of about 4 to 1. Various combination of intensity/exposure time and colour channels were acquired.

MUSICAL images were first generated for a single time-point for various number of frames (10-400) and threshold values (based on image stack singular values) to select suitable reconstruction parameters before generating MUSCIAL time-lapses from raw, low intensity image sequences (up to 3000 frames per colour channel). The MUSICAL image reconstructions were generated using a new Python implementation of MUSICAL optimized for high-speed reconstruction, making it feasible to generate MUSICAL time-lapse images of large, multi-colour image stacks [Sebastian citation?].

3. RESULTS AND DISCUSSION

For the current state of MUSICAL, thresholds must be selected manually. We found that a suitable way of selecting the threshold for reconstruction to be such that image features appearing in-focus were emphasized and out-of-focus features were suppressed in the MUSICAL images. 50 frames were in most cases found to give a suitable compromise between motion-smearing and fluctuation data for the algorithm, though longer sequences often appeared to provide a more reliable reconstruction if the organelles remained reasonable stationary within the sequence. For time-lapse sequences where considerable dynamics occurred, the algorithm were found to give a very strong image intensity. At one hand, structural motion created strong image intensity fluctuation that are outcompeting the intrinsic intensity fluctuations of the fluorophores, on the other hand the algorithm appears to provide a window into a new way of analyzing and understanding sub-cellular dynamics, which is a key to understanding living processes and systems.

Figure 1 shows results applying the algorithm in fast acquisition for one colour channel at the time for mitochondria (upper panels) and endoplasmic reticulum (ER; lower panels). Figure 2 displays results applying MUSICAL to four-colour time-lapse data of (b) endosomes, (c) mitochondria, (d) ER and and (e) nucleus. Though no challenge for the algorithm in itself, four-colour acquisition was slow due to necessary switching of fluorescent filters between each time-point (number of channels > number of cameras), translating in to algorithmic challenges in reconstruction the moving objects even for short sequences.

4. CONCLUSION

In conclusion, we have explored the fluctuation nanoscopy algorithm MUSICAL in fast, multicolour nanoscopy of living cardiomyoblasts. The algorithm seems promising with its multi-colour abilities and live-cell compatibility. Challenges include fast three-dimensional motion of living cellular features, modest intrinsic fluorescent intensity fluctuations compared to motion-induced signal fluctuations. Though a analytical challenge, this feature of the algorithm could be exploited in analyzing feature dynamics of e.g. cellular organelles.

Competing interests statement

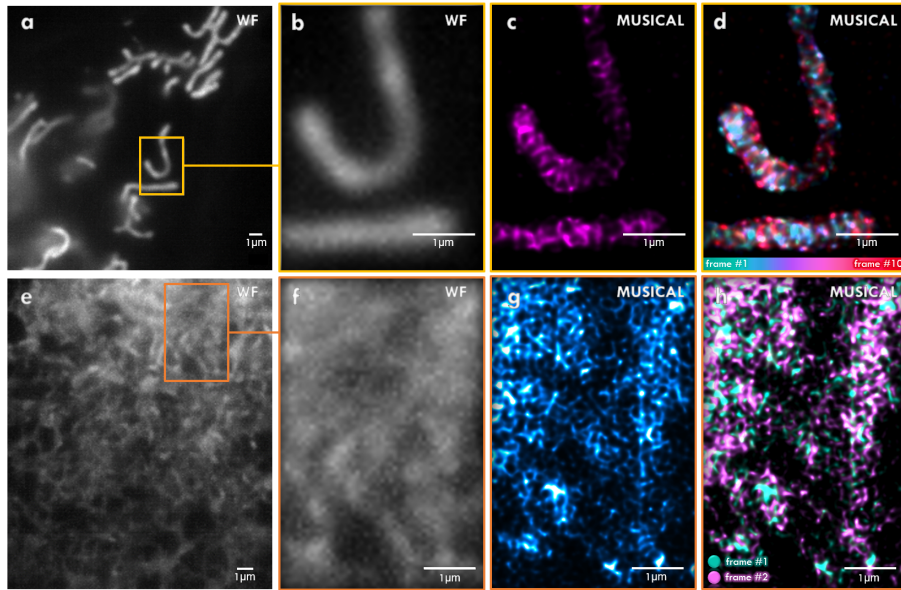The authors declare no competing interests.

Figure 1. **Pushing the limits of time resolution in live-cell nanoscopy**. Obtainable MUSICAL frame rate is limited by achievable system imaging speed and the fluorescence intensity fluctuations of the raw time-lapse image data. *Upper panels*: mitochondrial inner membrane, *lower panels*: endoplasmic reticulum (ER). (a) and (e): widefield images (50 frames low intensity images maximum intensity projected) with indicated regions magnified on the right. (c) and (g) corresponding MUSICAL images obtained from the 50 frames projected on the left. (d) and (h) MUSICAL time-lapse images colour projected as indicated in the colour bars. The different colour projected time-points are 0.22s apart (acquisition time for a sequence of 50 wide-field images).
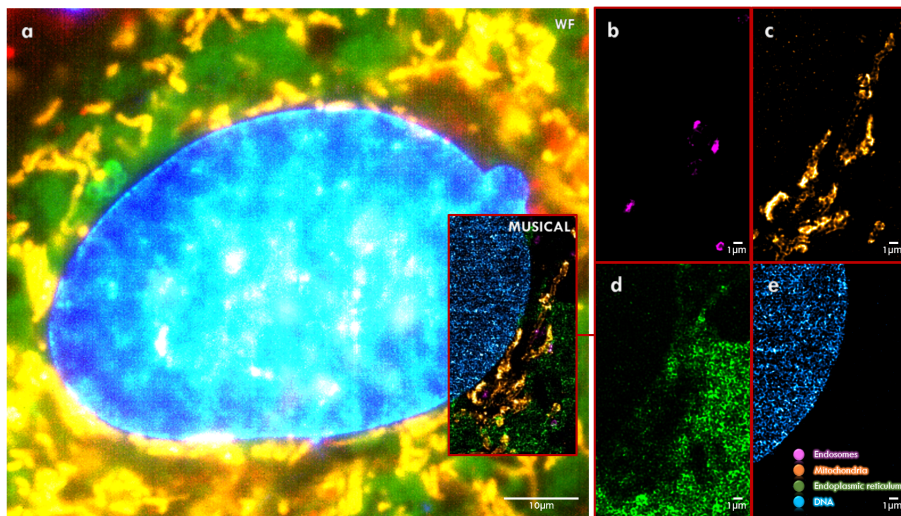

Figure 2. **Multi-colour imaging with MUSICAL.**(a) Single frame widefield image (raw MUSICAL data) of a living cardiomyoblast, labelled with four different fluorescent tags. The four-colour MUSICAL insert is shown to the right with separated channels. (b) endosomes labelled using the membrane probe mCLING-ATTO647N, (c) mitochondrial inner membrane (dsRed-mito). (d) ER fusion protein eGFP-KDEL. (e) Nucleus labelled using Hoechs.

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