### Native Chilean Fruits and the Effects of Their Functional Compounds on Human Health

#### research-article

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#### **Abstract**

In recent years, there has been great interest in the nutraceutical compounds of fruits from native Chilean plant species. In this context, fruits of *Amomyrtus meli* (Meli), *Aristotelia chilensis* (Maqui), *Berberis microphylla* (Calafate), *Luma apiculata* (Arrayán), *Luma chequén* (Chequén), and *Ugni molinae* (Murtilla) growing predominantly in Chilean forests have been studied. This chapter has compiled the existing information about antioxidant activity and antioxidant compound contents of the above mentioned fruit species and their association with the prevention of pathophysiological disorders in humans, such as inflammation, diabetes, and cardiovascular diseases. Results show that the antioxidant compounds of these species, particularly anthocyanins, decrease inflammation as well as the risk of diabetes and cardiovascular diseases. Therefore, consumption of these fruits is a good alternative for preventing cardiovascular and age-related diseases and pathophysiological disorders.

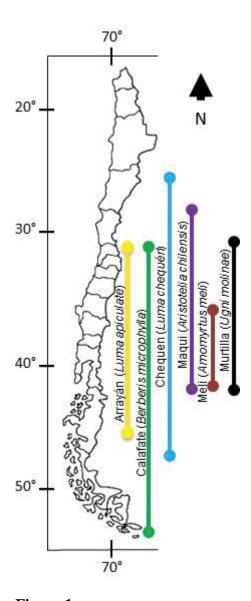
**Keywords:** *Amomyrtus meli*, *Aristotelia chilensis*, arrayán, antioxidant, *Berberis microphylla*, calafate, cardiovascular, carcinogenesis, chequén, diabetes, inflammatory, *Luma apiculata*, *Luma chequén*, maqui, meli, murtilla, phenolic compounds, *Ugni molinae* 

#### 1. Introduction

Chile is a South American country bordered by Perú to the north, Bolivia to the northeast, and Argentina to the east. The continental Chilean territory is a long, narrow strip of land between the Andes in the east and the Pacific Ocean in the west. Its island territories include the Pacific islands of Juan Fernández, Salas y Gómez, Desventuradas, and Pascua and Easter Island in Oceania [1, 2]. Due to its long surface and geographical location, Chile has a large range of different climates from tropical to polar, creating great diversity in its ecosystems and richness in its biodiversity [1–4]. This promotes better use of natural resources for improving the bioeconomy of the country [1, 3]. Traditionally, native Chilean fruits have been used as medicine by Chilean ethnic groups [5]. Currently, studies into native Chilean fruits have focused on identifying and defining their key compounds with medicinal effects [6]. Native Chilean species with edible fruits include woody or shrub forest species belonging to the Elaecarpaceae, Berberidaceae, and particularly Myrtaceae families (Table 1). These families are distributed from Illapel to Tierra del Fuego (Figure 1) and are species-rich in antioxidant and nutraceutical compounds with benefits to human health, including Amomyrtus meli (Meli) (Figure 2), Aristotelia chilensis (Maqui) (Figure 3), Berberis microphylla (Calafate) (Figure 4), Luma apiculata (Arrayán) (Figure 5), Luma chequén (Chequén) (Figure 6), and *Ugni molinae* (Murtilla) (Figure 7) [3, 7–11]. These species inhabit the Valdivian Evergreen Forest (Bosque siempreverde Valdiviano) and the Evergreen Patagonian Forest (Bosque siempreverde Patagónico) together with other Chilean forest species that have not been domesticated. Interestingly, Murtilla is the most domesticated among them [10, 12– 16].

Scientific name of plant	Common name	Family	References
species			
Amomyrtus meli (Phil.) D. Legrand & Kausel.	Meli	Myrtaceae	[ <u>39</u> , <u>54</u> ]
Aristotelia chilensis (Mol.) Stuntz.	Maqui	Elaeocarpaceae	
Berberis microphylla G. Forst.	Calafate	Berberidaceae	
Luma apiculata (DC.) Burret.	Arrayán, Arrayán rojo, Luma, Palo colorado, Temu	Myrtaceae	
Luma chequen (Mol.) A. Gray.	Arrayán Blanco, Chequén	Myrtaceae	
Ugni molinae Turcz.	Murtilla	Myrtaceae	

**Table 1.**Scientific and common name as well as families of the native Chilean fruits.



**Figure 1.**Geographical distribution of native species in the forests of continental Chile.

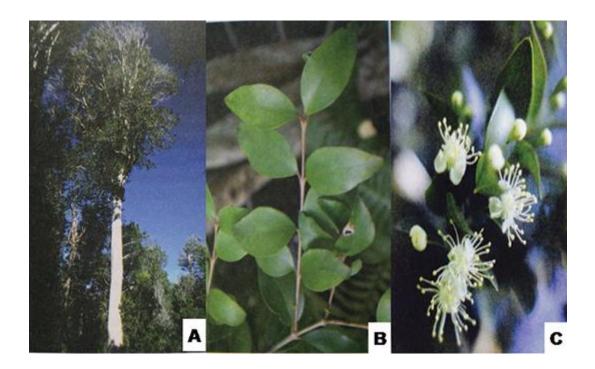


Figure 2.

Pictures of *Amomyrtus meli* (Meli): tree (A), leaves (B), and flowers (C). Extracted from Donoso [40].



**Figure 3.**Pictures of *Aristotelia chilensis* (Maqui): tree (A), leaves (B), and fruits (C).



Figure 4.

Pictures of *Berberis microphylla* (Calafate): flowers (A), fruits (B) and composed pictures of fruits, leaves, and thorns (C).

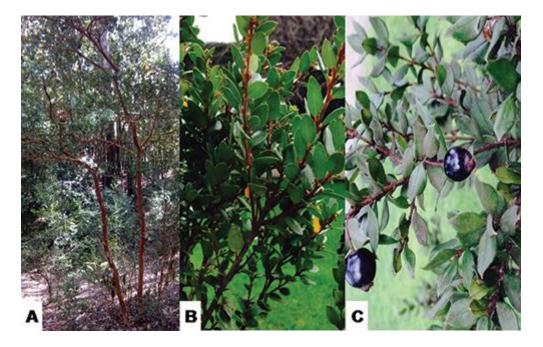
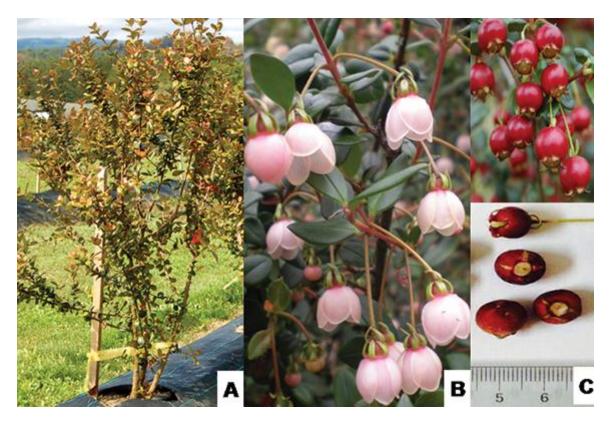


Figure 5.

Pictures of *Luma apiculata* (Arrayán): tree (A), leaves (B), and fruits (C). The picture was replaced by an our picture.



**Figure 6.**Pictures of *Luma chequen* (Chequén): tree (A), leaves (B), and immature fruits (C).



**Figure 7.**Pictures of *Ugni molinae* (Murtilla): tree (A), flowers (B), and fruits (C).

Scalbert et al. [17] and Lila et al. [18] reported a relationship between consumption of fruits with high antioxidant contents and a reduction in oxidative stress in humans. The richness and abundance of enzymatic and nonenzymatic antioxidants in plant species decrease the oxidative damage, helping to inhibit the formation of free radicals by several mechanisms: (1) inhibiting the initiation of the peroxidation, (2) preventing formation of reactive oxygen species (ROS), and (3) breaking the autoxidation chain reaction in humans [19–22]. ROS are normally produced in humans but are exacerbated under exogenous stresses (ozone, cigarette smoking, air pollutants, etc.) [23–27]. Chemical effectiveness of antioxidants against ROS is due to the protection of biomolecules such as lipids, proteins, and carbohydrates to prevent damage by oxidative stress in biomembranes. For this reason, it is very important that people consume antioxidant-rich fruits or foods to decrease ROS [28]. The human diseases associated with oxidative stress are the inflammatory process, cardiovascular disorders, and carcinogenesis (gastric and colorectal) [29–32]. Therefore, fruits with a high antioxidant power are of great benefit in disease prevention related with the inflammatory process and in particular associated with oxidative stress [6, 33–35].

Regarding native fruits rich in antioxidants and other bioactive compounds, it is important to detect the quality and quantity of the compounds responsible for the health beneficial properties [36–38]. In this context, the aim of this review is to associate the main bioactive compounds, antioxidant activity, and content of individual phenolic compounds of some native Chilean fruits such as Arrayán (Figure 5), Calafate (Figure 4), Chequén (Figure 6), Maqui (Figure 3), Meli (Figure 2), and Murtilla (Figure 7) with the prevention of diseases and pathophysiological disorders in humans.

# 2. Morphological characterization, geographical distribution, and ethnobotany of native Chilean fruits

#### 2.1. A. meli (Meli) (Phil.) D. Legrand & Kausel

A. meli is a deciduous tree endemic to Chile, reaching a height up to 20 m (Figure 2A). It has a very distinctive white trunk, which makes it unmistakable among the species of Chilean trees [39]. It grows mostly on moist and shaded sites from the Arauco to Chiloé Regions (Figure 1) in the coastal rainforest, growing on the banks of waterways or under the canopy of other larger species [40]. No conservation problems have been reported for this species, although it is present in a few protected areas such as Chiloé National Park, the Valdivian Coast Reserve, and Oncol Park [39]. Despite its palatability, its fruits are not consumed because they have been reported to cause headaches. The infusion of its leaves is recommended as medicinal use for hypertension (Figure 2B) [14, 39]. It has great ornamental potential due to its abundant and very fragrant flowering (Figure 2C).

#### 2.2. A. chilensis (Maqui) (Mol.) Stuntz

A. chilensis is an evergreen species endemic to Chile (Figure 3A), and it is distributed from Illapel (Coquimbo Region) to Chiloé (Los Lagos Region) (Figure 1) [37]. It grows naturally, forming wild populations named "macales" and can protect against erosion, since it grows rapidly in abandoned, burned, or overexploited soils [41]. The bright green Maqui leaves are oval-lanceolate with a serrated edge (Figure 3B) [42]. The fruits are small, purple bright berries about 5 mm of diameter (Figure 3C), with two to five small seeds inside and are eaten fresh, in juice or in jam, and also used as food coloring and in wine-making [42, 43]. Maqui

fruits have been of great interest to consumers for their high antioxidant activity, mainly due to phenolic compounds (<u>Figure 3C</u>) [44–46], which means this species has been considered a "superfruit" [47].

#### 2.3. B. microphylla (Calafate) G. Forst

B. microphylla, commonly called Calafate, is a native Chilean plant species belonging to the Berberidaceae family (Table 1) [10, 15]. It is an evergreen shrub 2–3 m high with thorns and single yellow flowers (Figure 4A). Its fruit (barberry) is a dark blue-purple berry 7–11 mm in size (Figure 4B and C) [15, 48, 49]. Berberis genus incorporate among 20–60 species in South America, and the majority is commonly known as Michay or Calafate [50, 51]. In Chile, Calafate is distributed from Curicó to Tierra del Fuego at different altitudes and grows under a wide range of ecological conditions (Figure 1) [15, 48, 52]. This species is not cultivated and grows in the wild or in small gardens. Its fruits are used to prepare jam, juice, and wine [10, 15, 48, 50].

#### 2.4. Luma apiculata (Arrayán) (DC) Burret

L. apiculata or Arrayán is an evergreen tree of the Valdivian Evergreen Forest (Figure 5A). Its bark is smooth reddish with whitish parts, reaching heights from 12 to 15 m tall with a crooked trunk around 50 cm in diameter (Figure 5A) [14, 40, 53]. The flowers are hermaphrodites and axillary arranged in groups of three or five, which form a berry, a small, black edible fruit (1.3–1.5 cm in diameter) (Figure 5C) [39, 51]. It is distributed in Chile from the Valparaíso to Aysén Regions (Figure 1) on lakeshores and riverbeds and on very moist soils. Trees of this species are conserved in various National Parks, including La Campana, Radal Siete Tazas, Laguna del Laja, Huerquehue, Puyehue, and Chiloé. Arrayán roots have medicinal uses as an anti-hemorrhagic and astringent, while the bark is used to treat herpes and ulcers, and the leaves (Figure 5B) to heal wounds, and treat stomach disorders [55].

#### 2.5. Luma chequen (Chequén) (Mol.) A. Gray

The Chequén is an endemic tree of the Andean forests in Chile (**Figure 6A**). It is distributed from Coquimbo to Capitán Prat (**Figure 1**) [39]. It grows only in areas of high humidity, on the banks of streams, and in gulches [39]. It is a medium-sized branched shrub, which reaches 9 m in height, with grayish brown bark, and the whole plant gives off a pleasant aroma (**Figure 6A**) [39]. The leaves are oval, wide, and short (**Figure 6B**). The berry fruits are edible and dark purple 1 cm in diameter, ripening in early autumn (**Figure 6C**) [14, 39].

#### 2.6. Ugni molinae (Murtilla) Turcz

Natural habitats of Murtilla are forests and coastal mountains from Valparaiso to Aysén (**Figure 1**). It is an evergreen shrub from 1 to 2 m tall (**Figure 7A**) [56, 57]. The dark green leaves are lanceolate, and the flowers are hermaphrodite (**Figure 7B**) [56]. The fruits are bright red globose berries 5–15 mm in diameter with a sweet taste and a strong aroma (**Figure 7C**) [56, 58]. Murtilla is adapted to most soils and is resistant to drought, wind, and cold, but not frost [59]. The traditional uses are for jams, juices, chocolates, and liqueur production (enmurtado or enmutillado).

# 3. Effect of Chilean fruits on diseases and pathophysiological disorders: beneficial effects on human health

#### 3.1. Bioactive compounds in native Chilean fruits

Native Chilean fruits are naturally rich in phenolic compounds beneficial to human health (Tables 2 and 3) [8, 14, 48]. Phenolic compounds are plant secondary metabolites with bioactive properties and are generally involved in the defense against stress conditions in plants [60, 61]. They can be generally characterized by astringency, color, flavor, odor, and oxidative stability [62, 63]. In recent years, the phenolic compounds of wild or domesticated Arrayán, Chequén, Maqui, Meli, and Murtilla have been studied, with the results highlighting the high antioxidant activity of their leaves and fruits [8, 14–16, 48]. The main phenolic compounds in these fruits can be divided into phenolic acids, flavonoids, flavanols, and anthocyanins [63, 64] (Tables 2 and 3). Ruiz et al. [48] performed a comparison using total antioxidant activity and Trolox equivalent antioxidant capacity (TEAC) methods among edible fruits of Calafate, Maqui, and Murtilla, showing that Maqui and Calafate had a higher antioxidant activity with 88.1 and 74.5 Trolox equivalent (TE) g<sup>-1</sup> of FW, respectively, followed by Murtilla with 11.7 TE g<sup>-1</sup> FW. Afterward, this was confirmed by Dai and Mumper [65], who determined antioxidant activity in Maqui and Murtilla fruits with one of the most commonly used methods, that is, the 2,2-diphenylpicrylhydrazyl (DPPH) assay. The antioxidant activity was higher in Maqui (399.8) than in Murtilla (82.9) in milligrams of crude extract per Liter<sup>-1</sup> (mg of crude extract L<sup>-1</sup>). This method is based on the measurement of the antioxidant compounds able to scavenge the stable free radical DPPH. It is a simple, rapid, and inexpensive method [66]. Usually, the results are expressed as milligrams of a sample that bleached 50% of the DPPH solution (IC<sub>50</sub>) [67]. Therefore, low IC<sub>50</sub> values show a high antioxidant activity. In this antioxidant assay, Brand-Williams et al. [68] reported that to reach IC<sub>50</sub> only 0.0016 g L<sup>-1</sup> of Maqui fruit is necessary; meanwhile, for "Blueberries" (Vaccinium corymbosum), "Strawberries" (Fragaria ananassa), and "Raspberries" (Rubus idaeus) an average of 0.03 g L<sup>-1</sup> of fruits is needed. The result of IC<sub>50</sub> for Maqui fruits was confirmed by Fredes et al. [69] and Céspedes et al. [70], who reported IC<sub>50</sub> values of 0.0012 and 0.0019 g L<sup>-1</sup> by DPPH assay, respectively. This means that Maqui fruits exhibited the highest antioxidant activity compared with other berries cultivated in Chile. Additionally, Dai and Mumper [65] compared phenolic compounds of leaves and fruits, showing that in Magui leaves, concentrations were 200% and in Murtilla, 50% higher than in other fruits. Another commonly used method to determine total antioxidants is the oxygen-radical absorbing capacity (ORAC), which measures the antioxidant values as TE and includes both inhibition time and extent of oxidation inhibition [71, 72]. With this method, Prior et al. [73] reported that fruits of wild Calafate have 25 and 150% higher antioxidant activity than Magui and Murtilla, respectively.

Phenolic compounds	Arrayán	Calafate (B.	Chequén	Maqui	Meli	Murtilla
profile	( <b>L</b> .	microphylla)	( <b>L</b> .	(A.	(A.	(U.
	apiculata)		chequen)	chilensis)	meli)	molinae)
Caffeic acid	ND	+	ND	ND	ND	ND
Catechin	ND	ND	ND	+	ND	+
Chlorogenic acid	+	+	+	ND	+	+

Cinnamicbenzenepropenoic acid         ND         ND         ND         ND         ND         ND         +           Coumaric acid         ND         ND	Phenolic compounds profile	Arrayán (L. apiculata)	Calafate (B. microphylla)	$(\bar{L}.$	Maqui (A. chilensis)	(A.	Murtilla (U. molinae)
Dimethoxy-quercetin		• ,	ND	_			
Ellagic acid ND	Coumaric acid	ND	+	ND	ND	ND	ND
Epigallocatechin gallate	Dimethoxy-quercetin	ND	ND	ND	+	ND	ND
Ferulic acid         ND         +         ND         ND         ND           Feruloyl-quinic acid         +         +         +         ND         +         +         +         +         Feruloyl-quinic acid         +         +         +         ND         +         +         +         +         +         +         +         +         ND         <	Ellagic acid	ND	ND	ND	+	ND	+
Feruloyl-quinic acid	Epigallocatechin gallate	+	ND	ND	ND	ND	ND
Furosinin	Ferulic acid	ND	+	ND	ND	ND	ND
Gallic acid         ND         +         ND         +         ND         +           Hyperoside         +         +         +         +         ND         +         +           Isorhamnetin         ND         +         +         +         ND         ND         ND           7-glucoside         ND         +         ND         ND         ND         ND         ND           1 Sorhamnetin-3-rutinoside         ND         +         ND         N	Feruloyl-quinic acid	+	+	+	ND	+	+
Hyperoside	Furosinin	ND	ND	+	ND	ND	ND
Isoquercitrin	Gallic acid	ND	+	ND	+	ND	+
Isorhamnetin ND + + + ND ND ND ND Isorhamnetin-3-rutinoside-7-glucoside Isorhamnetin-3-galactoside ND + ND ND ND ND ND ND Isorhamnetin-3-galactoside ND + ND ND ND ND ND ND ND Isorhamnetin-3-glucoside ND + ND ND ND ND ND ND ND Isorhamnetin-3-rutinoside ND + ND	Hyperoside	+	+	+	ND	+	+
Isorhamnetin ND + + + ND ND ND ND Isorhamnetin-3-rutinoside-7-glucoside Isorhamnetin-3-galactoside ND + ND ND ND ND ND ND Isorhamnetin-3-galactoside ND + ND ND ND ND ND ND ND Isorhamnetin-3-glucoside ND + ND ND ND ND ND ND ND Isorhamnetin-3-rutinoside ND + ND	Isoquercitrin	+	+	+	ND	+	+
7-glucoside         Isorhamnetin-3-galactoside         ND         +         ND         ND	<del>-</del>	ND	+	+	ND	ND	ND
Isorhamnetin-3-glucoside ND + ND ND ND ND ND Isorhamnetin-3-rutinoside ND + ND ND ND ND ND ND Isorhamnetin-3-(600- ND + ND		ND	+	ND	ND	ND	ND
Isorhamnetin-3-rutinoside ND + ND ND ND ND ND Isorhamnetin-3-(600- ND + ND	Isorhamnetin-3-galactoside	ND	+	ND	ND	ND	ND
Isorhamnetin-3-(600- ND + ND ND ND ND ND ND acetyl)-hexoside  Isorhamnetin-3- ND + ND	Isorhamnetin-3-glucoside	ND	+	ND	ND	ND	ND
acetyl)-hexoside  Isorhamnetin-3- ND + ND ND ND ND ND ND malonylgalactoside  Isorhamnetin-3- ND + ND ND ND ND ND ND ND malonylglucoside  Kaempferol ND + ND	Isorhamnetin-3-rutinoside	ND	+	ND	ND	ND	ND
malonylgalactoside  Isorhamnetin-3- malonylglucoside  Kaempferol ND + ND	•	ND	+	ND	ND	ND	ND
Isorhamnetin-3- ND + ND ND ND ND ND ND malonylglucoside  Kaempferol ND + ND ND ND ND + Kaempferol-deoxyhexoside ND + ND		ND	+	ND	ND	ND	ND
Kaempferol-deoxyhexoside ND + ND ND ND ND ND ND Myricetin + + + + + + + + + + + + + + + + + + +	Isorhamnetin-3-	ND	+	ND	ND	ND	ND
Myricetin + + + + + + + + + + + + + + + Myricetin-3-rutinoside-7- ND	Kaempferol	ND	+	ND	ND	ND	+
Myricetin-3-rutinoside-7- glucoside  Myricetin-3-glucoside  ND + ND + ND ND  Myricetin-3-rutinoside  ND + ND ND ND  Myricetin-3-rutinoside  ND + ND ND ND  Myricetin-3-O-rhamnose + ND ND ND ND  Myricetin-3-galactoside  ND + ND ND ND ND  Myricetin-3-galactoside  ND + ND ND ND ND  ND ND ND  Neochlorogenic acid + + + ND + +  Protocatechuic  ND ND ND ND ND ND  Protocatechuic 3,4-  ND ND ND ND ND ND ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND + ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND + ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND + ND + ND + ND  Protocatechuic 3,4-  ND ND ND ND ND ND + ND + ND + ND + ND	Kaempferol-deoxyhexoside	ND	+	ND	ND	ND	ND
glucoside  Myricetin-3-glucoside  ND  +  ND  +  ND  ND  Myricetin-3-rutinoside  ND  +  ND  ND  ND  ND  ND  ND  Myricetin-3-O-rhamnose  +  ND  ND  ND  ND  ND  ND  ND  ND  ND	Myricetin	+	+	+	+	+	+
Myricetin-3-rutinoside ND + ND ND ND ND ND ND Myricetin-3-O-rhamnose + ND	· ·	ND	+	ND	ND	ND	ND
Myricetin-3-O-rhamnose + ND	Myricetin-3-glucoside	ND	+	ND	+	ND	ND
Myricetin-3-galactoside ND + ND ND ND ND ND ND ND NEochlorogenic acid + + + + ND + + + ND	Myricetin-3-rutinoside	ND	+	ND	ND	ND	ND
Neochlorogenic acid + + + + ND + + Protocatechuic ND ND ND ND + ND ND Protocatechuic 3,4- ND ND ND ND ND ND + Dihydroxybenzoic p-coumaric acid ND ND ND ND + ND +	Myricetin-3-O-rhamnose	+	ND	ND	ND	ND	ND
Protocatechuic ND ND ND + ND ND ND Protocatechuic 3,4- ND ND ND ND ND ND + Dihydroxybenzoic p-coumaric acid ND ND ND ND + ND + ND +	Myricetin-3-galactoside	ND	+	ND	ND	ND	ND
Protocatechuic 3,4- ND ND ND ND ND + Dihydroxybenzoic p-coumaric acid ND ND ND + ND + ND +	Neochlorogenic acid	+	+	+	ND	+	+
Dihydroxybenzoic p-coumaric acid ND ND ND + ND +	Protocatechuic	ND	ND	ND	+	ND	ND
		ND	ND	ND	ND	ND	+
Quercetin + + + + + +	p-coumaric acid	ND	ND	ND	+	ND	+
	Quercetin	+	+	+	+	+	+

Phenolic compounds profile	Arrayán (L. apiculata)	Calafate (B. microphylla)	$(\hat{L}.$	Maqui (A. chilensis)	(A.	Murtilla (U. molinae)
Quercetin-3-rutinoside-7-glucoside	ND	+	ND	ND	ND	ND
Quercetin-3-galactoside	ND	+	ND	+	ND	ND
Quercetin-3-rutinoside	ND	+	ND	ND	ND	ND
Quercetin-3-glucoside	ND	+	ND	+	ND	ND
Quercetin-3- malonylgalactoside	ND	+	ND	ND	ND	ND
Quercetin-3- malonylglucoside	ND	+	ND	ND	ND	ND
Quercetin-3-rhamnoside	ND	+	ND	ND	ND	ND
Quercetin-galloyl-hexoside	ND	ND	ND	+	ND	ND
Quercetin-3-(600-acetyl)-hexoside 1	ND	+	ND	ND	ND	ND
Quercetin-3-(600-acetyl)-hexoside 2	ND	+	ND	ND	ND	ND
Quercetin-3-rhamnoside	ND	+	ND	ND	ND	ND
Quercetin-3-O-(6"-O-galloyl)-hexose	+	ND	ND	ND	ND	ND
Quercetin-3-O-glucose (isoquercitrin)	+	ND	ND	ND	ND	ND
Rutin	+	+	+	ND	+	+
Rutin hydrate	ND	ND	ND	+	ND	ND
Vanillic 4-Hydroxy-3- methoxybenzoic acid	ND	ND	ND	ND	ND	+
Unknown quinic acid derivative	+	ND	+	ND	ND	ND
Unknown gallotannin	ND	ND	+	ND	ND	ND
References	[ <u>8</u> , <u>11</u> , <u>14</u> ]	[ <u>14</u> , <u>15</u> , <u>48</u> ]	[ <u>8</u> , <u>14</u> ]	[ <u>48</u> , <u>81</u> ]	[ <u>14</u> ]	[ <u>14</u> , <u>74</u> , <u>79</u> ]

Table 2.

Identification of phenolic compounds in native Chilean fruits.

<sup>[</sup>i] - +, presence of compounds in fruits.

<sup>[</sup>ii] - ND, not detected.

Anthocyanin profile	Arrayán (L. apiculata)	Calafate (B. microphylla)	Chequén (L. chequen)	Maqui (A. chilensis)	Meli (A. meli)	Murtilla (U. molinae)
Delphinidin-3-glucoside	ND	+	ND	+	ND	+
Cyanidin-3- glucoside	ND	+	ND	+	ND	+
Petunidin-3- glucoside	ND	+	ND	ND	ND	ND
Peonidin-3- glucoside	ND	+	ND	ND	ND	+
Malvidin-3- glucoside	ND	+	ND	ND	ND	ND
Delphinidin-3- rutinoside	ND	+	ND	ND	ND	ND
Cyanidin-3-rutinoside	ND	+	ND	ND	ND	ND
Petunidin-3- rutinoside	ND	+	ND	ND	ND	ND
Peonidin-3- rutinoside	ND	+	ND	ND	ND	ND
Malvidin-3- rutinoside	ND	+	ND	ND	ND	ND
Delphinidin-3- sambubioside	ND	ND	ND	+	ND	ND
Cyanidin-3- sambubioside	ND	ND	ND	+	ND	ND
Delphinidin-3- rutinoside-5- glucoside	ND	+	ND	ND	ND	ND
Petunidin-3- rutinoside-5- glucoside	ND	+	ND	ND	ND	ND
Malvidin-3- rutinoside-5- glucoside	ND	+	ND	ND	ND	ND
Delphinidin-3- sambubioside-5- glucoside	ND	ND	ND	+	ND	ND
Cyanidin-3- sambubioside-5- glucoside	ND	ND	ND	+	ND	ND
Delphinidin-3,5-dihexoside	ND	+	ND	ND	ND	ND
Cyanidin-3,5-dihexoside	ND	+	ND	ND	ND	ND

Anthocyanin profile	Arrayán (L. apiculata)	Calafate (B. microphylla)	Chequén (L. chequen)	Maqui (A. chilensis)	Meli (A. meli)	Murtilla (U. molinae)
Petunidin-3,5-dihexoside	ND	+	ND	ND	ND	ND
Peonidin-3,5-dihexoside	ND	+	ND	ND	ND	ND
Malvidin-3,5- dihexoside	ND	+	ND	ND	ND	ND
Delphinidin-3,5-diglucoside	ND	ND	ND	+	ND	ND
Cyanidin-3,5- diglucoside	ND	ND	ND	+	ND	ND
Delphinidin-3- <i>O</i> -arabinoside	ND	+	ND	ND	ND	ND
Peonidin-3- <i>O</i> -arabinoside	ND	+	ND	ND	ND	+
Peonidin-3- <i>O</i> -di hexoside	ND	+	ND	ND	ND	+
Cyanidin-3- <i>O</i> -di- hexoside	+	ND	ND	ND	ND	ND
Cyanidin-3- <i>O</i> -galactoside	ND	ND	+	ND	+	ND
Petunidin-3- <i>O</i> -galactoside	+	+	+	ND	+	+
Malvidin-3- <i>O</i> -galactoside	+	ND	ND	ND	ND	ND
Delphinidin-3- <i>O</i> -glucoside	ND	+	ND	ND	ND	ND
Cyanidin-3- <i>O</i> -glucoside	+	ND	+	ND	+	ND
Petunidin-3- <i>O</i> -glucoside	+	ND	+	ND	+	+
Peonidin-3- <i>O</i> -glucoside	+	+	+	ND	ND	+
Malvidin-3- <i>O</i> -glucoside	+	+	+	ND	+	ND
Cyanidin-3- <i>O</i> -rutinose	ND	ND	ND	ND	ND	+
Petunidin-3- <i>O</i> -rutinoside	ND	+	ND	ND	ND	+
Cyanidin-3- <i>O</i> -(6-succinoyl)-glucoside	ND	+	ND	ND	ND	+

Anthocyanin profile	Arrayán (L. apiculata)	Calafate (B. microphylla)	Chequén (L. chequen)	Maqui (A. chilensis)	Meli (A. meli)	Murtilla (U. molinae)
Malvidin-3- <i>O</i> -(6-coumaroyl) glucoside	ND	+	ND	ND	ND	ND
Petunidin-3- <i>O</i> -(6-acetyl) glucoside	ND	+	ND	ND	ND	ND
Malvidin-3- <i>O</i> -(6-acetyl) galactoside	+	+	ND	ND	ND	ND
References	[ <u>7</u> , <u>8</u> , <u>10</u> ]	[7, 8, 10, 14, 48, 78]	[8,10,14,]	[ <u>38</u> , <u>46</u> , <u>48</u> , <u>75</u> , <u>78</u> , <u>80</u> , <u>81</u> ]	[ <u>8</u> , <u>14</u> ]	[ <u>10</u> , <u>14</u> , <u>48</u> , <u>79</u> ]

Table 3.

Identification of anthocyanins in native Chilean fruits.

[i] - +: presence of compounds in fruits.

[ii] - ND, not detected.

Genotype, environmental factors, and geographical location are among the main causes for the differences in the antioxidant capacity in native Chilean fruits, since in all studies, fruits were collected in different locations. Mariangel et al. [15] analyzed Calafate fruit antioxidant capacity by DPPH method from different sites in southern Chile (Mañihuales and El Blanco; Aysén and Temuco and Lonquimay; Araucanía). They found that more southern provenances (Mañihuales and El Blanco) showed a higher antioxidant capacity—9.4 and 7.5 TE g<sup>-1</sup> of DW, respectively—than northernmost provenances (Temuco and Lonquimay) with 5.2 and 3.3 TE g<sup>-1</sup> DW, respectively. This suggests that growth conditions have a direct influence on the content of nutraceutical compounds in fruits, as total phenols in Calafate also showed the same trend: 16.1 mg gallic acid equivalent (GAE) g<sup>-1</sup> DW (Lonquimay) and 34.6 mg GAE g<sup>-1</sup> DW (Mañihuales). Thus, fruits from the southernmost region exhibited higher levels of total phenols.

Several types of phenolic compounds have been reported in native Chilean "superfruits," including caffeic acid, ferulic acid, gallic acid, myricetin, *p*-coumaric acid, and others (Tables 2 and 3) [8, 14, 15, 47, 48, 74]. In native berries, total phenolics have been analyzed by Ruiz et al. [48] using the Folin-Ciocalteu method. The results showed a higher total phenol content for Maqui (97 μmol GAE g<sup>-1</sup> FW) followed by Calafate (87 μmol GAE g<sup>-1</sup> FW) and Murtilla (32 μmol GAE g<sup>-1</sup> FW). However, no statistically significant differences were found between Maqui and Calafate. Afterward, Brito et al. [14] found higher values for Calafate (65 mg GAE g<sup>-1</sup> DW) than for Arrayán (27 mg GAE g<sup>-1</sup> DW), Meli (17 mg GAE g<sup>-1</sup> DW), Murtilla (9 mg GAE g<sup>-1</sup> DW), and Chequén (5 mg GAE g<sup>-1</sup> DW). In addition, these studies have recognized anthocyanins as the most important compounds in native Chilean fruits (Table 3) [10, 14, 47, 48, 75–80].

Anthocyanins in Calafate fruits from the Aysén and Magallanes Regions, analyzed by HPLC-DAD, showed total anthocyanin concentrations between 14 and 26 μmol g<sup>-1</sup> FW,

corresponding to the highest values in fruits from the Aysén Region [48]. Comparable anthocyanin values as in Calafate were found in Maqui (16 and 20 μmol g<sup>-1</sup> FW), whereas the lowest values were found in Murtilla (mean 0.2 μmol g<sup>-1</sup> FW) and in Blueberry (2.0 μmol g<sup>-1</sup> FW). The lowest results in Murtilla could be explained by the weaker coloration (rose) of their fruits compared with the black and blue-purple color of the other analyzed fruits [48]. In this context, Mariangel et al. [15] reported differences among Calafate fruits collected in different sites of the Araucanía (Temuco and Lonquimay) and Aysén Regions (Mañihuales and El Blanco). Higher values of cyanidin were found in El Blanco (0.6 mg g<sup>-1</sup> DW), followed by Temuco (0.2 mg g<sup>-1</sup> DW), Mañihuales (0.1 mg g<sup>-1</sup> DW), and Lonquimay (0.06 mg g<sup>-1</sup> DW). These results suggest that anthocyanin concentrations vary depending on the different agro-characteristics of the growth areas and the fruit-ripening time. Brito et al. [14] reported anthocyanin contents (in mg cyaniding 3-O-glucoside g<sup>-1</sup> of DW) with higher values in Calafate fruits (51.6) than in Arrayán (15.2), Meli (13), Murtilla (6.85), and finally Chequén (1). Therefore, it is reported that there is higher anthocyanin content in fruits of Calafate and Maqui compared with other native berries [7, 14, 48, 81].

Due to the difficulty of having fresh fruits rich in antioxidants for consumption out of season, it is very important to know the effect of fruit preservation techniques on the content of bioactive compounds. Among the common techniques in use (convective hot-air, freeze drying, and direct cold), native Chilean fruits have demonstrated minor variation in the concentration of phenolic compounds and antioxidant activity compared with the fresh fruits [46, 82]. The effect of freeze-drying and direct cold on the content of bioactive compounds in native Chilean fruits are the least studied of these techniques, and therefore, they should be explored because cold storage in the postharvest period of these berries may produce fewer changes in the antioxidant levels.

## 3.2. Oxidative stress and antioxidant response in human pathophysiological disorders

Under normal conditions, the human body produces free radicals and other reactive oxygen species (ROS) [29, 83]. ROS are molecules characterized by an unpaired electron in an atomic orbital, being highly unstable, such as hydroxyl radical, superoxide anion radical, hydrogen peroxide, oxygen singlet, hypochlorite, nitric oxide radical, and peroxynitrite radical [83]. In the human body, the ROS molecules are produced in the mitochondria, peroxisomes, in inflammation, phagocytosis processes, and ischemia. They are exacerbated by exposure to stress conditions such as ozone, cigarette smoking, air pollutants, and industrial chemicals, among others [84–86]. The ROS have a high affinity for organic molecules (proteins, carbohydrates, and lipids), causing oxidative stress, damage in biomembranes, and altering body homeostasis [87]. The body counteracts the damage induced by ROS by activating antioxidant systems, which are stable molecules able to donate an electron to the free radical, neutralizing it and reducing ROS damage [88, 89]. The human metabolism produces enzymatic antioxidants such as superoxide dismutase, catalase, xanthine oxidase, lipogenase, and cyclooxygenase, and nonenzymatic compounds such as glutathione, ubiquinol, and uric acid [90]. However, it is also necessary to supply other antioxidants in the diet to strengthen the antioxidant capacity. Among them, vitamin E (α-tocopherol), vitamin C (ascorbic acid), and B-carotene are important. These antioxidants reduce the process of lipid peroxidation, preventing or decreasing oxidative reactions and cell damage [91, 92]. In tissues injured by infection, heat, hypertoxia trauma, and toxin-enhanced oxidative stress, processes are induced in the short-term [36]. To counteract the oxidative damage in tissues, transport of antioxidant enzymes (e.g., xanthine oxidase, lipogenase, and cyclooxygenase) and activation

of phagocytes related to the release of free iron, copper ions, or a disruption of the electron transport chains of oxidative phosphorylation take place [36]. However, when the conditions of oxidative damage are higher than antioxidant defense responses, a critical imbalance between free radical generation and antioxidant defenses occurs. This imbalance in the human body induces complications such as diabetes mellitus, onset, promotion or progression of cancer, and neurodegenerative damage such as Parkinson's disease, among others [93–95]. Epidemiological evidence suggests that diets high in antioxidants reduce the incidence of heart disease, cancer, and neurological disorders, altering the inflammatory process common in these diseases [96–100]. Despite the wealth of information that relates to the consumption of fruits in general with the prevention of diseases linked to oxidative stress, studies into native Chilean fruits are limited.

#### 3.3. Native Chilean fruits as a source of antioxidants

Antioxidant compounds in Chilean berries, such as phenolic acids, flavonoids, flavanols, anthocyanins, and procyanidins, among others, have been widely studied for their highly protective effect on human health, particularly with respect to age-related diseases and pathophysiological disorders related to oxidative stresses [6, 11, 37, 69, 82, 86, 101–104]. Antioxidants have an important role as anti-inflammatory or cancer chemopreventive compounds and against degenerative disorders, decreasing the risk of oxidative stress [6, 99, 105]. Furthermore, anthocyanins in berry fruits have a positive effect on human health, related to their capacity to act as antioxidants, and the protective effect in chronic diseases such as diabetes, cardiovascular diseases, and different type of cancers [6, 10, 14, 64, 106, 107]. Regarding the health beneficial properties of fruits, it is important to study different aspects: the species, the composition of bioactive compounds, their antioxidant capacity, and the capacity of different compounds to modulate the transcription factors of enzymes that induce inflammatory diseases [108, 109]. In this context, the traditional use of the plants is also important. Folk medicine in Chile has used leaves and fruits of Arrayán, Calafate, Chequén, Maqui, Meli, and Murtilla to treat throat pain, ulcers, inflammation, and kidney pain disorders [5, 41, 43]. Interestingly, the richness, abundance, and diversity of bioactive compounds in berries of the Chilean native species have shown effects against some pathophysiological disorders (inflammation, diabetes, and cardiovascular) as reported by Fredes et al. [69], Reyes-Farias et al. [6], Alonso [37], Wellen and Hotamisligil [101], Glass and Witztum [104], Lipfert et al. [86], Fuentes et al. [11], and Genskowsky et al. [82].

Inflammation is a defensive mechanism of the organism to specific noxa (factor producing damage), involving different cellular or humoral agents. Its purpose is to restore the body homeostasis, eliminating the noxae. When the inflammatory process becomes self-perpetuating, it ceases to be beneficial and becomes harmful [110–113]. Cell membrane damage activates the phospholipase A2, favoring the synthesis of arachidonic acid, which serves as a substrate for the formation of lipoxygenase and promotes the oxygenase cycle and leukotriene synthesis, which in turn induce the synthesis of prostaglandins and thromboxanes [114]. This promotes neutrophil chemotaxis, ultimately phagocytizing the damaged cell [115]. In addition, leukotriene favors vascular permeability, facilitating the influx of neutrophils to the injured tissue. In general, berries are important inhibitors of inflammatory processes due to the phenolic compounds like anthocyanins present in their fruits [116–121]. This has been supported by reports about moderate consumption of raspberry, strawberry, and bilberry (*Vaccinium myrtillus*) juices and green and black tea that can help prevent the development of early atherosclerosis [118]. Consumption of lingonberry (*Vaccinium vitis-idaea*) juice for 10 weeks has an anti-inflammatory effect on salt-induced hypertension in rat models,

probably due to high polyphenol concentrations in the juice [119]. In wild blueberries, the fruits have provided *in vivo* evidence of the improvement or the prevention of metabolic disturbances associated with developing obesity, particularly a systemic low-grade inflammation and hypertension in mice [120]. In the case of strawberry, its tannins (as enriched extract or as pure compounds) are able to act on gastric epithelial cells, thereby inhibiting the inflammatory response [121]. More detailed information about the berries and their anti-inflammatory properties has been reported by Yang and Kortesniemi [122] and Joseph et al. [123]. Studies with native berries such as Maqui and Calafate fruits have given evidence that the compounds of these fruits can be considered a good anti-inflammatory agent [116, 124]. The anti-inflammatory activity has been shown *in vivo* in the ears of mice and *in vitro* using macrophages and guinea pigs [125–127]. Rouanet et al. [118] also showed that leaf extracts of these species have anti-inflammatory activity in mice, showing that leaf extracts containing quercetin and kaempferol can reduce inflammation.

Diabetes has been recognized as one of the most important chronic diseases in the world [128]. According to the International Diabetes Federation (IDF), 400 million people worldwide had diabetes in 2013, and it could reach 642 million by 2040 [128]. In 2013 alone, US\$548 billion was spent on diabetes management [128]. Therefore, it is a great challenge to find alternatives to reduce this global epidemic. Type II diabetes comprises disease groups of diverse etiology characterized by the presence of chronic hyperglycemia, altering the secretion and action of insulin, as well as alterations in the metabolism of carbohydrates, proteins, and lipids [128, 129]. Type II diabetes represents over 90% of diabetes cases, and its etiology involves both genetic and environmental factors [130]. In the last few years, a gradual increase in incidences has been reported, inducing metabolic alterations and cardiovascular complications [129–131]. Early treatment with dietary and/or pharmacological hygienic measures in patients with prediabetic states can reduce the incidence of diabetes [132]. However, clearly there is a genetic predisposition that is favored by some factors such as obesity or a sedentary lifestyle [133–135]. In this sense, the adipose tissue sets free inflammatory mediators such as interleukins, tumor necrosis factor (TNF-alpha), or free fatty acids, which increase insulin resistance and oxidative stress [130, 136, 137]. Most of the reports about this disease comprise the effects of commercial berries [138]. Thus, preclinical and clinical studies have suggested that consumption of commercial berries has health benefits with preventive effects on diabetes, improving insulin resistance [122, 139–142]. Studies in obesity-prone rats with a diet containing 2% (wt/wt) freeze-dried powder of highbush blueberry reduced the phenotypes of metabolic syndrome, affecting the gene transcripts of the peroxisome proliferator-activated receptors in adipose and muscle tissues involved in fat and glucose metabolism [143]. This may be due to fibers and/or polyphenols present in the berry fruits. Powder from lingonberries did not change the insulin curve in humans, when consumed together with added glucose [139]. Mursu et al. [140] reported that the intake of berries in the diet may reduce risk of type 2 diabetes in Finnish men. In addition, consumption of blackcurrant (Ribes nigrum) extract in amounts roughly equivalent to 100 g in fruit drinks with low sugar and administered immediately before a high-carbohydrate meal reduced postprandial glycemia, insulinemia, and incretin secretion, being beneficial to human health [141]. Further information about the link between berries and their anti-diabetic properties is available in Yang and Kortesniemi [122] and Tsuda [142]. Nevertheless, there are more limited reports in native Chilean fruits in the form of in vitro and in vivo studies. In this context, Dai and Mumper [65] reported that in in vitro studies, Maqui fruits have hypoglycemic effects that inhibit  $\alpha$ -amylases and  $\alpha$ -glucosidases, enzymes involved in the carbohydrate metabolism. In line with this evidence, the inhibition of both enzymes by Maqui fruits was also confirmed by Schreckinger et al. [144]. Using in vivo assays, Fredes et al. [78]

reported the anti-diabetic properties of Maqui fruits in mice. They showed that oral administration of a standardized anthocyanin-rich formulation from Maqui fruits decreased blood glucose in obese hyperglycemic mice. It has been suggested that Maqui fruits could act by inhibiting sodium glucose cotransporter in the small intestine [145]. However, more studies are needed to confirm this physiological mechanism involved in reducing blood glucose. The results of *in vitro* studies of the biological effects of phenolic compounds are questioned due to the limited bioavailability and absorption of these compounds in the human body [146].

Cardiovascular diseases are a heterogeneous group of pathologies, the common substrate of which is the alteration of different arteries of the body regardless of caliber of arteries [147, 148]. In particular, these supply the brain, heart, lower limbs, and a rta [149]. The cascade of events for atherosclerosis are proliferation of smooth muscle cells, recruitment of inflammatory cells, and lipid deposits within the blood vessel walls, forming plaques of atheroma [149, 150]. This formation prevents normal tissue irrigation, inducing ischemia reduction or loss of blood flow in a tissue [150]. If the atheromatous plaque ruptures, the body in an attempt to repair activated coagulation cascade induces the formation of a platelet plug, which totally or partially obstructs the flow distally [151]. Therefore, the tissues supplied by the artery suffer hypoxia or anoxia due to the induction of tissue necrosis [149, 152, 153]. Plant antioxidants have been shown to reduce cellular oxidative damage and to protect against cardiovascular diseases [154]. Berry consumption has been known to benefit human health with preventive effects on cardiovascular diseases. Thus, Erlund et al. [155] reported that the consumption of moderate amounts of berries such as bilberries, nectar of lingonberries, blackcurrant, strawberry puree, cold-pressed chokeberry (Aronia melanocarpa), and raspberry juice resulted in favorable changes in platelet function, high density lipoprotein (HDL) cholesterol, and blood pressure in male and female volunteers, concluding that the berries may play a role in the prevention of cardiovascular disease. Afterward, Basu et al. [156] deepened the evidence about berry-rich diet that controls the risk of chronic diseases among them the cardiovascular risk. Interestingly, Oudot et al. [157] reported that a high salt diet (8% NaCl) with 2 g/day berries can prevent the cardiac alterations independently of changes in systolic pressure in rats. This was verified by Yang and Kortesniemi [122] and Huang et al. [158], who highlighted that berries are an essential fruit group in heart-healthy diets as a supplementary option to better prevent and control cardiovascular disease in humans. More details are available in Zhu et al. [159] and Rodriguez-Mateos et al. [160]. In the case of native Chilean fruits, the preclinical and clinical studies are more limited, although a recent interest in the study of these berries as a beneficial food to protect the heart is growing. Fredes et al. [69] found that Maqui fruits can significantly reduce the cardiac injury produced by ischemia-reperfusion (I/R) in rat heart in vivo. The I/R injury occurs after a myocardial ischemia, and it is known to generate free radicals, heart injury, and necrosis [69, 161, 162]. Therefore, the high level of phenolic compounds and antioxidant activity of this endemic berry can scavenge free radicals produced by I/R and protect the heart [69]. Similarly, concentrated Maqui juice has a high capacity for the oxidation of low-density lipoproteins, which is considered one of the first steps in the development of atherosclerosis [163]. In this sense, Fuentes et al. [11] suggested that Arrayán fruit extracts could protect endotheliumdependent vasodilation (measure to probe endothelial function in different pathophysiological disorders), which is impaired by high glucose [11]. Consequently, they consider that the extract may have an important use in the prevention of vascular damage induced by high glucose. In addition, Falkenberg et al. [164] showed an inhibition of "platelet aggregation" (induced by adenosine diphosphate and collagen) in sheep and human blood through the application of Arrayán and Chequén extracts, which was confirmed by the inhibition of

platelet surface activation markers. Afterward, research showed that Murtilla fruits demonstrated vasodilator activity in the aortic rings [165]. Another important damage parameter used as a marker for the risk of developing heart disease is the low-density lipoproteins (LDL). Maqui extracts may reduce oxidative modifications of LDL in overweight people and smokers [124].

Finally, all the properties mentioned of the native Chilean fruits place them in the category of "superfruits" due to their excellent biological effects on human health. Thus, these "superfruits" could be used as nutraceuticals and functional foods with potential use in the human health industry.

#### 4. Conclusion

We conclude that Arrayán, Chequén, Maqui, Meli, and Murtilla fruits are rich in antioxidant compounds with high antioxidant power, such as anthocyanins. The results show that the consumption of native Chilean fruits reduces the risk of pathophysiological disorders such as inflammation, diabetes, and cardiovascular disease due to their bioactive compounds. Therefore, these berries have a potential for increased use in the functional food and nutraceutical industries.

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