

1 **Surface etched carbon nanofiber companied ytterbium oxide for pinch level**
2 **detection of fungicides carbendazim**

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23 **Abstract**

24 In the present work, we demonstrate for the first time the use of a ytterbium oxide nanorod/carbon
25 nanofiber ($\text{Yb}_2\text{O}_3/f\text{-CNF}$) hybrid nanocomposite for the electrochemical detection of carbendazim
26 (CBZ) fungicide. Various physicochemical methods such as Field Emission Scanning Electron
27 Microscopy (FE-SEM), Transmission electron microscopy (TEM), and X-ray diffraction (XRD)
28 spectroscopy have been used to confirm the formation of Yb_2O_3 , $f\text{-CNF}$, and $\text{Yb}_2\text{O}_3/f\text{-CNF}$
29 composite. Cyclic voltammetry and differential pulse voltammetry were employed to investigate
30 different electrode electrochemical properties for the detection of CBZ. Compared with Yb_2O_3 and
31 $f\text{-CNF}$, the $\text{Yb}_2\text{O}_3/f\text{-CNF}$ nanocomposite exhibits the highest electrocatalytic activity toward the
32 oxidation of CBZ. Under optimized conditions, the developed sensor shows a low detection limit
33 (6 nM) and a wide linear range (50 nM to 3035 μM) with good sensitivity ($0.2899 \mu\text{A } \mu\text{M}^{-1} \text{cm}^{-2}$).
34 Moreover, the fabricated sensor has demonstrated excellent stability, anti-interference ability,
35 reproducibility, repeatability, and practicality. In addition, these sensor electrodes are easily
36 prepared, affordable, portable, and effective at detecting CBZ fungicide residues in food and the
37 environment.

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42 **Keywords:** Yb_2O_3 nanorods; $f\text{-CNF}$; Nanocomposites; Carbendazim; Electrochemical sensor

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44 1. Introduction

45 Globally, pesticide treatments during cultivation are gradually increasing to further the control and
46 eradication of a variety of crop diseases [1], [2], [3]. Pesticide residues can be found in vegetables,
47 fruits, processed food, water, air, and soil due to excessive and uncontrolled usage [4], [5], [6].
48 Carbendazim (methyl 2-benzimidazole carbamate, CBZ) is a harmful fungicide that belongs to the
49 -IV family and is extensively used to eradicate weeds post-harvest and before planting seeds [7],
50 [8], [9]. A stable benzimidazole ring makes CBZ resistant to environmental degradation.
51 Moreover, the CBZ can persist in the environment for a long time, resulting in severe health
52 problems and environmental risks [10], [11]. According to the Brazilian National Health
53 Surveillance Agency (BANVISA) and China, 0.02-0.03 mg/kg of CBZ can be used in certain fruits
54 like citrus fruits, apples, grapes, and bananas [12], [13]. On the other hand, the International Codex
55 Alimentarius Commission (ICAC) recommends levels of CBZ varying from 0.05 to 20 mg kg⁻¹
56 for crops [14][15]. However, CBZ traces can adversely affect humans, such as infertility,
57 embryotoxicity, carcinogenic and mutagenic effects, testicular damage, and endocrine disruption
58 [16]. The World Health Organization (WHO) has classified CBZ as a hazardous chemical because
59 of its toxicity [17]. Even though CBZ has been banned in many countries (USA, Australia,
60 Sweden, European Union), it is still used in cultivation due to its low prices and high yields [9].
61 Therefore, it is crucial to monitor CBZ levels accurately and rapidly in food products and water
62 sources [18–20]. A variety of advanced analytical techniques have been used to detect CBZ in
63 food and water to date, including chemiluminescence [21], high-performance liquid
64 chromatography [22], Raman scattering [23], UV-Vis spectrometry [24], and electrochemical
65 detection [15]. There are disadvantages to traditional chromatographic and spectrometric methods,
66 such as their complex free treatment process, expansive instrumentation, inability to observe in

67 real-time, and long duration [25], [26]. In contrast, electrochemical methods have generated
68 considerable attention due to their ease of operation, high accuracy, inexpensive equipment, quick
69 response, and minimal chemical usage [27], [28]. Therefore, this study uses an electrochemical
70 method to detect CBZ in food and water samples.

71 Lanthanide-based nanomaterials play a significant role in a variety of fields due to their unique
72 properties such as unquenched magnetic moments, high magnetic susceptibilities, various
73 oxidation states, electropositive properties, electronic relaxation times, partially filled semi-core
74 4f subshells, long excited-state lifetimes (>1 second), high dielectric constants, high resistivity,
75 and large band gap energies [29]. Among them, lanthanide oxides have received sustained
76 attention in diverse fields, such as catalysis, gas sensors, electrochemical sensors, temperature
77 sensors, solar cells, and supercapacitors [30], [31]. They also have excellent mechanical, chemical,
78 and thermal stability, high corrosion resistance, and low toxicity [32], [33], [34]. Ytterbium oxide
79 (Yb_2O_3) nanomaterials have been chosen in this study because of their desirable catalytic
80 properties, electrochemical redox properties, optical properties, and high electron transfer
81 mediators on their surfaces. In addition, the high surface energies make them highly sensitive to
82 the detection of targeted analytes [35], [36]. To date, Yb_2O_3 has been synthesized with different
83 structures by many different methods, including hydrothermal [37], solution-doping technique
84 [38], coprecipitation [39], sol-gel [40], electro-spinning [41], etc. However, the
85 synthesized Yb_2O_3 nanoparticles have agglomerated on the electrode surface, reducing its
86 efficiency during electrochemical analysis [42], [43]. It is possible to overcome these obstacles by
87 using a chemically functionalized carbon nanofiber (*f*-CNF) substrate since carbonaceous
88 nanomaterials are renowned candidates for electrode fabrication and can prevent agglomeration
89 [33], [44]. It is worth noting that *f*-CNF has gained desirable properties, including considerable

90 mechanical strength, porosity, and uniformity of its surface. Additionally, its unique surface
91 properties can enhance electron conductivity and electrochemical performance due to its
92 cylindrical nanostructure and surface functional groups [45], [46], [47]. The hollow structure of
93 the carbon nanofiber network structure facilitated molecular interaction, electrochemical activity
94 on the electrode surface, and mechanical interlocking of the nanofibers facilitated electron transfer.
95 Additionally, the *f*-CNF has a large surface area and appropriate pore size distribution, which make
96 it a valuable candidate for electrochemical sensors [48], [49]. The combined nanocomposite of
97 Yb₂O₃ with highly conducting properties and negative charge surfaces of *f*-CNFs can provide
98 additional synergistic properties. Furthermore, the *f*-CNF enhances Yb₂O₃ electrocatalytic activity
99 and allows its use in modified electrode materials for highly sensitive and selective detection of
100 pesticides. [50], [51]. According to a literature survey, Yb₂O₃ nanoparticles modified
101 carbonaceous electrodes were used for electrochemical determination of biological and hazardous
102 molecules. For instance, Raja et al. [52] Ytterbium oxide/graphene oxide has been prepared with
103 a green approach method, and a modified electrode was used to detect the 3-nitro-L-tyrosine.
104 Mohammad et al. [32] RGO/ Yb₂O₃ NPs modified glassy carbon electrode for sensitive detection
105 of metoclopramide in human fluids. Similarly, Carolina et al. used graphene/Yb₂O₃ composite-
106 based carbon paste electrodes with good sensitivity and low detection limit (LOD) for the
107 electrochemical detection of acetaminophen [53]. In the present study, Yb₂O₃ nanorods decorated
108 with *f*-CNF (Yb₂O₃/*f*-CNF) hybrid composite-modified electrodes were prepared for selective and
109 sensitive detection of toxic CBZ. In the literature, there are no sensor studies on CBZ that use
110 either Yb₂O₃ or its composite so this study would draw the researcher's attention to CBZ's toxicity
111 and detection mechanisms. In this study, we developed a novel electrochemical sensor using
112 Yb₂O₃/*f*-CNF nanocomposite to detect CBZ in food and water samples (carrots, reddish water,

113 lake water, pond water). The sensor proposed in this study has a wider linear range and a lower
114 LOD for the detection of CBZ. Additionally, real samples were analyzed to demonstrate the
115 applicability of the developed CBZ sensor.

116 **2. Materials and methods**

117 **2.1. Materials and reagents**

118 Ytterbium acetate ($\text{Yb}(\text{C}_2\text{H}_4\text{O}_2)$, 99.9%), ammonium acetate ($\text{C}_2\text{H}_7\text{NO}_2$, 98%), and carbendazim
119 (CBZ) were purchased from Sigma-Aldrich and used without further purification. Monosodium
120 phosphate (NaH_2PO_4 (s), $\geq 99\%$) and disodium phosphate (Na_2HPO_4 (s), $\geq 98.5\%$) were used to
121 make different concentrations supporting electrolyte solution during electrochemical analysis, and
122 HCl and NaOH were used to adjust the pH value. Nitric acid (HNO_3) and sulfuric acid (H_2SO_4)
123 were purchased from Sigma Aldrich. Potassium ferricyanide ($\text{K}_3\text{Fe}(\text{CN})_6$ (s), $\geq 99\%$), potassium
124 ferrocyanide ($\text{K}_4\text{Fe}(\text{CN})_6$ (s), $\geq 99\%$), potassium chloride (KCl (s), $>99\%$), sodium hydroxide
125 (NaOH, $>98\%$), and ethanol (CH_2OH (l), $>98\%$) were purchased from Sigma-Aldrich.

126 **2.2. Synthesis of Yb_2O_3 nanorods**

127 A coprecipitation method was used to synthesize Yb_2O_3 nanorods. Initially, 0.1 M of
128 $\text{Yb}(\text{C}_2\text{H}_4\text{O}_2)$ was dissolved in 50 mL of DI water and stirred continuously at room temperature. In
129 addition, a few drops of ammonium acetate were gently added to the solution and continuously
130 stirred for 2 hours at 350 rpm. A uniform, clean blue precipitate was obtained at the end of the
131 reaction. The reaction mixture in the beaker was sonicated for 1 h for homogeneous formation
132 and the resulting residue was washed with deionized (DI) water and ethanol to remove any external
133 impurities, then centrifuged at 400 rpm to remove any residual impurities. The precipitate was then
134 dehydrated at 55°C for 24 h and calcined in a muffle furnace at 600°C for 6 h.

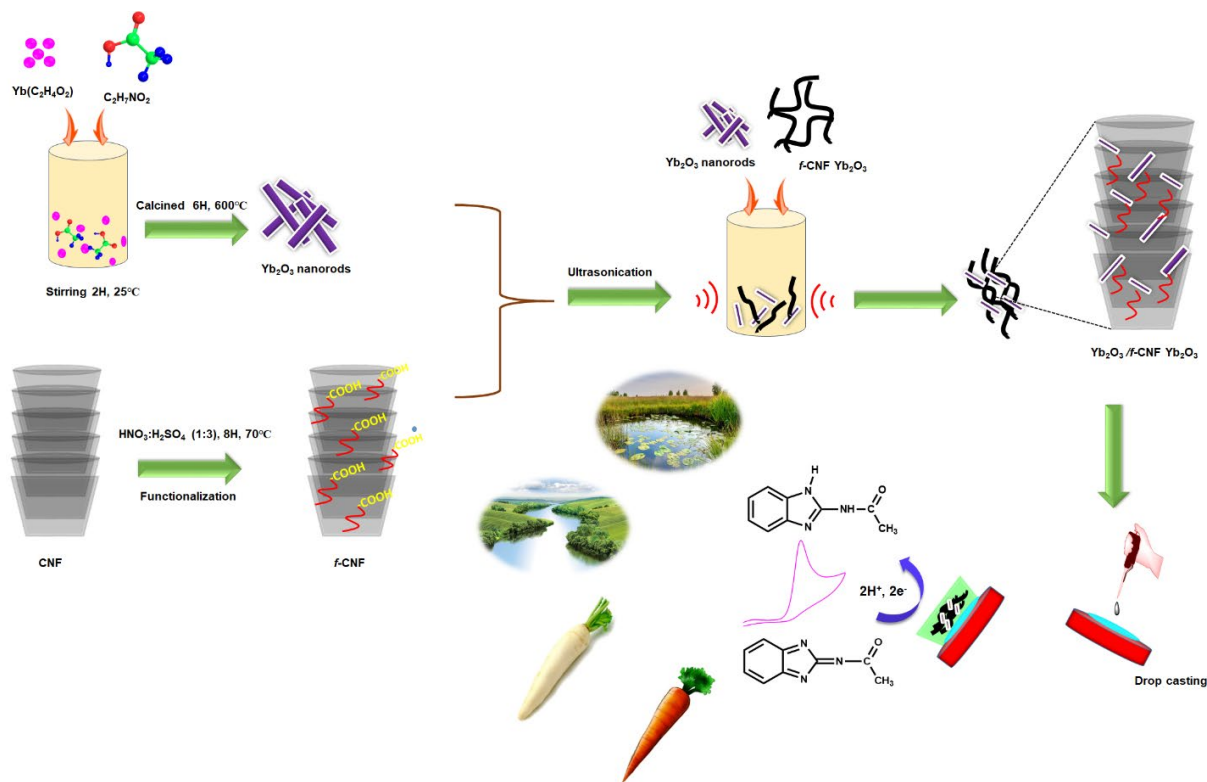
135 **2.3. Synthesis and fabrication of $\text{Yb}_2\text{O}_3/f$ -CNF**

136 To prepare the $\text{Yb}_2\text{O}_3/f\text{-CNF}$, 5 mg of as-prepared Yb_2O_3 and 2 mg of $f\text{-CNF}$ were added to 1 mL
137 of DI water. The composite suspension was kept under the ultrasonication bath until it reached a
138 homogeneous suspension. The suspension was centrifuged for 15 minutes at 400 rpm to remove
139 impurities and washed numerous times with DI water and ethanol. The attained $\text{Yb}_2\text{O}_3/f\text{-CNF}$
140 composite was dried at 80 °C for 8 h. As expected, the presence of oxidized carboxylic functional
141 groups of $f\text{-CNF}$ holds on the wall to the Yb_2O_3 nanorod over the outer interlayers of the
142 structure during ultrasonication. The overall synthesis process of $\text{Yb}_2\text{O}_3/f\text{-CNF}$ is shown
143 in **Scheme 1**. For the electrode preparation, about 5 μL of the above-prepared $\text{Yb}_2\text{O}_3/f\text{-CNF}$
144 composite suspension was dropped on the glassy carbon electrode (GCE) and dried in an air oven.
145 The fabricated $\text{Yb}_2\text{O}_3/f\text{-CNF}$ composite electrode was utilized for further electrochemical and
146 catalytic studies. The same procedure was followed to prepare the other electrode.

147 **2.4. Apparatus and measurements**

148 The powder X-ray diffraction (XRD) patterns of the synthesized Yb_2O_3 , $f\text{-CNF}$, and $\text{Yb}_2\text{O}_3/f\text{-CNF}$
149 were measured by a PAN analytical X'Pert PRO diffractometer equipped with Cu $K\alpha$ radiation (λ
150 = 1.5418 Å). The crystal structure of all samples was investigated from 10 to 90° 2θ with a step
151 size of 0.02°. A JASCO FT-IR 460 Plus spectrophotometer was used to measure the IR spectrum
152 of synthesized samples. The morphology, size, and surface features were examined using a High-
153 resolution Transmission Electron Microscope (HR-TEM) and scanning images, selected area
154 electron diffraction (SEAD), and elemental mapping was obtained using an HR-TEM: JEOL -
155 2100F. X-ray photoelectron spectroscopy (XPS, Thermo Scientific Multi-Lab 2000) was used to
156 analyze the prepared samples' elemental composition and oxidation state. pH measurements were
157 performed with a Horiba-L aqua pH meter calibrated with a standard buffer of the defined pH. All
158 electrochemical experiments were carried out with a conventional three-electrode system equipped

159 with a modified GCE (3 mm diameter) as a working electrode, platinum wire as an auxiliary
 160 electrode, and Ag/AgCl (sat. KCl) as a reference electrode. The electrochemical studies were
 161 carried out using CHI 750A electrochemical workstation.



162
 163 **Scheme 1.** Synthesis of individual Yb₂O₃ nanorods, *f*-CNF, and *f*-CNF/Yb₂O₃ composite and
 164 detection mechanism of CBZ in food and water samples.

165 3. Results and discussion

166 3.1. XRD and FTIR analysis

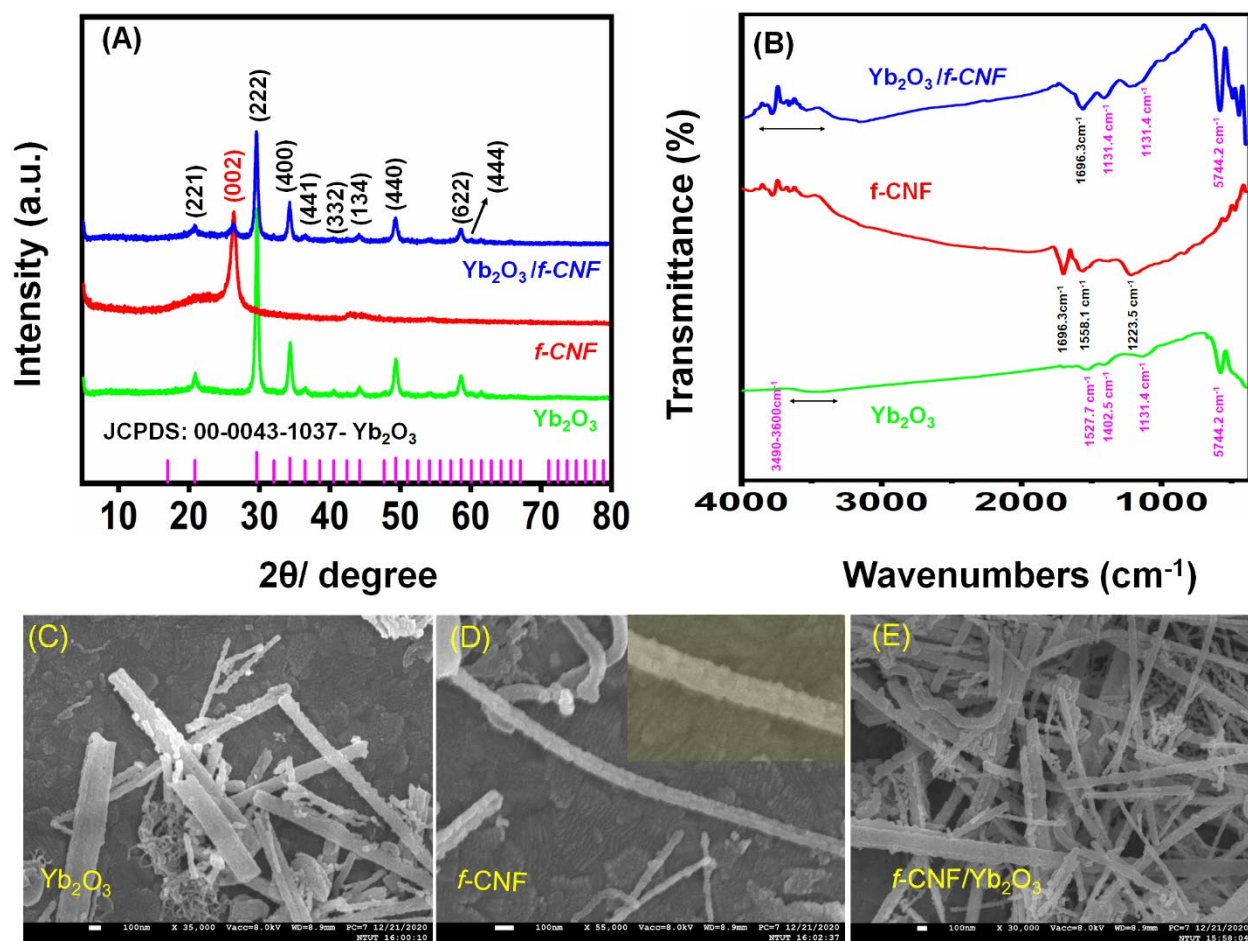
167 The crystal structure and phase purity of as-synthesized Yb₂O₃, *f*-CNF, and *f*-CNF/Yb₂O₃
 168 composite were explored using XRD. The XRD pattern of Yb₂O₃ in **Fig. 1A** showed diffraction
 169 peaks at 2θ of 20.8°, 29.6°, 34.8°, 36.5°, 40.5°, 44.2°, 49.3, 58.6, and 61.5°, indicating lattice
 170 planes (211), (222), (400), (441), (332), (134), (440), (622) and (444), respectively, which are
 171 associated with BCC phase, space group-La₃ and well-matched with the previously reported

172 literature [JCPDS:0-0043-1037] [54]. The average crystalline size of Yb₂O₃ was calculated
173 through ‘Debye–Scherrer’s equation’[55]:

$$174 \quad D_{XRD} = K\lambda / \beta_{hkl} \cos\theta \quad (1)$$

175 where λ is the wavelength of X-ray (0.15406 Å), D represents the size of crystallites (nm), θ is the
176 diffraction angle, k is a constant value, and β is full with half maximum (FWHM) of the diffraction
177 peak. The calculated average crystalline size (D) of the Yb₂O₃ was 19.08 nm. The lattice unit cell
178 parameters were calculated as = 27.16 Å with a cell volume of 1136.59 Å³. These results show that
179 the synthesized Yb₂O₃ consists of only Yb and O, with no impurities detectable. The XRD pattern
180 of the *f*-CNF is shown in **Fig. 1A**; the observed diffraction peaks at 24.4° and 42.2° were well
181 indexed with (002) and (100), respectively, confirming the presence of carbon source [56].
182 Combining Yb₂O₃ with *f*-CNF resulted in lower metal oxide influence on the carbon surface
183 because of the intensity of *f*-CNF.

184



185
 186 **Fig. 1.** (A) XRD and (B) FTIR spectrum of Yb₂O₃, f-CNF, and Yb₂O₃/f-CNF. FE-SEM images of
 187 Yb₂O₃ (C), f-CNF (D), and Yb₂O₃/f-CNF (E).

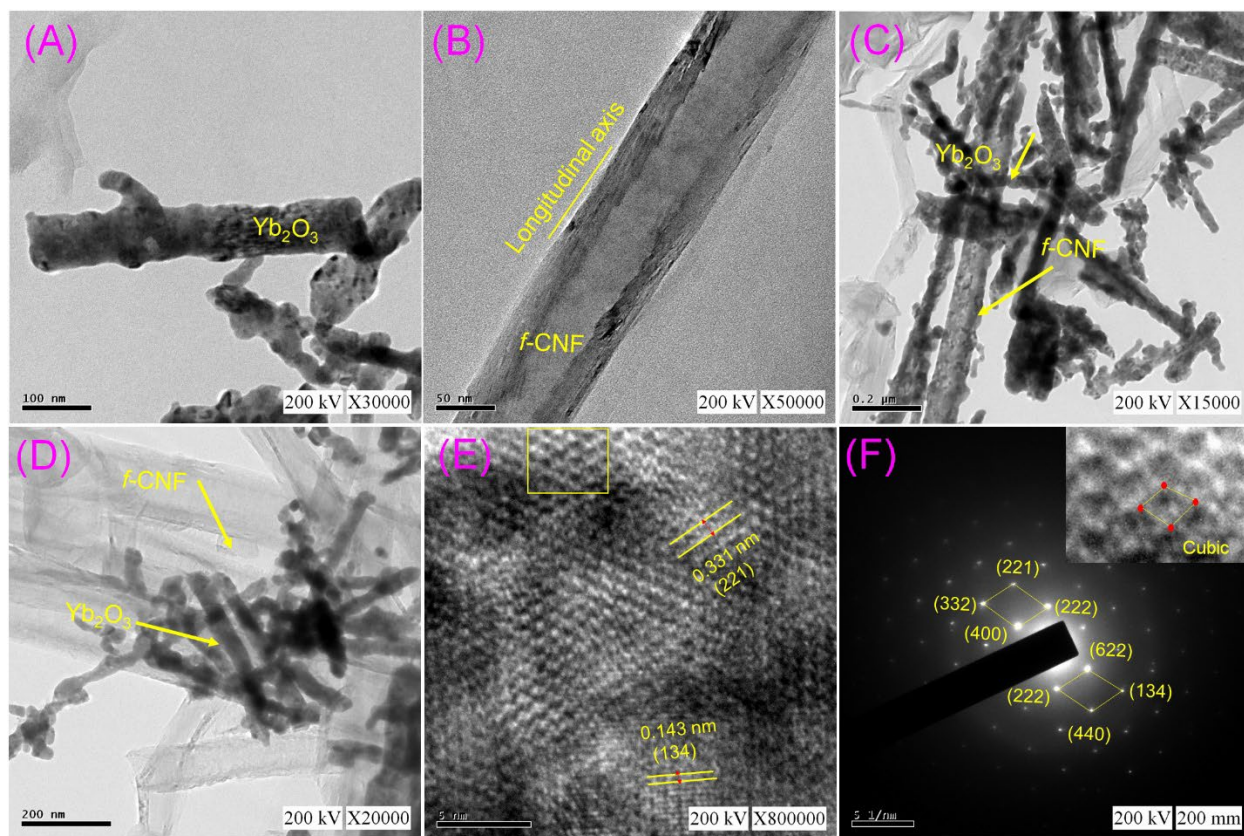
188 FTIR spectroscopy was used to find the functional groups of the as-prepared Yb₂O₃, f-
 189 CNF, and f-CNF/Yb₂O₃ nanocomposite. The FTIR spectrum of Yb₂O₃ showed a broad peak at
 190 3490 – 3600 cm⁻¹ corresponding to the adsorption of -OH stretching vibration, and three blend
 191 peaks noticed at 1131.4, 1402.5, and 1527.7 cm⁻¹ are attributed to the CO₃²⁻ stretching vibration
 192 (Fig. 1B) [57]. A significant peak appears at 574.2 cm⁻¹, indicating the formation of Yb-O
 193 stretching vibration (metal-oxygen bond), which proves the formation of Yb₂O₃ nanoparticles
 194 [40][58]. The FTIR spectrum of f-CNF revealed sustainable peaks at 1223.5, 1558.1, 1696.3, 1701,
 195 and 3462 cm⁻¹, which belong to the acid-treated functional groups C-O, C=C, C=O, and -OH,

196 respectively [46][59]. Finally, all observed peaks appeared in the *f*-CNF/Yb₂O₃ nanocomposite
197 without any other impurities.

198 3.2. Surface and structural analysis

199 The surface structure, morphology, and size of Yb₂O₃, *f*-CNF, and Yb₂O₃/*f*-CNF were confirmed
200 using FE-SEM and HR-TEM studies. **Fig. 1C** shows a single nanorod structure morphology of
201 Yb₂O₃. It is possible for two-dimensional Yb₂O₃ nanorods to provide more active sites and more
202 electrode/electrolyte contact areas, and accelerate electron transfer activity at the electrochemical
203 reaction by their uniform surface morphology. **Fig. 1D** depicts the morphological features of
204 surface functionalized *f*-CNF, the one-dimensional hollow structure, and the rough surface caused
205 by the chemisorption of the organic substrate on the carbon surface after functionalization. As
206 shown in **Fig. 1E**, the ultrasonically fabricated composite of the Yb₂O₃/*f*-CNF revealed Yb₂O₃
207 fused to the surface of the *f*-CNF. Also, the HR-TEM images confirmed the individual structure
208 of Yb₂O₃ nanorods and *f*-CNF (**Fig. 2A** and **B**). From **Fig. 2A**, the diameter and length of Yb₂O₃
209 nanorods were calculated as 98 and 274 nm, respectively. The higher and lower magnification
210 TEM images of the composite reveals that the Yb₂O₃ nanorods are fused around the surface walls
211 of the *f*-CNF (**Fig. 2C** and **D**), which result from the strong attraction of chemically induced
212 oxygen functional groups (-COOH, -OH) by Yb₂O₃ nanorods. As shown in the HR-TEM image
213 of **Fig. 2E**, the d-spacings calculated for the Yb₂O₃ cubic crystal system (maximized in **Fig. 2F**)
214 were 0.331 and 0.143 nm, respectively, which is consistent with XRD results. The lattice fringes
215 of Yb₂O₃ are visible, indicating its high crystallinity, and *f*-CNF did not affect the crystallinity of
216 Yb₂O₃ nanorods in the nanocomposite (**Fig. 2F**). Based on the Energy-dispersive X-ray (EDX)
217 analysis (**Fig. S1A**), the percentages of the elements were determined as Yb (61.1%), O (7.44%),

218 and Yb (31.18%). Additionally, the elemental mapping images of S1. B-C reveals Yb, O, and C
 219 in the Yb₂O₃/f-CNF composite.

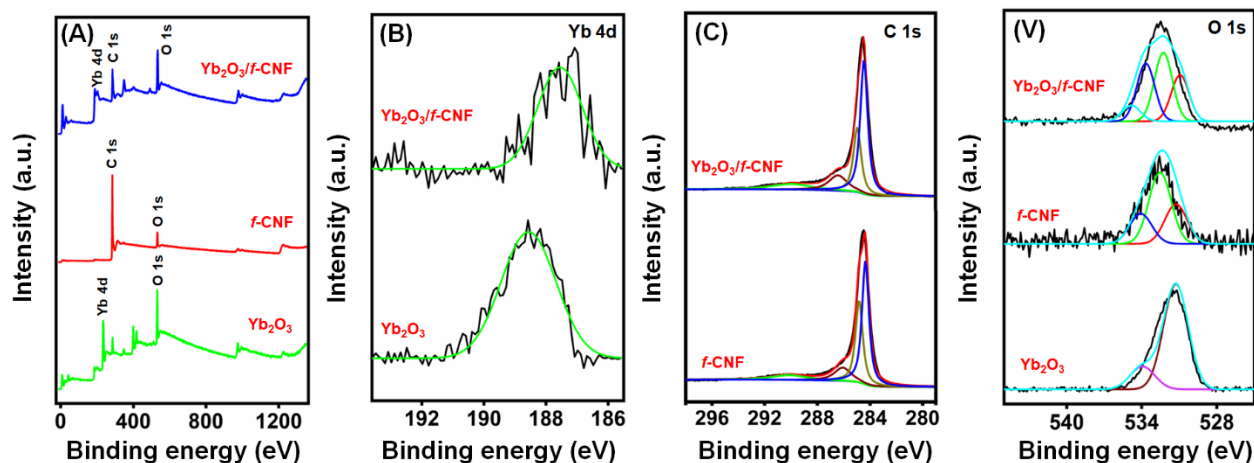


220
 221 **Fig. 2.** TEM images of (A) Yb₂O₃, (B) f-CNF, (C and D) Yb₂O₃/f-CNF, and (E) Lattice fringe and
 222 (F) SEAD pattern of Yb₂O₃.

223 3.3. XPS analysis

224 The chemical composition and oxidation state of Yb₂O₃/f-CNF nanocomposite are revealed by
 225 XPS. The overall scanning survey spectra (**Fig. 3A**) of Yb₂O₃ peaks observed at 236.1 eV (Yb 4d),
 226 533.2 eV (O1s), and f-CNF at 282.5 eV (C 1s), 532.1 eV (O 1s), respectively. The Yb₂O₃/f-CNF
 227 nanocomposite shown in **Fig. 3A**, which exhibits the presence of Yb 4d, C1s, and O1s, is located
 228 at 189.6, 285.6, 533.2 eV, respectively, with minor distortion on the position. **Fig. 3B** shows the
 229 Yb 4d XPS spectrum of Yb₂O₃ at 188.5 eV, which corresponds to Yb³⁺ [60][61], and in Yb₂O₃/f-

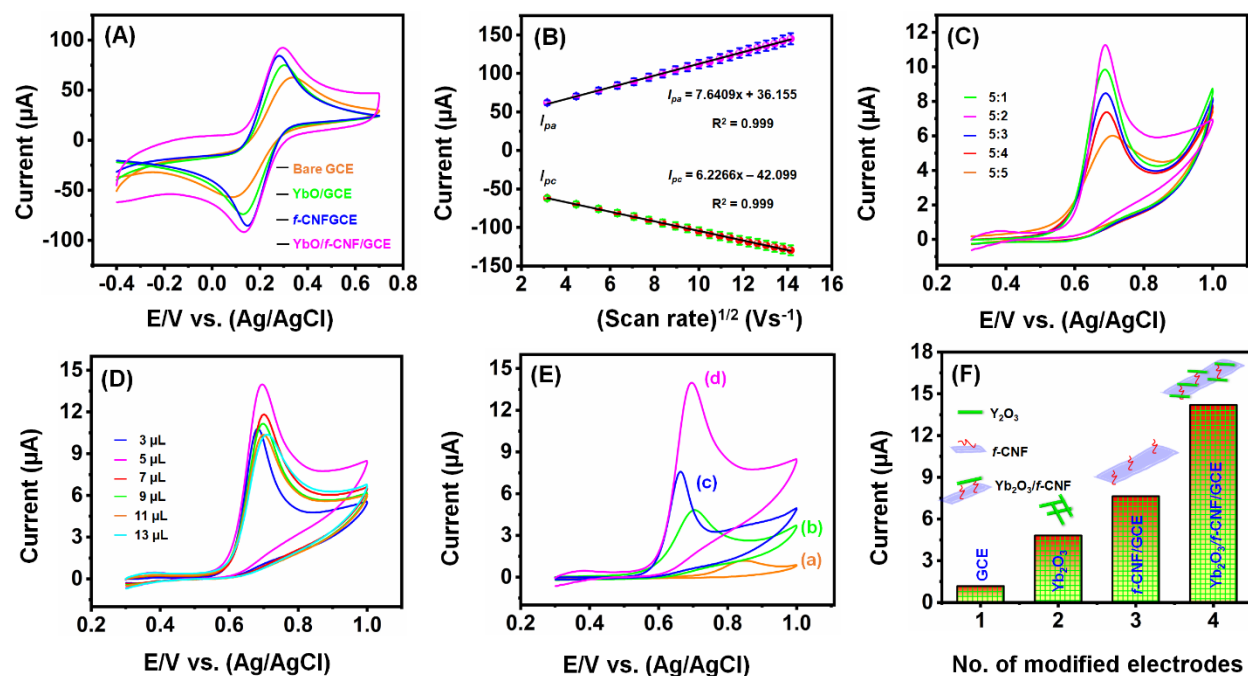
230 CNF nanocomposite, the peak moved around +1.2 eV due to carbon-metal interactions. The
 231 individual C1s spectrum in **Fig. 3C** contains the four different oxygen peaks with binding energies
 232 at 290.2, 286.3, 284.8, and 284.3 eV. These are relevant to the functional groups CO₂H, C=O/C-
 233 O, and C-C and C=C, respectively. The same peaks were also obtained on Yb₂O₃/*f*-CNF
 234 nanocomposite with +0.21 eV positive shift and reduced from the original intensity [62], [63]. In
 235 Fig. 3D, the two significant O1s peaks are attributed to lattice oxygen (534.0 eV) and metal-oxygen
 236 bond (531.3 eV) on Yb₂O₃. The *f*-CNF showed three significant peaks at 531.2, 532.5, and 534.2
 237 eV, which correspond to the oxygen double bond (O=C-O/COO-), a hydroxyl group (C-OH) or
 238 C-O functional group, and oxygen single bond (O=C-O), respectively. The intensity of these peaks
 239 was reduced due to metal attracted to the oxygen functional groups [62][64]. As a result of the
 240 combination of Yb₂O₃/*f*-CNF nanocomposite, there are more active sites for electrochemical
 241 reactions, which enhances electrocatalytic activity.



243 **Fig. 3.** The overall survey spectra for Yb₂O₃, *f*-CNF, and Yb₂O₃/*f*-CNF nanocomposite. Individual
 244 high-resolution XPS spectra of (A) Yb 4d, (B) C1s, and (C) O 1s.

245 4. Electrochemical analysis

246 4.1. Electrocatalytic performance of the modified electrode



247
 248 **Fig. 4.** (A) CV response of bare GCE, Yb₂O₃/GCE, f-CNF/GCE, and Yb₂O₃/f-CNF/GCE in 5 mM
 249 of [Fe(CN)₆]^{3-/4-} comprises of 0.1 M of KCl at a scan rate of 50 mVs⁻¹, (B) The linear relationship
 250 between the square root of scan rate (mVs⁻¹) vs. anodic and cathodic peak current (I_{pa} and I_{pc}). (C)
 251 Different ratios of catalyst optimized in the presence of 200 μM CBZ. (D) Various loading
 252 amounts of catalyst optimized in the presence of 300 μM CBZ (E) CV curves of different
 253 electrodes at scan rate 50 mVs⁻¹ presence of 300 μM CBZ, (F) Bar chart of different individual
 254 electrodes current response for the detection of 300 μM CBZ.

255 The electron transfer and electroactive surface area of the different electrodes were
 256 evaluated by performing CV. The electrochemical performance of modified and unmodified
 257 electrodes was carried out in a mixed solution of 5 mM [Fe(CN)₆]^{3-/4-} and 0.1 M KCl. **Fig. 4A**
 258 showed that redox peak intensities of the bare GCE display a weak redox peak response due to
 259 weak electron transfer. When GCE was modified with Yb₂O₃/GCE, f-CNF/GCE, and Yb₂O₃/f-
 260 CNF/GCE, its electrochemical performance gradually increased in the redox peak current of
 261 Fe^{II}/Fe^{III}. The redox peak current for Yb₂O₃/f-CNF/GCE was more significant than those of other

262 electrodes, implying that the Yb₂O₃ interacts with a strong π - π bond on *f*-CNF, which may
263 contribute to the higher electrocatalytic activity and electron transfer kinetics of Yb₂O₃/*f*-
264 CNF/GCE. Additionally, the electroactive surface area of the unmodified and modified electrodes
265 was determined using CV in a mixed solution of 5 mM [Fe(CN)₆]^{3-/4-} and 0.1 M KCl solution at
266 different scan rates (10-200 mVs⁻¹), as shown in **Fig. 4B**. The Rendles-Sevick Eqn. (1) [65] was
267 used to calculate the active surface area of the Yb₂O₃/GCE, *f*-CNF/GCE, and Yb₂O₃/*f*-CNF/GCE,
268 and was found to be 0.081, 0.090, and 0.99 cm², respectively.

$$269 \quad I_p = 2.69 \times 10^5 A n^{3/2} D^{1/2} \text{red } C \times V^{1/2} \quad (1)$$

270 Where I_p is the redox peak current (μ A), A is the electroactive surface area (cm²), D is the
271 diffusion coefficient (7.6×10^{-6}), n is the number of electrons ($n=1$), and v is the scan rate (Vs⁻¹).
272 Finally, Yb₂O₃/*f*-CNF/GCE possesses the most electroactive surface area among the other
273 electrodes. Hence, the large surface area of the nanocomposite offers the robust channeling of an
274 electron across the interphase and enhances electrocatalytic performance.

275 4.2. Optimization of conditions

276 In order to optimize the Yb₂O₃/*f*-CNF for electrode modifications, the different weights of
277 *f*-CNF containing Yb₂O₃ composite dispersions were prepared using DI water. The resulting
278 composite drop-coated modified electrodes were analyzed for detection of CBZ (200 μ M) at a
279 scan rate of 50 mVs⁻¹. **Fig. 4C** shows the CV response of Yb₂O₃ with different ratios of *f*-CNF
280 (5:1, 5:2, 5:3, 5:4, 5:5) in which the amount of Yb₂O₃ kept as 5 mg/mL. It can be seen that the CBZ
281 oxidation peak current gradually increased upon increasing the amount of *f*-CNF with Yb₂O₃, and
282 the peak current of CBZ was decreased over 2 mg/mL of *f*-CNF. A thick layer of *f*-CNF may
283 prevent CBZ molecules from accessing binding sites because of excess amounts of *f*-CNF. A 5:2
284 mg ratio of the electrode modifier was found to produce the maximum peak current, so this ratio

285 was chosen for electrode modification. Then, the different casting amounts of above-optimized
286 $\text{Yb}_2\text{O}_3/f\text{-CNF}$ were examined using CV, and the obtained results are shown in **Fig. 4D**. The CBZ
287 oxidation peak current increased remarkably when the catalyst loading was increased. The
288 significant oxidation peak current response of CBZ was observed for the 5 μL suspension drop-
289 coated composite electrode. Thus, the above-optimized ratio and drop coating amount of $\text{Yb}_2\text{O}_3/f\text{-}$
290 CNF were used for the entire electrochemical experiment.

291 The electrochemical response of CBZ (200 μM) at different electrodes was investigated by
292 CV in pH 7.0 at a scan rate of 50 mV s^{-1} with a broad oxidation peak potential (E_{pa}) window (0.3
293 to 1.0 V). In GCE, a low oxidation peak current (I_{pa}) and high oxidation potential (0.83V) was
294 noticed due to the slower electron transfer of bare GCE. Oxidation peak current and peak potential
295 in the other electrodes gradually increased. The modified electrodes are arranged in increasing
296 order of $\text{Yb}_2\text{O}_3/\text{GCE}$ ($I_{\text{pa}}=4.8 \mu\text{A}$, $E_{\text{pa}} = 0.70 \text{ V}$) > $f\text{-CNF}/\text{GCE}$ ($I_{\text{pa}}=7.6 \mu\text{A}$, $E_{\text{pa}} = 0.66 \text{ V}$) > $\text{Yb}_2\text{O}_3/f\text{-}$
297 CNF/GCE ($I_{\text{pa}}=13.9 \mu\text{A}$, $E_{\text{pa}} = 0.69 \text{ V}$), as shown in **Fig. 4E** and **4F**. $\text{Yb}_2\text{O}_3/f\text{-CNF}/\text{GCE}$ exhibited
298 a higher oxidation peak current and lower peak potential due to the synergistic effect of Yb_2O_3
299 nanorods and $f\text{-CNF}$. The synergistic effect of $\text{Yb}_2\text{O}_3/f\text{-CNF}$ composite results in the enhanced
300 electron transfer between the electrode surface and analyte (CBZ) than bare, Yb_2O_3 , and $f\text{-CNF}$
301 modified GCEs. Moreover, Yb_2O_3 over the surface of $f\text{-CNF}$ bonded through $\pi\text{-}\pi$ interaction, and
302 its synergistic effect improved the electrocatalytic activity toward the CBZ oxidation. No reduction
303 peak was attained during the reverse scan, which proved the irreversible reaction process and only
304 happening oxidation on the modified electrode surface. Besides, the anodic peak represents the
305 oxidation of the double bond of carbamate in carbendazim to form an amide and heterocyclic
306 radical. Likewise, the oxidation of CBZ on the $\text{Yb}_2\text{O}_3/f\text{-CNF}/\text{GCE}$ surface involves two electrons
307 and two protons (**Scheme 2**).

308 4.3. CBZ concentrations and scan rates

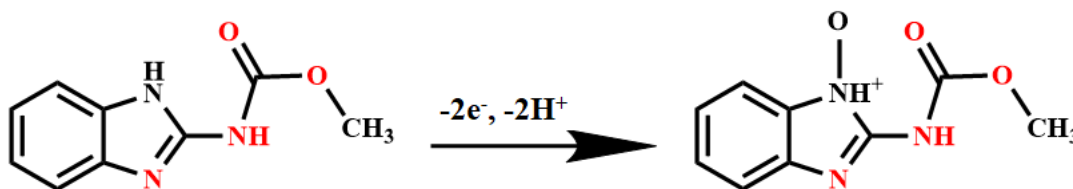
309 The effect of various scan rates was investigated in the range from 10 – 100 mVs⁻¹ for CBZ
310 (200 μM) detection at Yb₂O₃/f-CNF/GCE in pH 7.0. The I_{pa} of CBZ increased gradually with the
311 increasing scan rates, and the peak current was proportional to the scan rate in the potential range
312 (Fig. 5A). In Fig. 5B, the relationship plot between scan rates (mVs⁻¹) and I_{pa} was described by
313 the equation of I_{pa} = 0.0927v + 6.9951 (R²=0.991). Also, the linear relationship expressed for the
314 I_{pa} vs. square root of scan rate (mVs⁻¹) equation is I_{pa} = 0.0706v (mVs⁻¹) + 3.1377 (R² = 0.98),
315 which suggests that the electrochemical reaction of CBZ at Yb₂O₃/f-CNF/GCE followed the
316 surface adsorption-controlled process. The linear relationship of E_{pa} vs. log scan rates expressed
317 in the linear equation is E_{pa} (V) = 0.5839 – 0.5632 (R²=0.999). The Butler-Volmer Eqn. (2) [65]
318 was used to calculate the number of electrons participating in the electrochemical reaction during
319 the CBZ oxidation at Yb₂O₃/f-CNF/GCE.

$$320 \quad E_{pa} = E^{\circ} - \left(\frac{RT}{\alpha nF}\right) \ln \left(\frac{RT}{\alpha nF} \frac{K_s}{\alpha nF}\right) + \left(\frac{RT}{2} \alpha nF\right) \ln v \quad (2)$$

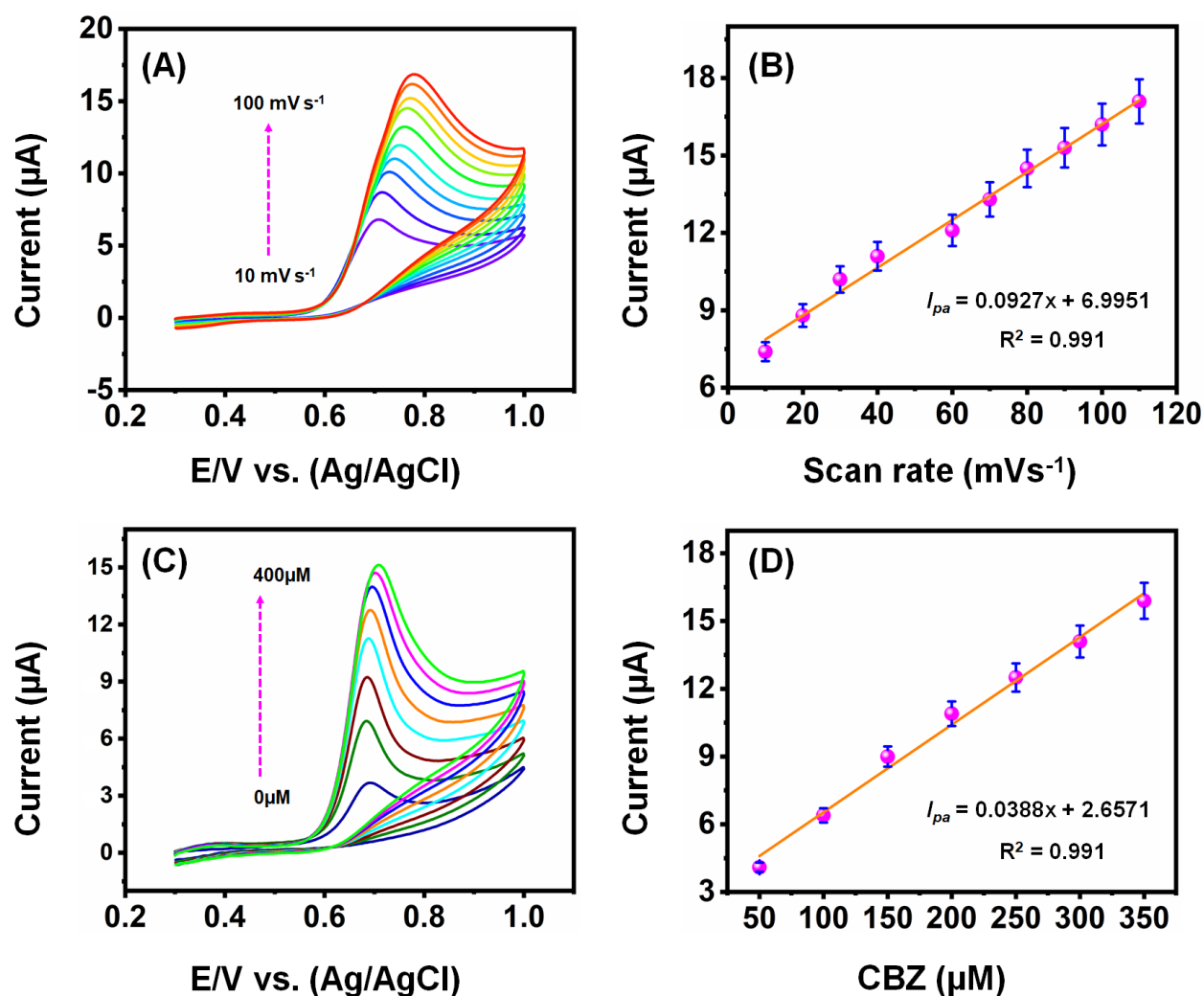
321 Where E^o is the standard potential, α is the electron transfer coefficient, R is the gas constant, T is
322 the surrounding temperature, F is the Faraday constant, and v is the scan rate. The value of α n is
323 1.03, calculated from the obtained slope value of 0.5632, and the value of α is 0.5. The calculated
324 value is 1.98 and is close to 2; thus, two protons and two electrons participated during the CBZ
325 oxidation at Yb₂O₃/f-CNF/GCE [69], [70].

326 The CBZ was quantitatively determined through voltammograms obtained with CV at various
327 concentration range from 50 to 400 μM at a scan rate of 50 mVs⁻¹, as shown in Fig. 5C. When the
328 concentration of CBZ increased linearly, the oxidation peak current also increased equally without
329 change its oxidation potential (0.69 V). This indicates the high stability of the electrode at high
330 concentrations, with more readily available active sites. In Fig. 5D, the oxidation peak current and

331 various concentration of CBZ expressed in the linear regression equation as $I_{pa} = 0.0388_{CBZ} +$
 332 $2.6571(R^2 = 0.991)$. These results indicate the excellent kinetics and higher electrocatalytic activity
 333 of $Yb_2O_3/f-CNF/GCE$ towards CBZ.



335 **Scheme 2.** The possible electrochemical oxidation of CBZ on $Yb_2O_3/f-CNF/GCE$.



336

337 **Fig. 5.** (A) CV response of CBZ (200 μM) at $Yb_2O_3/f-CNF/GCE$ with different scan rates (10–100
 338 mVs^{-1}). (B) The linear relationship between the different scan rates vs. I_{pa} . (C) CV response for

339 different concentrations addition of CBZ (0-400 μM) at $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ at a scan rate of 50
340 mVs^{-1} . (D) The linear relationship between the $[\text{CBZ}]$ vs. I_{pa} .

341 4.4. Influence of pH

342 The pH for CBZ sensing at $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ was investigated using different pHs from 3.0 –
343 11.0 at a scan rate of 50 mVs^{-1} (**Fig. 6A**). The I_{pa} and E_{pa} of CBZ have been moved towards the
344 negative region when varying the pH of an electrolyte from the low to high, suggesting that the
345 protons are taking part in the electrode reaction process. The maximum oxidation current of CBZ
346 was observed at pH 7.0 and was selected as the optimal pH for this study. In **Fig. 6B** revealed the
347 relationship between the different pH vs. E_{pa} can be found as $E_{\text{pa}} (\text{V}) = -0.054 \text{ pH} + 0.829$ ($R^2 =$
348 0.994). The obtained slop value of 54 mV/pH was compared with the Nernst theoretical value
349 (**Eqn. 3**). The obtained value is close to the theoretical value of 59 mV/pH , indicating the equal
350 ratio ($m/n=1$) of electron and proton ($2e^-$ and 2H^+) involved in the electrochemical reaction of CBZ
351 oxidation.

$$352 E_{\text{pa}} = E^\circ - \left(\frac{2.303mRT}{nF}\right) \text{pH} = E^\circ - 0.059 \frac{m}{n} \text{pH} \quad (3)$$

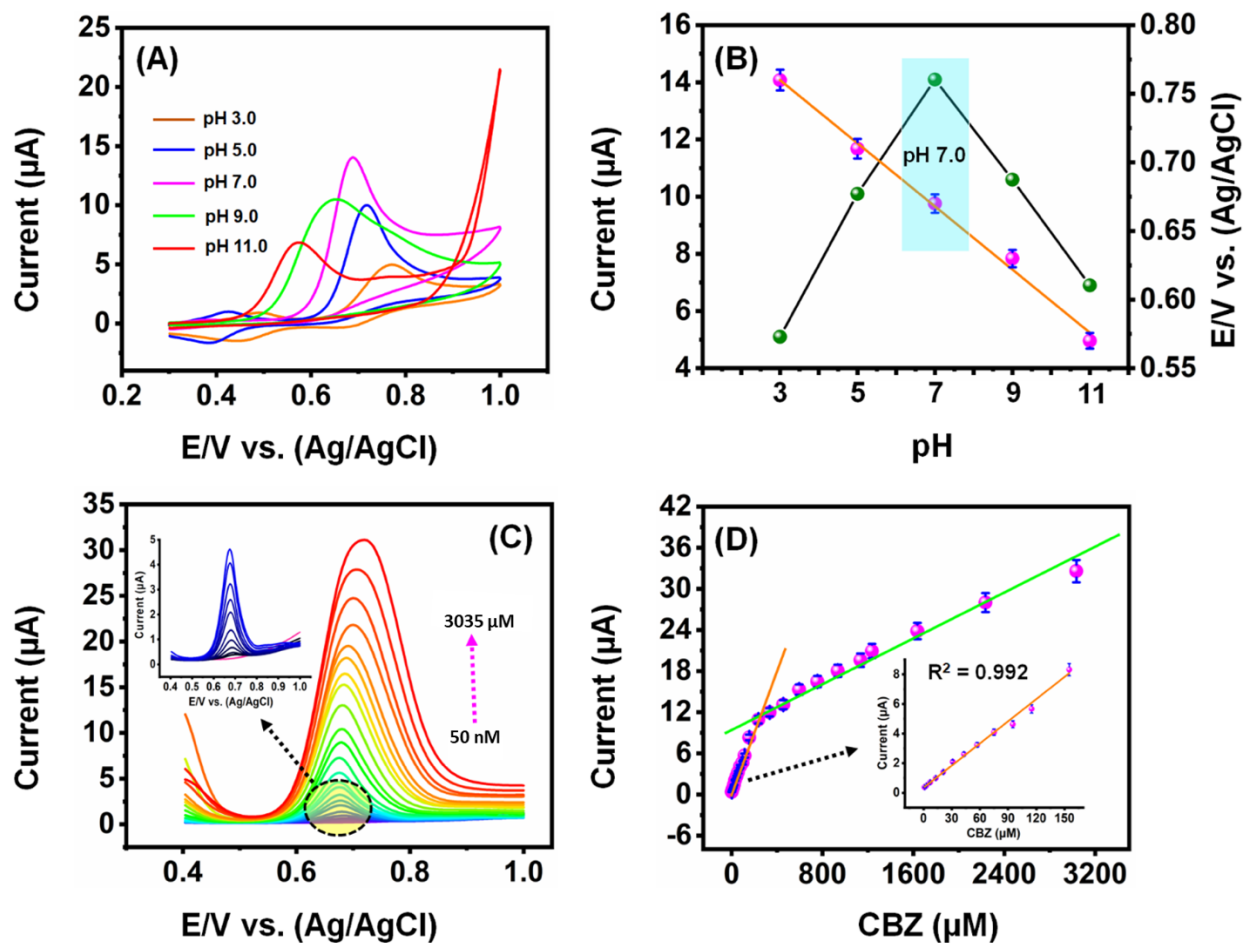
353 4.5. Determination of CBZ on $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$

354 DPV is an ideal electrochemical method for the electrochemical determination of various analytes
355 due to its higher sensitivity, lower detection limit, and lower background current response [66–
356 68]. Therefore, the response current of CBZ at the $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ sensor was explored through
357 the DPV curves in the CBZ concentration range of 50 nM to 3035 μM (**Fig. 6C**). Under the
358 optimized condition, the response current of CBZ presents a significantly enhanced trend with the
359 increase of CBZ concentration. The two linear curves are plotted at low and high concentrations
360 of CBZ. The first linear equation is expressed as $I_{\text{pa}} (\mu\text{A}) = 0.049C_{\text{CBZ}} + 0.3719$ ($R^2 = 0.993$) in the
361 CBZ concentration range of 50 nM-155 μM , as seen in **Fig. 6D**. Second equation is expressed as

362 $I_{pa} (\mu A) = 0.0078C_{CBZ} + 10.227 (R^2 = 0.993)$ in the CBZ concentration range of 156-3035 μM . As
363 a result of the consecutive linear range, when injecting CBZ, the molecules rapidly bind to the
364 electrode surface because free active sites are initially unused. As concentration increases, CBZ
365 curves shift toward a more positive region with a low oxidation current. The oxidation of CBZ at
366 modified $Yb_2O_3/f-CNF/GCE$ surfaces is comparatively slow since there are fewer unoccupied
367 active sites. It was revealed that the occupied sites had a low electroactive area and sensitivity.
368 Therefore, the first linear equation is used to calculate the low detection limit (LOD) by Eqn. (4)
369 [66, 67].

$$LOD = 3S_b/S \quad (4)$$

371 Where S_b is the standard deviation of the blank signal ($S_b=3$), and S is the slope value of the
372 primary linear range. The calculated LOD is 6 nM, and the sensitivity is $0.2899 \mu A \mu M^{-1} cm^{-2}$
373 (Sensitivity = slope value of the curve/surface area of $Yb_2O_3/f-CNF//GCE$), respectively. From
374 this experiment, the obtained results were compared with previously reported literature. The
375 $Yb_2O_3/f-CNF/GCE$ sensor displayed a better LOD, sensitivity, and wide linear range than
376 previously reported many CBZ sensors, as shown in **Table 1**.



377
 378 **Fig. 6.** (A) CV response of $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ for CBZ ($300 \mu\text{M}$) in different pH (pH 3.0 – 7.0)
 379 at scan rate 50 mVs^{-1} . (B) The linear relationship between the pH vs. I_{pa} and E_{pa} . (C) DPV response
 380 of the $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ for successive addition of CBZ from 50 nM to $3035 \mu\text{M}$. (D) The linear
 381 relationship between the CBZ concentrations (μM) vs. current response.

382 **Table 1.** Comparison of sensing performance of the fabricated CBZ sensor with previously
 383 reported various CBZ sensors.

Electrode	Method	Linear range (μM)	Limit of detection (nM)	Refs.
$\text{ZnFe}_2\text{O}_4/\text{SWCNT}/\text{GCE}^{\text{a}}$	DPV ^c	0.5-100	90	[71]
$\text{SiO}_2/\text{MWCNT}/\text{GCE}^{\text{a}}$	SWV ^d	0.2-4.0	56	[72]
$\text{Ti}_3\text{C}_2\text{T}_x\text{MXene}/\text{GCE}^{\text{a}}$	DPV ^c	0.05-100	10.3	[73]

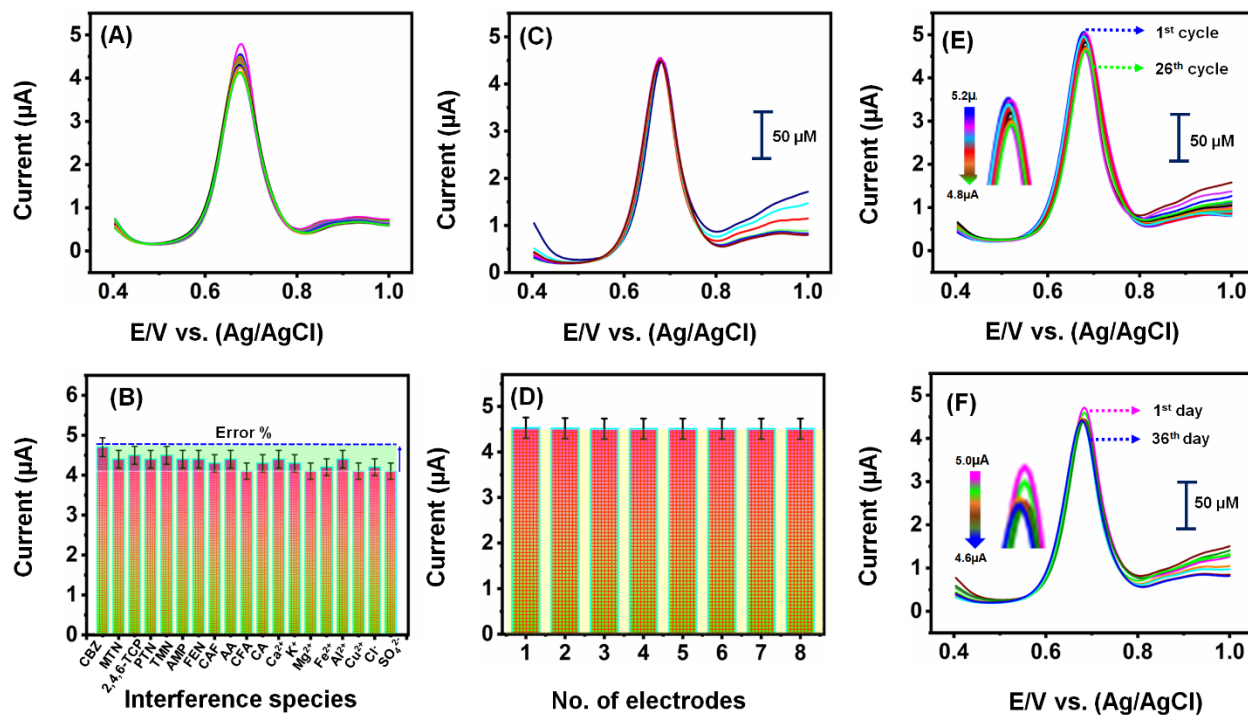
P-HCNFs/GCE ^a	DPV ^c	1-35	38	[74]
NP-Cu/RGO/GCE ^a	DPV ^c	0.5-30	90	[75]
La-Nd ₂ O ₃ /CPE ^b	DPV ^c	0.08-50	27	[76]
D-PC/GCE ^a	SWV ^d	0.01-1.00	6	[7]
Ce-dope ZnWO ₄ /GCE ^a	DPV ^c	0.01-5.5	3	[77]
WO _{3.0.33} H ₂ /GCE ^a	SWV ^d	0.1-250	22	[78]
Fe-CuV/GCE ^a	DPV ^c	0.01-83.1	5	[79]
MBC@CTS/GCE ^a	DPV ^c	0.1-20	20	[22]
QD-rGO/CPE ^a	DPV ^c	0.1-11.8	91.6	[80]
TCP/CPE ^a	DPV ^c	0.5-10	300	[81]
MWCNTs/GCE ^a	SWV ^d	0.256-3.11	54.9	[82]
Yb₂O₃/f-CNF/GCE^a	DPV^c	0.05-155 156-3035	6	This work

384 ^aGCE-Glassy carbon electrode, ^bCPE-Carbon paste electrode, ^cDPV- Differential pulse
385 voltammetry, ^dSWV- Square wave voltammetry,

386 4.6. Selectivity, reproducibility, repeatability, and storage stability

387 It is essential to ensure the electrode's selectivity to use it effectively for practical
388 applications. As shown in **Fig. 7A**, the Yb₂O₃/f-CNF/GCE response current for CBZ (at 50 mVs⁻¹
389 scan rate) was not affected in the presence of excessive amounts of interfering compounds (like
390 pesticides, biological compounds, and metal ions, including amatryn, parathion, thiamethoxam,
391 2,4,6-trichlorophenol, carbofuran, calcium, potassium, magnesium, iron, aluminum, copper,
392 chlorine, and sulfate). The RSD was calculated as 3.66 ± 0.158. The above result confirms the
393 highly selective nature of Yb₂O₃/f-CNF/GCE toward CBZ. The relative error bar is shown in **Fig.**
394 **7B**. The obtained results suggest that the CBZ oxidation peak current was not affected by the
395 presence of the above-mentioned interfering species. Hence, the fabricated sensor electrode can
396 be used for real-time CBZ monitoring. The reproducibility, repeatability, and storage stability of
397 the modified Yb₂O₃/f-CNF/GCE sensor were examined for the detection of CBZ (50 μM). Eight
398 different Yb₂O₃/f-CNF/GCEs were prepared and evolved using DPV (Fig. 7C-D) for the

399 reproducibility studies. The current response of the eight different electrodes had the same
 400 potential as the peak current of CBZ. From these DPV curves, the calculated RSD was 4.51 ± 0.51 .
 401 Then repeatability of the $\text{Yb}_2\text{O}_3/f\text{-CNF}/\text{GCE}$ sensor was evaluated with the same experimental
 402 conditions as mentioned above (**Fig. 7E**). In this experiment, 26 cycles were continuously
 403 operated, and RSD was calculated as 4.77 ± 2.63 . Even after 26 cycles, the CBZ oxidation peak
 404 current was retained ($-0.4 \mu\text{A}$) from the initial value, indicating the high repeatability of the
 405 sensor. In addition, the long-term stability (**Fig. 7F**) of the modified $\text{Yb}_2\text{O}_3/f\text{-CNF}/\text{GCE}$ was
 406 examined at intervals of 4 days, with the electrode stored in a refrigerator at 4°C when not in
 407 use, for detection of CBZ ($50 \mu\text{M}$, 0.01M). The peak current value is still 96% of the original value
 408 (maximized in **Fig. 7F**) after 36 days, proving the electrode's long-term stability. According to the
 409 above results, the modified $\text{Yb}_2\text{O}_3/f\text{-CNF}/\text{GCE}$ sensor was reproducible, repeatable, and stable
 410 toward the detection of CBZ.

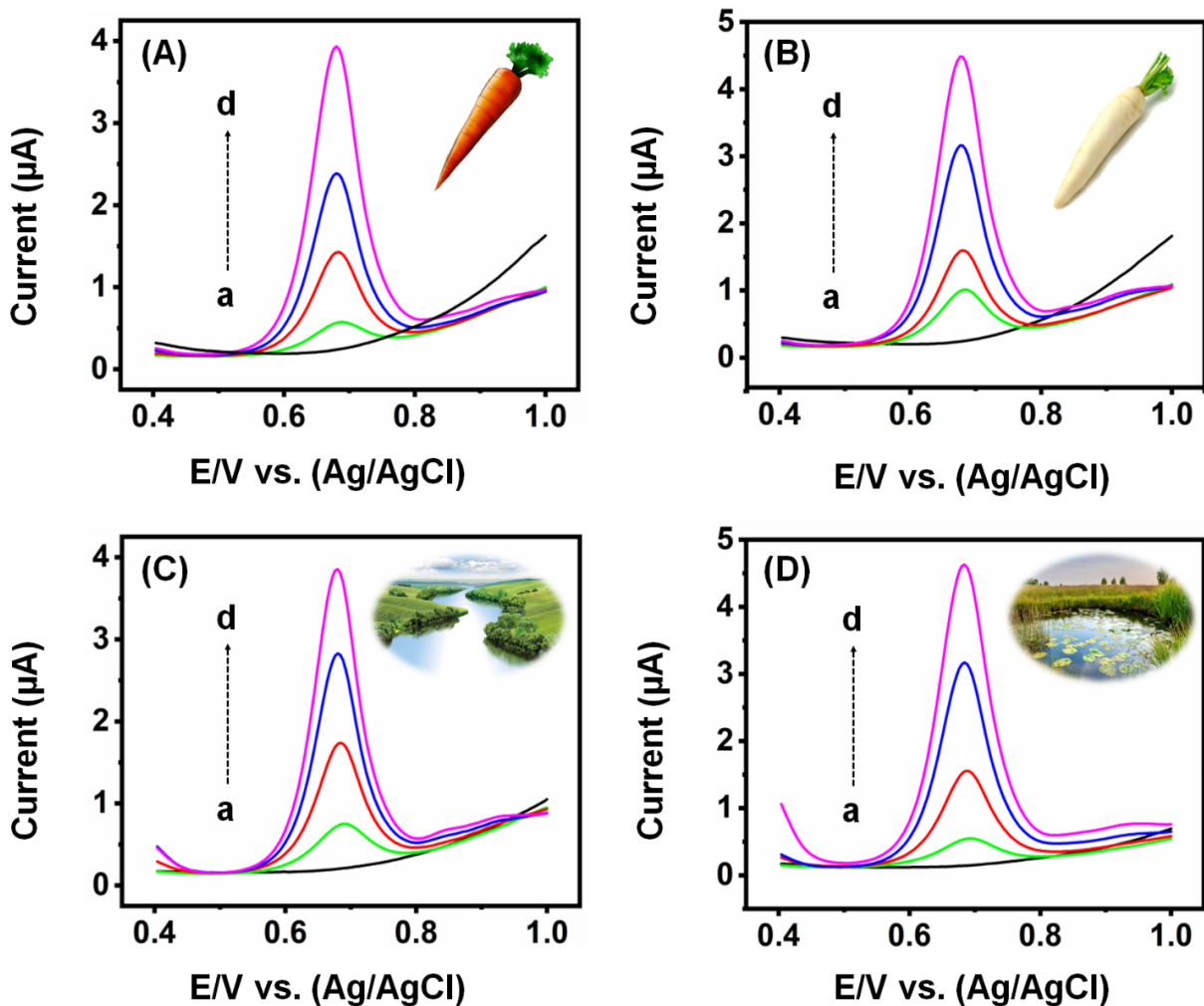


411

412 **Fig. 7.** (A-B) Selectivity & error bar diagram, (C-D) Reproducibility & bar diagram, (E)
413 Repeatability, and (F) Long-term stability of Yb₂O₃/f-CNF/GCE toward CBZ.

414 **4.7. Real-time application**

415 The real-time detection capability of the proposed CBZ sensor has been evaluated in food and the
416 environment to use it in practical applications. The real samples of carrot, radish, lake water, and
417 pond water were used to evaluate the practical feasibility of the Yb₂O₃/f-CNF composite. The
418 vegetables were purchased from a local market in Taiwan. The vegetable samples were peeled, cut
419 into small pieces, and crushed with an electric blender before analysis. After collecting the extract,
420 it was centrifuged for 15 minutes at 3000 rpm. First, the 1 mL of separated supernatant extract was
421 diluted with 9 mL DI water. The pH was adjusted to pH 7.0. The real samples in the absence and
422 presence of different concentrations of CBZ were studied using DPV. The obtained results are
423 shown in **Fig. 8A-D**. It can be seen that the carrot (A), radish (B), lake water (C), and pond water
424 (D) real samples did not show any DPV response in the absence of CBZ, which indicates that these
425 real samples are CBZ-free. A sharp DPV response was observed for real samples containing CBZ
426 (5 μM), and the DPV response current increased with an increasing CBZ concentration (10, 15,
427 and 20 μM). The standard addition method was used to calculate the recovery of CBZ in the real
428 sample, and the obtained recovery values are listed in **Table 2**. The recovery rate of the sensor
429 ranged from 96.0% to 99.4%, with an acceptable relative standard deviation (RSD). According to
430 the above results, the detection method based on the Yb₂O₃/f-CNF composite electrode can be
431 used to determine CBZ presence.



432

433 **Fig. 8.** DPV response of CBZ for the addition of 5, 10, 15, and 20 μM at $\text{Yb}_2\text{O}_3/\text{f-CNF}/\text{GCE}$ in
 434 different real-time samples of (A) carrot, (B) radish, (C) lake water, and (D) pond water.

435

436 **Table 2.** Real-time determination of CBZ in food and water sample.

Sample	Added (μM)	Found (μM)	Recovery	RSD (%)
Carrot	5	4.80	96.07	1.00 \pm 1.69
	10	9.81	98.10	1.56 \pm 0.79
	15	14.66	97.79	3.19 \pm 2.32
	20	19.68	98.43	4.42 \pm 0.87
Pond water	5	3.85	96.49	0.56 \pm 1.45

Radish	10	9.79	97.91	1.40±2.35
	15	14.55	97.05	2.32±1.65
	20	19.79	98.98	3.88±1.39
Lake water	5	4.90	98.18	0.52±4.98
	10	9.61	98.08	1.51±3.97
	15	14.71	98.10	3.12±1.08
Pond water	20	19.78	98.92	4.57±1.26
	5	4.86	98.64	0.73±2.86
	10	9.94	99.42	1.72±0.72
	15	14.73	98.23	2.80±0.93
	20	19.75	98.70	3.81±0.86

437

438 5. Conclusion

439 S novel hybrid nanocomposite of Yb₂O₃/f-CNF has been prepared for the electrochemical
440 detection of CBZ in vegetables and fruit samples. The morphological and structural characteristics
441 of the developed Yb₂O₃/f-CNF nanocomposite were examined using various analytical and
442 spectroscopic methods. The Yb₂O₃/f-CNF nanocomposite-modified GCE demonstrated a low
443 LOD, a broad linear range, and relatively high sensitivity for the detection of CBZ. Additionally,
444 the sensor exhibited high selectivity, stability, and anti-interference ability toward CBZ sensing,
445 and showed satisfactory identification of CBZ in vegetables and water samples. The fabricated
446 sensor could potentially be used to detect CBZ in real time.

447 Declaration of Competing Interest

448 The authors declare that they have no competing financial interests or personal relationships
449 relevant to this paper.

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453

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