

Evaluation of Prebiotic and Probiotic Effects on the Intestinal Gut Microbiota and Histology of Atlantic salmon (*Salmo salar* L.)

Mads Kristiansen^{1,2}, Daniel L. Merrifield³, Jose L. Gonzalez Vecino⁴, Reidar Myklebust⁵ and Einar Ringø^{1*}

¹Norwegian College of Fishery Science, Faculty of Bioscience, Fisheries and Economics, University of Tromsø, Brevikva, Norway.

²Present address. Veso Vikan, Aquamedical Contract Research, Vikan, N-7800 Namsos, Norway.

³Aquaculture and Fish Nutrition Research Group, School of Biomedical and Biological Sciences, The University of Plymouth, UK.

⁴EWOS Innovation AS, Dirdal, Norway.

⁵Molecular Imaging Centre (MIC), Institute for Biomedicine, University of Bergen, Bergen, Norway.

Abstract

In the present study the impact on gut microbiology and indigenous gut histology of Atlantic salmon (*Salmo salar* L.) was investigated following feeding of a control and a prebiotic (EWOS prebiosal®) diet and *ex vivo* exposure to saline or the probiotic bacterium *Carnobacterium divergens*. The results showed that *ex vivo* exposure of *C. divergens* at 10⁸ CFU ml⁻¹ did not cause cell damage to the intestine tract of Atlantic salmon. Furthermore, prior provision of dietary prebiotic elevated the ability of *C. divergens* to adhere to the epithelium or mucus layer in the proximal intestine, where culturable heterotrophic bacterial levels (which were identified as *C. divergens*) were elevated by 234% compared to the control. This effect was not apparent in the distal intestine. The ability of isolated carnobacteria from the *ex vivo* experiment to inhibit growth of two fish pathogenic bacteria (*Yersinia ruckeri* and *Aeromonas salmonicida* ssp. *salmonicida*) was assessed. Extracellular products from all 11 of the isolated carnobacteria strains, plus the type strain *Carnobacterium inhibens* CCUG 31728, inhibited the *in vitro* growth of *Y. ruckeri*. However, only extracellular products from *C. divergens* isolate 57 inhibited the growth of *A. salmonicida*

Keywords: Carnobacteria; Synbiotic; Electron microscopy; Pathogen; 16S rRNA

Introduction

Today it is generally accepted that the three major routes of infection in fish are through: a) skin, b) gills and c) the gastrointestinal (GI) tract [1]. The GI microbiota, including lactic acid bacteria (LAB), have been suggested to be important in fish health and it has been suggested that the autochthonous gut bacterial community may be responsible for controlling the colonization of potential pathogens by adhesion competition and production of antagonistic compounds [2-7]. If the GI tract is involved as an infection route, scientists have to address whether probiotic bacteria are able to adhere and colonise mucosal surfaces and outcompete endogenous bacteria and pathogens.

Investigating these topics effectively in *in vivo* models can be difficult as they are time consuming and have high cost. Furthermore, as the EU has recommend reductions of *in vivo* experiments and the numbers of animals used in experiments (Revision of the EU directive for the protection of animals used for scientific purposes [Directive 86/609/EEC]; 8th of September 2010), attempts have been made to use alternative *ex vivo* methods (e.g. the Ussing chamber, everted sack and intestinal sack methods) [8-15].

The first aim of the present study was to investigate possible effects of a prebiotic feed on epithelial histology and indigenous GI tract microbiota in the proximal intestine (PI) and distal intestine (DI) of Atlantic salmon. Furthermore, the same effects, including morphological changes of epithelial cells after *ex vivo* exposure of the intestinal tract to *Carnobacterium divergens*, a probiotic bacterium, are investigated by light microscopy and electron microscopy. The result of *Carnobacterium* exposure is of high importance to evaluate as translocation and cell damage are negative criteria when evaluating the use of probiotics in endothermic animals as well as in fish [16,17]. The 2nd aim of the present study was to evaluate the bacterial community of the PI and DI of salmon fed control or prebiotic diets, before and

after *ex vivo* exposure to probiotic bacteria, in order to investigate if the indigenous GI tract microbiota is modulated by the different treatments. Finally, we addressed the issue as to whether carnobacteria isolated in the *ex vivo* studies were able to inhibit *in vitro* growth of the pathogenic bacteria *Yersinia ruckeri* and *Aeromonas salmonicida* ssp. *salmonicida*.

Materials and Methods

Fish husbandry

Two hundred and forty vaccinated Atlantic salmon (*Salmo salar* L.) were held at the EWOS Innovation AS Research Station, Dirdal, Norway. The average weight at the start of the experiment was 350 g. Two hundred and forty fish were distributed equally (i.e. 40 fish per tank) into six tanks supplied with 500 liters of sea water and two diets were offered (i.e. triplicate tanks per diet). The control diet and prebiotic diet had the same ingredient composition (Table 1) and differed only in the inclusion of 0.2% EWOS prebiosal® in the prebiotic diet. EWOS prebiosal®, is described as a multi-component prebiotic specifically designed for salmonid fish; more detailed information about the composition of EWOS prebiosal® is not available for commercial reasons. Feeding was conducted twice a day with duration of 2.5 hour between each feeding for a period of 15 weeks. During the feeding

***Corresponding author:** Einar Ringø, Norwegian College of Fishery Science, Faculty of Bioscience, Fisheries and Economics, University of Tromsø, Brevikva, Norway, E-mail: einarringo@uit.no

Received August 02, 2011; **Accepted** October 15, 2011; **Published** October 25, 2011

Citation: Kristiansen M, Merrifield DL, Vecino JLG, Myklebust R, Ringø E (2011) Evaluation of Prebiotic and Probiotic Effects on the Intestinal Gut Microbiota and Histology of Atlantic salmon (*Salmo salar* L.) J Aquac Res Development S1:009. doi:10.4172/2155-9546.S1-009

Copyright: © 2011 Kristiansen M, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

period the water temperature and salinity ranged, with season, from 5.3-12.9°C and 26.7-30.9 g l⁻¹.

The samplings were carried out at two different points: at the start (week 0) and at the end of the trial (week 15). An overview of the different treatments and groups is listed in Table 2.

Probiotic bacteria

The probiotic bacterium used in this experiment was *Carnobacterium divergens* strain Lab01 originally isolated from juvenile Atlantic salmon fed a commercial diet [18]. The bacteria were stored in glycerol-containing cryotubes at -80°C and inoculated into tryptic soy broth (Difco, USA) with glucose (10g l⁻¹) and NaCl (10g l⁻¹), viz. TSBgs medium. After approximately 24 hours of pre-inoculation at room temperature with an agitation of 190rpm, 1% of the pre-culture was transferred to new TSBgs medium and growth (same growth conditions as above) was measured by optical density for evaluation of the growth cycle (data not shown). Bacterial viability was confirmed by plating bacterial suspensions on tryptic soy agar (Difco) + glucose (15g l⁻¹) and NaCl (15g l⁻¹) (TSAGs) plates. The results obtained from this study were used to calculate the bacterial concentration in the experimental bacterial solutions.

Ex vivo exposure to bacteria

Three fish were randomly selected from two of the tanks fed each diet and killed with a blow to the head. The entire intestine, from the last pyloric caeca to the anus, was removed aseptically and intestinal content was gently squeezed out, before the intestine was flushed three

times with sterile saline solution (0.9 % NaCl), in order to remove the allochthonous gut bacteria. The posterior end was tightly tied with cotton thread before filling (ca. 1.5ml) with the appropriate assay solution (Table 2), tying the anterior end and suspending the sealed intestinal tube in sterile saline solution. The intestinal sacks were then incubated at 10°C for 1 hr.

After incubation the intestine was cut open, the contents discarded and flushed three times with sterile saline solution.

Post ex vivo bacterial assays

Samples for bacteriology from each segment from the first sampling point (groups 1 and 2) were prepared by homogenizing 1 gram of intestinal tissue (PI or DI) in 1 ml sterile saline using a Stomacher (Seaward Laboratory, UK). Gut samples for bacteriology from the second sampling (groups 3-6) were prepared by gently scraping off mucus with a sterile scalpel. Thereafter, the segments were weighed. Both the homogenates and mucus were used to create serial ten-fold dilutions which were spread plated (100µl) on TSAGs plates and incubated at 6°C for up to 1 week to determine viable counts of culturable heterotrophic bacteria.

After sub-culturing on TSAGs to achieve pure cultures, phenotypic bacterial identification (Gram stain, colony morphology, oxidase - and catalase tests and glucose fermentation) was carried out on random colonies from all plates containing between 10-300 colonies. A total of 168 bacterial strains were isolated from the two sampling points.

16S rRNA characterization of isolates

The bacterial DNA was isolated following the protocol from a commercial kit (DNeasy Blood and Tissue, Qiagen, USA). Specific treatment for Gram-positive and Gram-negative isolates was carried out according to the manufacturer's instructions. Template-DNA was diluted to a concentration of approximately 20-30 ng µl⁻¹ using Milli-Q water. The PCR mix constituted of 8µl of template-DNA, 36µl Milli-Q water, 5µl 10x buffer F511, 0.25µl dNTP, 0.25µl 27F forward primer, 0.25µl 1492R reverse primer and 0.25µl DNA-polymerase yielding a total volume of 50µl. PCR thermal cycling consisted of initial denaturation of 94°C, followed by 35 cycles of 94°C for 20 s, 53°C for 20 s and 72°C for 90 s with a final extension step of 72°C for 7 min. To verify PCR products, samples were run on gel electrophoresis. The PCR-products were desalted by mixing 20µl of PCR-product with 40µl of 100% ethanol and 2µl of 3M NaOAc (pH 5.3) and vortexed well. Samples were then incubated on ice for 30 min followed by centrifugation for 20 minutes at 14,000 g using an Eppendorf Microcentrifuge Model 5417R. The supernatant was removed and pellet washed in 100µl of 80% ethanol and centrifuged for another 5 min at 14,000 g. The supernatant was removed and the pellet dried at room temperature for 60 min. The pellet was then resuspended in 30µl of Milli-Q water. Purified PCR products were sequenced as described elsewhere [19]. The resultant nucleotide sequences were submitted to a BLAST search in GenBank (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) to retrieve the closest known alignment identities for the partial 16S rRNA sequences. Gene sequences that showed higher than 95% similarity to a genus or species in GenBank were categorized accordingly.

In vitro growth inhibition of pathogens by LAB isolated from the ex vivo studies

Eleven randomly chosen LAB isolated from the intestinal tract after ex vivo exposure and one type strain, *Carnobacterium inhibens* (CCUG 31728), were tested for antagonistic effects against two different fish

| Ingredient (%) | |
|---|-------|
| Fish meal | 31.25 |
| North Atlantic fish oil | 13.50 |
| Vegetable protein concentrates ¹ | 25.76 |
| Vegetable oil | 14.01 |
| Carbohydrate-based binders ² | 13.00 |
| Micro premixes ³ | 2.48 |
| Chemical composition (%) | |
| Moisture | 6.9 |
| Protein ⁴ | 44.2 |
| Fat ⁴ | 29.1 |
| NFE ⁴ | 1.6 |
| Ash ⁴ | 8.4 |

¹Includes soy protein concentrate, pea protein concentrate, wheat gluten, sun-flower meal.

²Includes wheat and pea starch

³Includes vitamin, mineral, amino acid and pigment premixes and 0.2 % EWOS prebiosaal® added to the prebiotic diet (at the expense of an equal volume of carbohydrate-based binders)

⁴dry weight basis

Table 1: Dietary formulation and chemical composition of the experimental diets.

| Treatment group | Type of treatment | Type of feed | Week of feeding |
|-----------------|----------------------------------|--------------|-----------------|
| 1 | Saline | Control | 0 |
| 2 | <i>C. divergens</i> ¹ | Control | 0 |
| 3 | Saline | Control | 15 |
| 4 | <i>C. divergens</i> ² | Control | 15 |
| 5 | Saline | Prebiotic | 15 |
| 6 | <i>C. divergens</i> ² | Prebiotic | 15 |

¹10⁶ CFU ml⁻¹

²10⁸ CFU ml⁻¹

Table 2: Experimental treatments applied to Atlantic salmon intestine fed control and prebiotic diets.

pathogens. The pathogenic bacteria used in the present investigation were *Yersinia ruckeri* (CCUG 14190) and *Aeromonas salmonicida* ssp. *salmonicida* (Ass 4017). *C. inhibens* (CCUG 31728) was used as a positive control as previous investigations have demonstrated that this strain has an inhibiting effect towards *V. anguillarum* and *A. salmonicida* [19,20]. *In vitro* growth inhibition of the two fish pathogens by the twelve LAB was tested using a microtitre plate assay described in detail by Ringø and co-authors [3]. This method has been used in two recent studies [15,22]. The pathogenic bacterial levels at the start of assays were 10^6 cells ml^{-1} . Positive *in vitro* growth inhibition was defined

| Group | Proximal intestine | | | Distal intestine | | |
|-------|--------------------------------|----|--|--------------------------------|----|---|
| | TVC (log CFU g^{-1}) | N* | Bacteria % | TVC (log CFU g^{-1}) | N* | Bacteria % |
| 1 | 1.72 | 12 | <i>Psychrobacter aquimaris</i> – 16.7 % <i>Psychrobacter glancincola</i> – 16.7 % <i>Psychrobacter</i> spp. – 66.6 % | 1.73 | 11 | <i>Psychrobacter glancincola</i> – 9.0 % <i>Psychrobacter</i> spp. – 36.3 % <i>Pseudoalteromonas</i> – 36.3 % <i>Brevibacterium</i> sp. – 9.0 % <i>Moraxella</i> sp. – 9.0 % |
| 2 | 6.04 | 7 | <i>Carnobacterium divergens</i> – 100 % | 5.56 | 7 | <i>Carnobacterium divergens</i> – 100 % |
| 3 | 2.08 | 17 | <i>Carnobacterium divergens</i> – 70.6 % <i>Pseudomonas fluva</i> – 17.6 % <i>Pantoea</i> spp. – 5.9 % <i>Gammaproteobacteria</i> – 5.9 % | 2.69 | 15 | <i>Carnobacterium divergens</i> – 33.3 % <i>Pseudomonas fluva</i> – 6.6 % <i>Shewanella baltica</i> – 6.6 % <i>Vibrio splendidus</i> – 13.3 % <i>Gammaproteobacteria</i> – 40 % |
| 4 | 6.26 | 8 | <i>Carnobacterium divergens</i> – 87.5 % <i>Pseudomonas</i> spp. – 12.5 % | 6.68 | 8 | <i>Carnobacterium divergens</i> – 100 % |
| 5 | 2.34 | 47 | <i>Carnobacterium divergens</i> – 29.8 % <i>Carnobacterium</i> spp. – 51 % <i>Pseudomonas antarctica</i> – 2.1 % <i>Pseudomonas koreensis</i> – 2.1 % <i>Enterobacter hormaechi</i> – 8.5 % <i>Gammaproteobacteria</i> – 4.3 % Uncultured bacterial clone CK20 – 2.1 % | 1.71 | 44 | <i>Carnobacterium divergens</i> – 25 % <i>Carnobacterium</i> spp. – 52.3 % <i>Pantoea</i> spp. – 18.2 % <i>Enterobacter</i> spp. – 4.5 % |
| 6 | 6.63 | 25 | <i>Psychrobacter marincola</i> – 4 % <i>Pseudomonas</i> sp. – 8 % <i>Carnobacterium divergens</i> – 20 % <i>Carnobacterium</i> spp. – 68 % | 6.7 | 17 | <i>Acinetobacter</i> sp. – 5.6 % <i>Carnobacterium divergens</i> – 94.2 % |

* N = number of isolates identified

Table 3: Culturable heterotrophic bacterial levels (log CFU g^{-1} wet weight) and identity (as determined from phenotypic characteristics and 16S rRNA sequence analysis) obtained from different groups after the *ex vivo* assay.

when no growth (turbidity < 0.05 at optical density; OD_{600} nm) of the pathogen was detected. Sterile growth media and the pathogens were used as controls. Growth (at OD_{600}) of the pathogens without addition of sterile supernatant of LAB was approximately 0.6. Measurements were carried out each hour using an automatic plate reader (Bioscreen C, LabSystems, Finland).

Histology

Samples for light microscopy (LM) and transmission electron microscopy (TEM) were collected by excising approximately 5mm from the posterior part of the PI and DI. The samples were immediately fixed in McDowell's fixative [23] and stored at 4°C until processing. TEM and LM samples were processed as described elsewhere [9,24]. Morphological observations were made from multiple micrographs (8) from each intestinal region from two fish within each group. The following morphological parameters were observed; detached microvilli, enterocytes detached from the basal membrane, disintegrated cell junctions, presence of goblet cells, presence of absorptive vacuoles and presence of intraepithelial lymphocytes.

Results

Bacterial levels after *ex vivo* exposure

The adherent bacterial levels, as determined by using a stomacher (groups 1 and 2) or by the collection of mucus (and subsequent weighing of the segments the mucus was removed from) (groups 3 – 6), did not seem to differ which indicates that the different sampling methods were similarly effective. Table 3 presents an overview of the autochthonous bacterial levels isolated from each segment and each group exposed to either saline or *C. divergens*. All values are expressed as log colony forming units (CFU) g^{-1} . Autochthonous bacteria isolated from intestines of fish fed the control diet at the experimental start and exposed to saline was approximately log 1.7 CFU g^{-1} in both PI and DI, while the number of bacteria isolated from intestines of fish exposed to *C. divergens* was log 6.04 CFU g^{-1} in PI and log 5.56 CFU g^{-1} in DI. After 15 weeks of feeding slightly higher values were present in PI of fish fed the prebiotic diet post exposure to saline or *C. divergens* compared to fish fed the control diet. Indeed, the bacterial level in the prebiotic fed fish intestine exposed to *C. divergens* (group 6) was 234% greater than that of the control fed fish intestine exposed to *C. divergens* (group 4). In both dietary groups, a similar bacterial level ($\sim \log 6.70$ CFU g^{-1}) was detected in DI exposed to *C. divergens*. However, a higher bacterial level (log 2.69 CFU g^{-1}) was observed in the DI of control fed fish exposed to saline than that of prebiotic fed fish (log 1.71 CFU g^{-1}).

Isolation and identification of bacteria after *ex vivo* exposure

A total of 168 bacterial strains were isolated from the two samplings. Among these, 40 isolates were isolated from the first sampling point and 128 isolates were isolated from the second sampling point. All isolates were tested for morphology and biochemical properties (colony morphology, Gram-testing, oxidase - and catalase tests and glucose fermentation).

One hundred and eleven isolates were further identified by partial sequencing of the 16S rRNA gene. Isolates not identified by 16S rRNA gene sequencing but showing similar biochemical and physiological properties to those isolates identified by 16S rRNA genes were defined as “- like”. Table 3 provides an overview of the different bacterial species isolated in each experimental group.

Week 0 (experimental start)

Microbiota of fish fed control diet and intestines exposed to saline (group 1): Analysis of the adherent microbiota in the PI of fish fed the control diet and exposed to sterile saline (group 1) revealed that all isolates belonged to the genus *Psychrobacter*. Of the 12 strains isolated from the PI of this group, 2 strains showed 96 % similarity to *Psychrobacter aquimaris*, 2 strains were identified as *Psychrobacter glacincola* while 8 strains were identified as *Psychrobacter* spp. - like. The DI of fish exposed to saline at the first sampling point showed a more diverse community which consisted of 4 different bacterial genera. Of these, 10 strains were indentified to genus level and 1 strain was identified to species level. The bacteria identified to genus level belonged to *Pseudoalteromonas*, *Psychrobacter*, *Moraxella* and *Brevibacterium*, while the last strains showed high similarity (98 %) to *Psychrobacter glacincola*.

Microbiota of fish fed control diet and intestines exposed to *C. divergens* (group 2): All bacteria isolated from PI and DI of fish exposed to *C. divergens* at the first sampling (group 2) were identified as *C. divergens*. This observation indicates that *C. divergens* are able to adhere to the intestinal mucosa in both segments.

Week 15

Microbiota of fish fed control diet and intestines exposed to saline (group 3): After 15 weeks of feeding on the control diet, the isolated strains (17) from the PI exposed to saline were dominated by *C. divergens*; 70.6% of the isolates were identified as *C. divergens*, 17.6% were identified as *Pseudomonas fulva*, 5.9% belonged to *Pantoea* spp. while 5.9% of the isolates were identified as members of the class Gammaproteobacteria. The bacteria isolated from the DI were identified as *C. divergens*, 2 strains as *Vibrio splendidus*, 1 strain as *Shewanella baltica*, 1 strain as *Pseudomonas fulva* and 6 other strains were identified as Gammaproteobacteria.

Microbiota of fish fed control diet and intestines exposed to *C. divergens* (group 4): In the intestine of fish fed the control diet for 15 weeks and exposed to *C. divergens*, the identified bacterial strains isolated from both PI and DI were dominated by *C. divergens*. Only 1 strain, identified as *Pseudomonas* spp., isolated from the PI of 1 fish did not belong to the species *C. divergens*.

Microbiota of fish fed prebiotic diet and intestines exposed to saline (group 5): The intestine exposed to saline of fish fed the prebiotic diet for 15 weeks showed higher diversity compared to the other groups exposed to saline (groups 1 and 3). Of the 47 strains isolated from the PI, 14 were identified as *C. divergens*, 1 as *Pseudomonas antarctica*, 1 as *Pseudomonas korensis*, 4 as *Enterobacter hormaechei* and 1 as

uncultured bacterial clone CK20. The remaining isolated strains were identified as members of the *Carnobacterium* and *Acinetobacter* genera. The dominant bacteria in the PI of this group belonged to carnobacteria (81%) and 30% of total isolates were identified as *C. divergens*.

The bacterial composition of the isolates from the DI of fish fed the prebiotic diet for 15 weeks were relatively low in diversity. Of the total number of strains isolated (44) from the DI exposed to saline 34 were identified as *Carnobacterium*, 8 strains showed high similarity (%) to *Pantoea* spp. and 2 strains belonged to the genus *Enterobacter*.

Microbiota of fish fed prebiotic diet and intestines exposed to *C. divergens* (group 6): In group 6, fish fed the prebiotic diets for 15 weeks and exposed to *C. divergens*, the isolated strains in the PI were dominated by *C. divergens* and *C. divergens* - like strains. Of the 22 carnobacteria isolated, 5 were identified as *C. divergens* by 16S rRNA sequencing while 17 isolates were identified as *C. divergens* - like. Three other isolates were identified as members of the genera *Pseudomonas* (2 strains) and *Psychrobacter* (1 strain). Of the 17 strains isolated and identified from the DI of group 6, *C. divergens* and *C. divergens* - like strains dominated with only one isolate, which showed high similarity (99 %) to *Acinetobacter* spp., not belonging to this species.

Microscopical analyses

Light microscopy (LM): All LM micrographs, both from PI and DI of the prebiotic groups (5 and 6) showed no morphological differences compared to the control feeding regime (groups 1 - 4). All intestinal sections examined appeared normal and healthy; no signs of detached enterocytes, necrotic enterocytes, widened lamina propria or necrosis were observed and the number of goblet cells were similar in both treatments (examples are displayed in Figure 1).

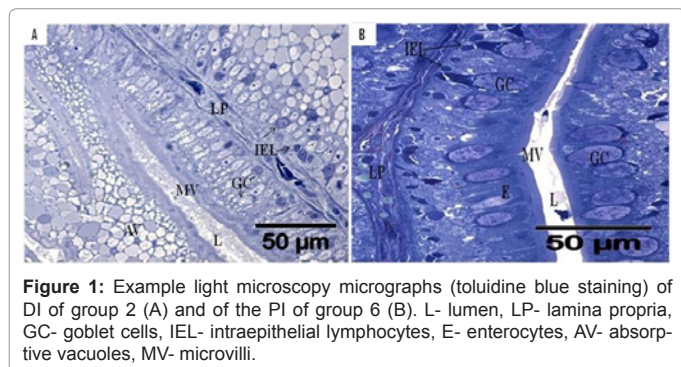


Figure 1: Example light microscopy micrographs (toluidine blue staining) of DI of group 2 (A) and of the PI of group 6 (B). L- lumen, LP- lamina propria, GC- goblet cells, IEL- intraepithelial lymphocytes, E- enterocytes, AV- absorptive vacuoles, MV- microvilli.

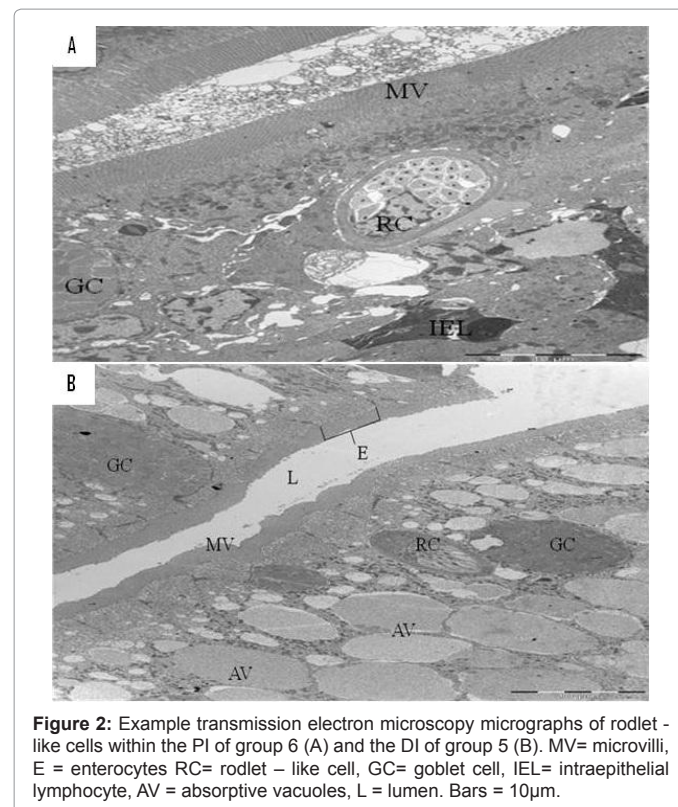


Figure 2: Example transmission electron microscopy micrographs of rodlet - like cells within the PI of group 6 (A) and the DI of group 5 (B). MV= microvilli, E = enterocytes RC= rodlet - like cell, GC= goblet cell, IEL= intraepithelial lymphocyte, AV = absorptive vacuoles, L = lumen. Bars = 10µm.

| Isolate code | Source group | Intestinal region | Closest known species | Strain | Accession no. | Identity (%) | Antagonism | |
|--------------|--------------|-------------------|---------------------------|------------|---------------|--------------|-------------------|-----------------------|
| | | | | | | | <i>Y. ruckeri</i> | <i>A. salmonicida</i> |
| 33 | Group 2 | Proximal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 98 | + | - |
| 40 | Group 2 | Distal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 98 | + | - |
| 75 | Group 3 | Proximal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 99 | + | - |
| 84 | Group 3 | Distal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 100 | + | - |
| 14 | Group 5 | Proximal | <i>Carnobacterium</i> sp. | H126a | EF204312.1 | 86 | + | - |
| 57 | Group 5 | Proximal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 99 | + | + |
| 17 | Group 5 | Distal | <i>Carnobacterium</i> sp. | H126a | EF204312.1 | 99 | + | - |
| 154 | Group 5 | Distal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 92 | + | - |
| 173 | Group 4 | Proximal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 99 | + | - |
| 127 | Group 4 | Distal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 99 | + | - |
| 99 | Group 6 | Proximal | <i>C. divergens</i> | LHICA_53_4 | FJ656716.1 | 99 | + | - |
| - | - | - | <i>C. inhibens</i> * | CCUG 31728 | - | - | + | - |

*- originally isolated from the digestive tract of Atlantic salmon (*Salmo salar*) [20]

Table 4: Identification of LAB strains and pathogen antagonistic activity of extracellular products used in the *in vitro* pathogen assays.

Transmission electron microscopy (TEM): Similar to the observations using LM, TEM revealed no differences between treatments or exposure groups; all micrographs revealed healthy epithelial brushborder, no deterioration of tight junctions was observed and microvilli appeared uniform. The presences of rodlet - like cells (as shown in Figure 2) were present in the PI and DI of all groups. The numbers of rodlet cells present in the PI displayed great differences between individual fish but were always observed in the upper half of the epithelium, above the underlying intraepithelial lymphocytes.

***In vitro* growth inhibition of two fish pathogens by extracellular extracts of LAB isolated from *ex vivo* studies**

Identification by partial sequences of the 16S rRNA genes of the eleven LAB strains isolated from the *ex vivo* experiments and subsequently used in the *in vitro* pathogen antagonism assays are displayed in Table 4. The results show that growth inhibition of *Y. ruckeri* was obtained from extracellular extracts from all strains of carnobacteria isolated from the *ex vivo* experiment. However, *in vitro* growth inhibition of *A. salmonicida* ssp. *salmonicida* was only obtained from the extracellular extract of *C. divergens* isolate 57. The extracellular products from the positive control, *C. inhibens* CCUG 31728, did not inhibit the growth of *A. salmonicida* ssp. *salmonicida*.

Discussion

The *ex vivo* intestinal sack method has been used in several studies to evaluate possible histological changes in the fish intestine after exposure to high levels of LAB [9,11,13-15]. The result of LAB exposure to the intestine is of high importance as translocation and cell damage have been proposed as important criteria when evaluating the use of probiotics in endothermic animals as well as in fish [16,17]. Recently, the effect of *ex vivo* LAB exposure on the gut microbiota in fish was documented [13-15], but to the authors' knowledge the effect of prebiotic supplementation and *ex vivo* LAB exposure of the fish intestine has not been investigated.

The culturable bacterial levels recovered on TSAgs plates from groups exposed to saline were relatively low, ranging from log 1.72 to 2.34 CFU g⁻¹. These values are low compared to autochthonous levels previously reported in Atlantic salmon [25] and rainbow trout *Oncorhynchus mykiss* [26,27]. This is likely due to the thorough rinsing process, 3 times prior and 3 times post probiotic/saline exposure. Culturable adherent bacteria in the PI observed in the prebiotic group

exposed to saline (group 5) was log 2.34 CFU g⁻¹. The fact that the value from this group is higher than in control groups exposed to saline water (log 1.72 and 2.08 CFU g⁻¹), might be due to a feeding effect of the prebiotic diet. However, this hypothesis merits further investigations.

Investigations of the qualitative and quantitative bacterial composition of the intestinal microbiota were based on the study of 168 pure cultured bacterial isolates. These isolates were biochemically tested in order to obtain a general classification. From this classification 111 isolates were selected by the lottery method and identified by 16S rRNA gene sequencing analysis.

The bacterial levels observed in groups exposed to saline varied between segments and feeding regime. By comparing different feeding groups it is clear that the indigenous microbiota of the PI were affected by the diet, while the effect of prebiotic feeding on the microbiota was less clear in the DI. In the intestine exposed to *C. divergens* the average number of bacteria were higher in the PI when the fish were fed the prebiotic diet compared to fish fed the control diet. Whether these findings can be related to a higher *C. divergens* colonization success in prebiotic fed fish merits further investigations.

The bacterial composition from the PI in control groups, i.e. fish fed control diet, and thereafter exposed to saline were dominated by members of a few genera (*Psychrobacter*, *Carnobacterium* and *Pseudomonas*). *Psychrobacter*, *Carnobacterium* and *Pseudomonas* spp. are commonly reported in the GI tract of fish, and these bacteria have previously been isolated and identified from the GI tract of salmonids [3,25-30]. *Carnobacterium* spp. have often been reported to be components of the gut microbiota of salmonids; indeed, *C. maltaromaticum*, *C. mobile*, *C. divergens* and *Carnobacterium* spp. have been identified from Atlantic salmon [29-33]. The consistency of isolation of these species indicates that these might be common core components of the GI microbiome of Atlantic salmon and are likely to be of importance to the host.

The bacterial composition isolated from the DI in the first control group exposed to saline (group 1) were dominated by *Pseudoalteromonas* spp. and *Psychrobacter* spp., while in the other control group (group 3) *Acinetobacter* spp. and *C. divergens* were the dominant bacteria isolated. All of these listed bacteria have previously been isolated from the intestine of Atlantic salmon [29,30,34]. The bacterial composition of the PI observed in the prebiotic fed fish exposed to saline (group 5) showed greater bacterial diversity than that observed from control fed fish exposed to saline (groups 1 and 3). The majority of the bacteria

from the PI of group 5 were *C. divergens* and *Carnobacterium* spp., which together accounted for 81% of all the isolated bacteria. The two other bacterial species that were isolated from this group were *Pantoea* spp. and an unidentified member of the class Gammaproteobacteria. The abundance of culturable adherent *Carnobacterium* spp. (77% of isolates identified) was higher in the DI of the prebiotic fed fish (group 5), compared to control groups (group 1 = 0% and group 3 = 36%). These results suggest that the prebiotic supplementation elevates autochthonous *Carnobacterium* spp. levels, particularly in the DI. To the authors' knowledge there is very little information regarding the effect of prebiotics on carnobacteria within the GI tract of fish. However, some studies suggest that the carnobacteria populations within the GI tract of salmonids are effected by various dietary factors such as krill meal [32] and oxytetracycline [29] in Atlantic salmon and dietary carbohydrates in Arctic charr (*Salvelinus alpinus* L.) [19,35]. However, it was observed that the presence of dietary inulin (a prebiotic-type carbohydrate) tended to lower culturable autochthonous carnobacteria levels (by ca. 90%) in the hindgut of Arctic charr and also elevated the proportion of *C. maltaromaticum* at the expense of *C. divergens* [19]. These findings suggest that different prebiotics may influence different carnobacteria strains in different fish species.

In all groups exposed to *C. divergens* in the *ex vivo* studies the same *C. divergens* strain was identified to dominate both the PI and DI after exposure. *C. divergens* levels were in the range of 10^4 - 10^6 CFU g^{-1} intestine which indicates that the bacteria are able to populate and potentially colonize the intestinal mucus and out-compete other adherent bacteria after only one hour of exposure. These results are in accordance with corresponding studies in that lactic acid bacteria are able to colonize the intestine of Atlantic salmon after one hour exposure [11].

Despite the plethora of information available on the prebiotic efficacy of elevating probiotic colonization (i.e. synbiotics) in various terrestrial species, little information is available in fish [13]. Further studies should focus on this topic as the present study demonstrated that the presence of the dietary prebiotic, prebiosal®, elevated the proportion of carnobacteria from 71% to 81% in the DI (as well as elevating total bacterial levels, effectively quadrupling the number of carnobacteria) and from 33% to 77% in the PI (although the total bacterial population was lower).

The histological effect of exposing the GI tract of Atlantic salmon to high levels of the *C. divergens* was investigated by light and electron microscopy. Furthermore, the intestinal effects of feeding a prebiotic diet to Atlantic salmon were evaluated.

Results from LM-investigations in the present study showed no apparent histopathological changes of the epithelium in the PI or DI, after exposure of *C. divergens*. In particular the micrographs demonstrated that enterocytes showed no signs of junctional rupture from the basement membrane which is in contrast to observations of the PI of Atlantic salmon after exposure to *Vibrio anguillarum* and *A. salmonicida* [9].

TEM observations confirmed the findings observed in LM regarding a lack of histological changes. TEM revealed no observable differences between the groups in respect to the presence of cell debris in the lumen, the amount of mucus, the number bacteria – like particles in the lumen and between the microvilli, disorganized microvilli and disintegrated

tight junctions. The enterocytes within all groups displayed normal cell contacts with unaffected tight junctions and *zona adherens*. The fact that *C. divergens* did not inflict damage to the intercellular junction is of *vital property* since the loosening of these junctions contributes to a paracellular port of entry for potential pathogens [17].

Rodlet cells were present in large numbers in the PI of all groups, while in the DI the number observed were lower. Since groups exposed to *C. divergens* did not display any clear differences in number of rodlet cells compared to groups exposed to saline, their presence in those groups may therefore not be related to an immunological function towards the exposed bacteria. On the other hand, the role of rodlet cells as immune cells and their large number in the PI compared to the DI may be a defense function towards potential invading bacteria of the PI. Since the PI has been confirmed as being an infection route for pathogenic bacteria by several studies [8,9,36], the role of rodlet cells as immune cells in the PI is possible and warrants further investigation.

The antimicrobial effects of LAB have long been utilized in food preservation by fermentation and several comprehensive reviews have been published on the ability of LAB to produce proteinaceous antimicrobial substances [2,3,7,37]. In fish studies, the antagonistic effect of LAB has been carried out on Gram-negative fish pathogens such as *V. anguillarum* and *A. salmonicida* [3,21,38-40]. In the present study strong growth inhibition of *Y. ruckeri* was recorded from extracellular extracts from late exponential growth phase from all of the eleven *Carnobacterium* strains isolated from the *ex vivo* experiments. However, the ability of the isolated strains to inhibit growth of *A. salmonicida* ssp. *salmonicida* was only observed from one strain isolated from the PI. The fact that only one (isolate 57) of the 11 strains displayed inhibitory effects towards *A. salmonicida* ssp. *salmonicida* is in accordance with the results of Ringø [22] who observed a lack of antagonism when challenging *A. salmonicida* ssp. *salmonicida* to extracellular extracts from *C. divergens* strain Lab01. These results indicate that the production of extracellular products only, might not be sufficient for strains of *C. divergens* in late exponential growth phase, to inhibit growth of *A. salmonicida* ssp. *salmonicida*.

The positive control bacteria, *C. inhibens* which Jöborn et al. [41] reported to display antagonistic effect against *A. salmonicida*, showed no sign of antagonism in the present study. This observation therefore indicates that antagonistic activity of *C. inhibens* is only effective when cells are actively incubated together or that antagonistic extracellular products are only produced by *C. inhibens* in the presence of *A. salmonicida*.

The ability of *C. divergens* as useful probiotics with effects against *Y. ruckeri* and *A. salmonicida* have previously been reported *in vivo* and *in vitro* [42]. Kim and Austin [42] observed that dietary provision of *C. divergens* strain B33, isolated from the intestine of healthy rainbow trout, increased survival of rainbow trout against *A. salmonicida* and *Y. ruckeri* challenge by 60% compared to the control group. Even though strains of *C. divergens* show antagonistic effects against pathogens, the precise mechanism of action of antimicrobial compounds isolated from fish remains unclear, but suggestions about their ability of penetrating cell walls by forming pores and channels, thus rendering it more fragile and incapable of carrying out normal metabolism has been proposed [43,44].

In order to confirm the *in vitro* probiotic effect of *C. divergens* against *Y. ruckeri* in Atlantic salmon, further investigations should therefore include *in vivo* challenges studies. By further applying electron

microscopy, the physical interference mechanisms between *C. divergens* and *Y. ruckeri* in the GI tract might be observed.

Acknowledgements

The authors thank the technical staff at EWOS Innovation AS for feed manufacture, analysis and running the feeding trial and thank Dr. Sigmund Sperstad and Dr. Chun Li, Norwegian College of Fishery Science, University of Tromsø for their inestimable help during 16S rRNA gene sequencing and *in vitro* growth inhibition. We also thank Randi Olsen and Helga Marie By at the EM department at University of Tromsø, and Anne Nyhaug at Molecular Imaging Centre, Institute for Biomedicine, University of Bergen for their inestimable help during light and electron microscopy analysis. This study was partially supported by grants from the Norwegian MABIT-program (project number AF0038).

References

- Birkbeck TH, Ringø E (2005) Pathogenesis and the gastrointestinal tract of growing fish. In: Microbial Ecology in Growing Animals. Holzapfel, W. and Naughton, P. (Eds.). pp. 208-234. Elsevier, Edinburgh, UK.
- Gatesoupe FJ (1999) The use of probiotics in aquaculture. Aquaculture 180: 147-165.
- Ringø E, Schillinger U, Holzapfel W (2005) Antibacterial abilities of lactic acid bacteria isolated from aquatic animals and the use of lactic acid bacteria in aquaculture. In: Holzapfel, W., Naughton, P. (Eds.), Microbial Ecology in Growing Animals, Elsevier, Edinburgh, UK, pp. 418-453.
- Austin B (2006) The bacterial Microflora of fish, revised. The Scientific World Journal 6: 931-945.
- Ringø E, Sperstad S, Myklebust R, Refstie S, Krogdahl Å (2006) Characterisation of the microbiota associated with the intestine of Atlantic cod (*Gadus morhua* L.). The effect of fish meal, soybean meal and a bioprocessed soybean meal. Aquaculture 261: 829-841.
- Caipang CMA, Brinchmann MF, Kiron V (2010) Antagonistic activity of bacterial isolates from intestinal microbiota of Atlantic cod, *Gadus morhua*, and an investigation of their immunomodulatory capabilities. Aquac Res 41: 249-256.
- Merrifield DL, Dimitroglou A, Foey A, Davies S J, Baker RTM, et al. (2010) The current status and future focus of probiotic and prebiotic applications for salmonids. Aquaculture 302: 1-18.
- Ringø E, Jutfelt F, Kanopathippillai P, Bakken Y, Sundell K, et al. (2004) Damaging effect of the fish pathogen *Aeromonas salmonicida* ssp. *salmonicida* on intestinal enterocytes of Atlantic salmon (*Salmo salar* L.). Cell Tissue Res 318: 305-311.
- Ringø E, Salinas I, Olsen RE, Nyhaug A, Myklebust R, et al. (2007) Histological changes in Atlantic salmon (*Salmo salar* L.) intestine following *in vitro* exposure to pathogenic and probiotic bacterial strains. Cell Tis Res 328: 109-116.
- Jutfelt F, Sundh H, Glette J, Mellander L, Björnsson BT, et al. (2008) The involvement of *Aeromonas salmonicida* virulence factors in bacterial translocation across the rainbow trout, *Oncorhynchus mykiss* (Walbaum), intestine. J Fish Dis 31: 141-151.
- Salinas I, Myklebust R, Esteban MA, Olsen RE, Meseguer J, et al. (2008) *in vitro* studies of *Lactobacillus delbrueckii* subsp. *lactis* in Atlantic salmon (*Salmo salar* L.) foregut: tissue responses and evidence of protection against *Aeromonas salmonicida* subsp. *salmonicida* epithelial damage. Vet Microbiol 128: 167-177.
- Khemiss F, Ahmadi S, Massoudi R, Ghoul-Mazgar S, Safta S, et al. (2009) Effect of *in vitro* exposure to *Vibrio vulnificus* on hydroelectrolytic transport and structural changes of sea bream (*Sparus aurata* L.) intestine. Fish Physiol Biochem 35: 541-549.
- Ringø E, Løvmo L, Kristiansen M, Salinas I, Myklebust R, et al. (2010) Lactic acid bacteria vs. pathogens in the gastrointestinal tract of fish: A review. Aquac Res 41: 451-467.
- Løvmo Martinsen L, Salma W, Myklebust R, Mayhew TM, Ringø E (2011) *Carnobacterium maltaromaticum* vs. *Vibrio (Listonella) anguillarum* in the midgut of Atlantic cod (*Gadus morhua* L.). An *ex vivo* study. Aquac Res early view doi:10.1111/j.1365-2109.2011.02784.x.
- Salma W, Zhou Z, Wang W, Askarian F, Kousha A, et al. (2011) Histological and bacteriological changes in intestine of beluga (*Huso huso*) following *ex vivo* exposure to bacterial strains. Aquaculture 314: 24-33.
- Salminen S, von Wright A, Morelli L, Marteau P, Brassart D, et al. (1998) Demonstration of safety of probiotics – a review. Int J Food Microbiol 44: 93-106.
- Ringø E, Myklebust R, Mayhew TM, Olsen RE (2007) Bacterial translocation and pathogenesis in the digestive tract of larvae and fry. Aquaculture 268: 251-264.
- Strøm E (1988) Melkesyrebakterier i fisketarm. Isolasjon, karakterisering og egenskaper. MSci. thesis. The Norwegian College of Fishery Science, University of Tromsø. 88 pp. In Norwegian.
- Ringø E, Sperstad S, Myklebust R, Mayhew TM, Olsen RE (2006) The effect of dietary inulin on aerobic bacteria associated with hindgut of Arctic charr (*Salvelinus alpinus* L.). Aquac Res 37: 891-897.
- Jöborn A, Olsson C, Westerdahl A, Conway P, Kjelleberg S (1997) Colonization in the fish intestinal tract and production of inhibitory substances in intestinal mucus and faecal extracts by *Carnobacterium* sp. strain K1. J Fish Dis 20: 383-392.
- Olsson C (1995) Bacteria with inhibitory activity and *Vibrio anguillarum* in the fish intestinal tract. Department of General and Marine Microbiology, Gothenburg University, Sweden. ISBN 91-628-1850-3.
- Ringø E (2008) The ability of carnobacteria isolated from fish intestine to inhibit growth of fish pathogenic bacteria: a screening study. Aquac Res 39: 171-180.
- McDowell EM, Trump BF (1976) Histologic fixatives suitable for diagnostic light and electron-microscopy. Arch Pathol Lab Med 100: 405-414.
- Ringø E, Lødemel JB, Myklebust R, Kaino T, Mayhew TM, et al. (2001) Epithelium-associated bacteria in the gastrointestinal tract of Arctic charr (*Salvelinus alpinus* L.). An electron microscopical study. J Appl Microbiol 90: 294-300.
- Hovda MB, Lunestad BT, Fontanillas R, Rosnes JT (2007) Molecular characterisation of the intestinal microbiota of farmed Atlantic salmon (*Salmo salar* L.). Aquaculture 272: 581-588.
- Merrifield DL, Burnard D, Bradley G, Davies SJ, Baker RTM (2009) Microbial community diversity associated with the intestinal mucosa of farmed rainbow trout (*Oncorhynchus mykiss* Walbaum). Aquac Res 40: 1064-1072.
- Merrifield DL, Bradley, G Baker RTM, Dimitroglou A, Davies SJ (2009) Soybean meal alters autochthonous microbial populations, microvilli morphology and compromises intestinal enterocyte integrity of rainbow trout (*Oncorhynchus mykiss* Walbaum). J Fish Dis 32: 755-766.
- Spanggaard B, Huber I, Nielsen J, Nielsen T, Appel KF, et al. (2000) The microflora of rainbow trout intestine: a comparison of traditional and molecular identification. Aquaculture 182: 1-15.
- Bakke-McKellep AM, Penn MH, Salas PM, Refstie S, Sperstad S, et al. (2007) Effects of dietary soyabean meal, inulin and oxytetracycline on intestinal microbiota and epithelial cell stress, apoptosis and proliferation in the teleost Atlantic salmon (*Salmo salar* L.). Br J Nutr 97: 699-713.
- Ringø E, Sperstad S, Kraugerud OF, Krogdahl Å (2008) Use of 16S rRNA gene sequencing analysis to characterize culturable intestinal bacteria in Atlantic salmon (*Salmo salar* L.) fed diets with cellulose or non-starch polysaccharides from soy. Aquac Res 39: 1087-1100.
- Ringø E, Bendiksen HR, Wesmajervi MS, Olsen RE, Jansen PA, et al. (2000) Lactic acid bacteria associated with the digestive tract of Atlantic salmon (*Salmo salar* L.). J Appl Microbiol 89: 317-322.
- Ringø E, Sperstad S, Myklebust R, Mayhew TM, Mjelde A, et al. (2006) The effect of dietary krill supplementation on epithelium-associated bacteria in the hindgut of Atlantic salmon (*Salmo salar* L.). A microbial and electron microscopical study. Aquac Res 37: 1644-1653.
- Skrodenyte-Arbaciauskienė V, Sruoga A, Butkauskas D, Skrupskelis K (2008) Phylogenetic analysis of intestinal bacteria of freshwater salmon *Salmo salar* and sea trout *Salmo trutta trutta* and diet. Fish Sci 74: 1307-1314.
- Ringø E, Wesmajervi MS, Bendiksen HR, Berg A, Olsen RE, et al. (2001) Identification and characterization of Carnobacteria isolated from fish intestine. Syst Appl Microbiol 24: 183-191.
- Ringø E, Olsen RE (1999) The effect of diet on aerobic bacterial flora associated with intestine of Arctic charr (*Salvelinus alpinus* L.). J Appl Microbiol 86: 22-28.
- Lødemel JB, Mayhew TM, Myklebust R, Olsen RE, Espelid S, et al. (2001) Effects of three dietary oils on disease susceptibility in Arctic charr (*Salvelinus alpinus* L.) during cohabitant challenge with *Aeromonas salmonicida* ssp. *salmonicida*. Aquac Res 32: 935-945.

37. Ringø E, Gatesoupe FJ (1998) Lactic acid bacteria in fish: a review. *Aquaculture* 160: 177-203.
38. Balcázar JL, Vendrell D, de Blas I, Ruiz-Zarzuola I, Gironés O, et al. (2007) *In vitro* competitive adhesion and production of antagonistic compounds by lactic acid bacteria against fish pathogens. *Vet Microbiol* 122: 373-380.
39. Balcázar JL, Vendrell D, de Blas I, Ruiz-Zarzuola I, Múzquiz JL, et al. (2008) Characterization of probiotic properties of lactic acid bacteria isolated from intestinal microbiota of fish. *Aquaculture* 278: 188-191.
40. Askarian F, Zhou Z, Olsen RE, Sperstad S, Ringø E (2011) Culturable autochthonous gut bacteria in Atlantic salmon (*Salmo salar* L.) fed diets with or without chitin. Characterization by 16S rRNA gene sequencing, ability to produce enzymes and *in vitro* growth inhibition of four fish pathogens. *Aquaculture* early view doi: 10.1016/j.aquaculture.2011.10.016.
41. Jöborn A, Dorsch M, Olsson JC, Westerdahl A, Kjelleberg S (1999) *Carnobacterium inhibens* sp nov., isolated from the intestine of Atlantic salmon (*Salmo salar*). *Int J of Syst Bacteriol* 49: 1891-1898.
42. Kim DH, Austin B (2006) Innate immune responses in rainbow trout (*Oncorhynchus mykiss*, Walbaum) induced by probiotics. *Fish Shellfish Immunol* 21: 513-524.
43. Steiner H, Andreu D, Merrifield RB (1988) Binding and action of cecropin and cecropin analogues: antibacterial peptides from insects. *Biochim Biophys Acta* 939: 260-266.
44. Park CB, Kim HS, Kim SC (1998) Mechanism of Action of the Antimicrobial Peptide Buforin II: Buforin II Kills Microorganisms by Penetrating the Cell Membrane and Inhibiting Cellular Functions. *Biochim Biophys Res Comm* 244: 253-257.

This article was originally published in a special issue, **Probiotic & Prebiotic Applications in Aquaculture** handled by Editor(s). Dr. Daniel L. Merrifield, University of Plymouth, UK; Prof. Zhigang Zhou, Chinese Academy of Agricultural Sciences, China