FACULTY OF HEALTH SCIENCES
DEPARTMENT OF COMMUNITY MEDICINE

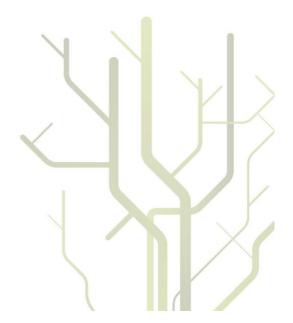
Staphylococcus aureus nasal carriage – Interplay between host, microbe and the environment.

-The Tromsø Staph and Skin Study



A dissertation for the degree of Philosophiae Doctor

Tromsø 2013



Staphylococcus aureus nasal carriage – Interplay between host, microbe and the environment.

Results from the Tromsø Staph and Skin Study

By

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PREFACE: FROM MICROBIOLOGY AND INFECTION CONTROL TO EPIDEMIOLOGICAL RESEARCH

During my more than 20 years as an MD in clinical medicine, infectious diseases, microbiology and infection control, I have met numerous patients with nosocomial- and community acquired infections with *Staphylococcus aureus*. For some patients, these infections caused severe diseases like deep surgical site infections, catheter related infections and septicemias. For others, however, colonization with methicillin-resistant *S. aureus* (MRSA) caused serious trouble for those in need of treatment from medical health services or for those working as a healthcare professional. This fact developed my interest for host susceptibility factors for *S. aureus* carriage and my main supervisor Anne-Sofie Furberg introduced me to this field.

Most of the patients with *S. aureus* infections or MRSA colonization were living in the community of Tromsø, and the Tromsø Study had for years explored the health of its inhabitants. The Tromsø 6 survey was in the planning phase when I prepared my PhD project. It is a short distance from the University Hospital in Tromsø to the epidemiological setting at the Department of Community Medicine where I met Anne-Sofie Furberg. She had a background both in microbiology, epidemiology and cancer research. During these last three years, I have had the privilege to more carefully investigate the interesting relationships between different host susceptibility factors like lifestyle, and metabolic and hormonal profile, and *S. aureus* nasal carriage.

ACKNOWLEDGEMENTS

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I am very grateful that I was given the opportunity to join the Tromsø Staph and Skin Study group and to perform the studies described in the thesis.

First and foremost, I would like to thank my main supervisor and mentor, Anne-Sofie Furberg. She has been an excellent teacher, always attentive and interested in my work, and always responding friendly and constructively to my sometimes less coherent thoughts and ideas. I appreciate her patience and for sharing her significant knowledge concerning aspects of microbiology and in particular *S. aureus* carriage, epidemiology and statistical methods. Her efficiency and sense of structure is impressing and inspiring, and has been essential along the way.

I will thank Gunnar Skov Simonsen, director at the Department of Microbiology and Infection Control, and one of my co-supervisors, for giving me the opportunity, for letting me in to this fascinating world of host-microbe interactions for *S. aureus* carriage, and for his significant knowledge concerning microbiology and *S. aureus* in particular.

Furthermore, I want to thank Inger Thune, my second co-supervisor, for her enthusiasm and critical guidance, and for her sharing of her significant experience and knowledge and for heaps of encouragements along the way.

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I want to thank Maria Sangvik, my fellow PhD candidate, for her contribution in *spa* typing the *S. aureus* isolates and for her discussions regarding bacterial factors facilitating *S. aureus* colonization. Tom Wilsgaard is thanked for both brief and more extensive discussions regarding statistical problems. I want to thank my other co-authors for good collaboration. My colleagues and staff at the Department of Microbiology and Infection Control are thanked for taking on a larger share of work during my period of absence.

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ENGLISH SUMMARY

Staphylococcus aureus (S. aureus) can act both as a human commensal that persistently colonizes 20–30% of the adult population, and as an invasive pathogen. S. aureus nasal carriage often precedes infection. Emergence and spread of antimicrobial resistance combined with increasing numbers of immune-compromised patients make infections increasingly difficult to treat. Thus, new insight into the predisposing factors of S. aureus nasal carriage may give novel clues to host-microbe-environmental interactions of importance for the carrier state, and thus, contributing substantially in reducing the burden of S. aureus disease.

In this thesis, we investigated whether host factors (gender, serum vitamin D levels, body mass index [BMI], and waist circumference [WC]), environmental factors (smoking, work in healthcare services, and residing with children) and microbe (*spa* types) were associated with *S. aureus* nasal carriage among women and men aged 30–87 years who participated in the Tromsø Staph and Skin Study– part of the sixth Tromsø survey (Tromsø 6) carried out from October 2007 to December 2008.

S. aureus nasal carriage was more common in men than in women (34.1% and 21.3%, respectively) and more common among non-smokers than among smokers. There was an inverse dose-response association between serum 25(OH)D concentration and the odds of *S. aureus* nasal colonization and carriage in non-smoking men.

We observed that young and premenopausal women with higher BMI and WC had increased odds of *S. aureus* nasal colonization independent of pre-diabetes/diabetes, and use of hormonal contraceptives. There was no association among older women and men while the association with higher WC was observed among young men.

Work in healthcare services was associated with increased odds of *S. aureus* nasal carriage among women. Odds were even higher among women residing with children. Among men, work in healthcare services and residing with children were associated with increased odds of common *spa* types. Our study suggests that a synergism between environmental risk factors (work and household) is of importance for the overall *S. aureus* carrier state in HCWs.

In summary, our cross-sectional study supports the view that there is a complex interplay between host-, microbial-, and environmental factors during colonization and carriage. Prospective studies are needed to determine causal relationships and targets for prevention.

SAMMENDRAG

Staphylococcus aureus er en av de viktigste årsakene til alvorlige infeksjoner hos mennesker. Bakterien kan kolonisere oss uten å skape sykdom, men den kan også invadere ulike typer vev og blodbanen og gi alvorlig infeksjon. S. aureus trives best i nesen, og oftest er det vår egen nesestamme som er årsak til infeksjonen. Effektiv behandling av S. aureus infeksjon er en klinisk utfordring pga globalt økende antibiotikaresistens og flere immunsupprimerte pasienter med mer kompliserte behandlingsforløp. Økt kunnskap om faktorer som fremmer kolonisering med bakterien, kan gi kunnskap om nye metoder for å forebygge infeksjon med S. aureus.

Ca 20–30% av den voksne normalbefolkning er bærere av *S. aureus* og årsakene til at noen er bærere mens andre ikke er det, er i stor grad ukjent. Både vert- og miljøfaktorer samt forhold ved mikroben synes å spille en rolle.

I denne avhandlingen har vi testet om ulike faktorer hos vert (kjønn, vitamin D-nivå i serum, kroppsmasseindeks og livvidde), miljø (røyking, arbeid i helsevesenet, bo med barn) og mikrobe (*spa* type) har betydning for bærerskap av *S. aureus* hos kvinner og menn i alderen 30–87 år som deltok i den befolkningsbaserte undersøkelsen–Tromsø 6 i 2007–2008.

Resultatene viser at menn er hyppigere bærere av *S. aureus* i nesen enn kvinner (34.1% versus 21.3%). Røykere har lavere prevalens av *S. aureus* bærerskap enn ikkerøykere. Høyere serum vitamin D var forbundet med lavere risiko for bærerskap hos ikkerøykende menn; en halvert risiko ble observert hos de med høyest serum vitamin D \geq 75 nmol/l versus de med lavest nivå <50 nmol/l.

Hos unge og premenopausale kvinner var høyere kroppsmasseindeks og livvidde forbundet med økt risiko for *S. aureus* nesebærerskap uavhengig av diabetes og bruk av hormonelle prevensjonsmidler. Sammenhengen med høy livvidde ble også funnet hos unge menn, mens det ikke ble funnet tilsvarende sammenhenger hos eldre kvinner og menn.

Kvinnelige helsearbeidere og især de som bodde sammen med barn, hadde økt risiko for nesebærerskap av *S. aureus*. Funnene var ikke signifikante hos mannlige helsearbeidere. *spa* type t012 og t015 var assosiert med jobb som helsearbeider. Resultatene tyder på at nesebæreskap av *S. aureus* bestemmes av både vert, miljø og mikrobielle faktorer. Prospektive studier er nødvendige for å avklare årsakssammenhenger og mål for forebygging.

LIST OF PAPERS

This thesis is based on the following three papers, which are referred to in the text by their Roman numerals.

Paper I

Olsen K, Falch BM, Danielsen K, Johannessen M, Sollid JUE, Thune I, Grimnes G, Jorde R, Simonsen GS, Furberg A-S.

Staphylococcus aureus nasal carriage is associated with serum 25-hydroxyvitamin D levels, gender and smoking status. The Tromsø Staph and Skin Study. European Journal of Clinical Microbiology & Infectious Diseases. 2012;31(4):465-473.

Paper II

Olsen K, Danielsen K, Wilsgaard T, Sangvik M, Sollid JUE, Thune I, Eggen AE, Simonsen GS, Furberg A-S.

Obesity and *Staphylococcus aureus* nasal colonization among women and men in a general population. Accepted in *PLoS ONE*, April 2013.

Paper III

Olsen K, Sangvik M, Simonsen GS, Sollid JUE, Sundsfjord A, Thune I, Furberg A-S.

Prevalence and population structure of *Staphylococcus aureus* nasal carriage in healthcare workers in a general population. The Tromsø Staph and Skin Study. *Epidemiology and Infection*. 2013;141(1):143-52.

1. INTRODUCTION

Staphylococcus aureus (S. aureus) can act both as a human commensal, that persistently colonizes 20–30% of the adult human population (S. aureus carriers), and as an invasive pathogen [1]. S. aureus is the major cause of skin and soft tissue infections, and the bacterium can invade any tissue in the body, causing other serious life-threatening diseases such as osteomyelitis, endocarditis, and pneumonia. S. aureus is a major cause of bloodstream infection and was the 2nd most common pathogen isolated in blood cultures in Norway in 2011 [2]. S. aureus nasal carriage often precedes infection. The bacterium colonizes the skin and mucosa of humans and several animal species [3, 4]. Although multiple body sites can be colonized in human beings, the anterior nares of the nose are the main body sites. Emergence and spread of antimicrobial resistance combined with increasing numbers of immunocompromised patients make infections increasingly difficult to treat [5]. Thus, new insight into the patophysiology as well as predisposing factors of S. aureus nasal carriage may give novel clues to host-microbe-environmental interactions of importance for the carrier state and provide new potential targets for prevention of infection.

1.1 Clinical significance

S. aureus is one of the most widespread human pathogens with the potential to cause serious and fatal diseases. The organism is well armed with potent virulence factors, survival fitness, and antimicrobial resistance determinants [6]. The spectrum of infections encompasses skin and soft tissue infections (SSTIs), muscle and visceral abscesses, septic arthritis, osteomyelitis, endocarditis, pneumonia, brain abscesses, meningitis and bacteremia, as well as toxinoses with toxic shock syndrome, scalded skin syndrome, and food poisoning [6]. Globally, S. aureus is the cause of a large proportion of bloodstream infections (22%), and skin and soft tissue infections (39%) [7]. In Norway, S. aureus is the second most common bacterial species in blood cultures, accounting for 14.2% of the isolates when skin contaminants are excluded [2].

The annual incidence of *S. aureus* bacteremia (SAB) varies between 19.7 and 50 per 100,000 populations in different countries such as Canada and the Scandinavian countries with the lowest incidence and USA with the highest incidence. These large geographical discrepancies may reflect differences in the prevalence of *methicillin-resistant S. aureus*

(MRSA), healthcare systems, infection control practices and the completeness of surveillance data [8]. There is substantial variation in the mortality rates (range 10–30%) [8] most likely attributable to differences in patient groups and complications due to bacteremiae, prevalence of MRSA and the mortality measurements used. Remarkably, in the Western part of the world, the 30-day all-cause mortality of SAB exceeds that of AIDS, tuberculosis and viral hepatitis, and is almost equal to that of breast and prostate cancers [8].

The Centers for Disease Control and Prevention (CDC) definition divides S. aureus infections into nosocomial (onset of infection >48 h after hospital admission), communityonset healthcare-associated (HA) [onset of infection in the community or <48 h after hospital admission and the presence of at least one of the following risk factors: a history of hospitalization, surgery, dialysis, or residence in a long-term healthcare facility within 1 year before the culture date; or the presence of a permanent indwelling catheter or percutaneous medical device at the time of culture; or previous isolation of methicillin-resistant S. aureus (MRSA)], and community-associated (CA) (onset of infection in the community or <48 h after hospital admission with none of the above risk factors) [5, 9]. About 20% of patients undergoing surgery acquire at least one nosocomial infection, leading to increased morbidity, mortality, hospital stay and costs [10-15]. Hospital treatment often requires that first line barriers for pathogens, of which skin is the most important one, are intentionally breached, resulting in an increased risk of infections. S. aureus is a predominant cause of endemic nosocomial infections, and is also responsible for large numbers of outbreaks of HA infections. Using hospital discharge data and infection surveillance (NNIS) system during 1999–2000, infections with S. aureus occur with an incidence of 9.13 per 1000 hospital discharges in the USA [16]. In a study from Calgary Health Region, approximately 29% of all nosocomial S. aureus infections were respiratory, 18% were associated with intravascular catheters, 18% arose from skin or soft tissue, and 13% represented bacteremiae without an identified source [17].

MRSA is associated with higher mortality, morbidity and financial costs compared to methicillin-sensitive *S. aureus* (MSSA) [14, 18-20]. MRSA is today accounting for 20–60% of all *S. aureus* infections in many countries and has thus become a great burden in most parts of the world [21]. In Europe, the prevalence varies considerably between geographic areas and countries from <1% to 50%, as shown in (Figure 1) [22].

Even though the percentage of MRSA among *S. aureus* isolates seems to stabilise, or even decrease in some European countries, MRSA remains a public health problem, since the proportion of MRSA is still above 25% in more than one fourth of the reporting countries

[23]. The Nordic countries and the Netherlands, are considered low-endemic countries regarding MRSA, as the frequency in bacteremia cases has remained <1 to 5%. However, since the late 1990s a substantial increase in the number of persons found MRSA positive has been observed in Norway as well as in other low endemic countries [2, 22, 24].

1.2 S. aureus carriage precedes infection

In about 80% of the cases, *S. aureus* infections are caused by the carrier strain already present on the skin or mucosa of the patient [25, 26]. *S. aureus* nasal carriage has been identified as a risk factor for the development of nosocomial infections among surgical patients [27, 28], patients on haemodialysis or continuous peritoneal dialysis [29-31], patients with liver cirrhosis and after liver transplantation [32-34], as well as HIV positive patients and patients admitted to intensive care units [35-38]. Previous studies have shown a three to six fold increase in risk of acquiring a nosocomial *S. aureus* infection in patients who are *S. aureus* nasal carriers with a large bacterial load versus non-carriers, or those with a low bacterial load

[5, 31, 39, 40]. A causal relation between *S aureus* nasal carriage and infection is supported by the fact that the nasal *S aureus* strain and the infecting strain share the same phage type or genotype [25, 26, 31] and that also eradication of *S. aureus* from the nares has proved to be effective in reducing the incidence of infection with the bacteria [27, 41-45]. Thus, prevention of the carrier state may provide new potential targets for prevention of infection.

1.3 S. aureus nasal carriage patterns

Staphylococcus aureus is part of the normal flora of humans and can also be found in other mammals as well as in birds [4]. In humans, the anterior nares are the most consistent sites of *S. aureus* colonization [3, 46]. Extra-nasal sites that typically harbour the organism include the skin, perineum, and throat [3, 47-49]. Other carriage sites including the gastrointestinal tract and vagina harbour *S. aureus* less frequently [1, 3]. Several studies have suggested that colonization of the throat is more prevalent than colonization of the anterior nares [50-53]. However, as decolonization of the nose usually has a decolonizing effect on skin and perineum, the nose is considered to be the major site of *S. aureus* colonization [42, 54, 55].

Most studies on S. aureus nasal carriage have used a cross-sectional study design with a single nasal swab culture to classify an individual as a carrier or not. However, based on longitudinal studies with repeated samples the population has been categorized into three S. aureus nasal carriage patterns: The persistent carriers, ~20% of individuals (range 12–32%), the intermittent carriers, ~30% of individuals (range 16–70%) and the non-carriers, ~50% (range 16–69%) [1, 46, 56-59]. The proportions of intermittent and non-carriers have a wide range, resulting from differences in culture methods, populations studied and interpretation guidelines [60]. The definition of persistent nasal carriage varies from study to study. There is an ongoing debate on how many cultures should be taken, at which interval, and the number or proportion of positive cultures to define persistence. One study has proposed a "culture rule" that combines qualitative and quantitative results of two nasal swabs taken with a 1week interval to accurately classify S. aureus nasal carriage [57]. The mean number of colony forming units (CFU) has been reported to be higher in persistent carriers than in intermittent carriers [61], resulting in an increased risk of infections [40, 62] and of spreading staphylococci to the surroundings [63]. It has also been shown that the genotypes of S. aureus isolated from repeated cultures differ more often among intermittent carriers than among persistent carriers [56]. This indicates that there may be differences in the determinants of

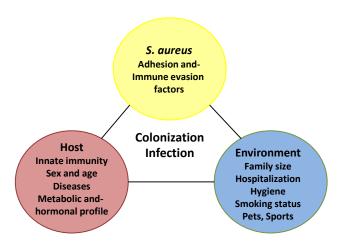
persistent and intermittent carriage. Recently, a reclassification of the *S. aureus* nasal carriage state has been proposed; the persistent carriers and the others (intermittent carriers) [64]. The proposed reclassification was based on results where intermittent carriers and non-carriers shared the same antistaphylococcal antibody profiles and responses to inoculation with a *S. aureus* mixture, as well as the previously described higher risk of infection among persistent carriers than in intermittent and non-carriers [25, 26, 62]. The study participants were first undergoing *S. aureus* eradication and then artificially inoculated with a mixture of different *S. aureus* strains. The originally persistent carriers were found to become carriers again with their original strain from the inoculation mixture, while the others (non-carriers and intermittent-carriers) quickly eliminated *S. aureus* cells from their nares [64, 65].

1.4 Determinants of S. aureus nasal carriage

Persistent nasal carriage of *S. aureus* has a high prevalence of 20–30% in healthy adults and is a major risk factor for infections with the bacterium. Nasal carriage of *S. aureus* is characterized by a subclinical inflammatory response that is insufficient to remove *S. aureus* from the nares [66, 67]. It seems as multiple mechanisms are involved in *S. aureus* nasal carriage, and that there is a fine-tuned interaction between the microbe and the host [68]. Host susceptibility factors (e.g. conditions influencing the immune response, or serious underlying diseases), environmental factors (e.g. crowding, hospitalization, and current smoking) and bacterial factors (e.g. cell-wall associated proteins, toxins and bacterial resistance mechanisms) may play important roles (Figure 2). The relative importance of these factors involved in *S. aureus* nasal carriage is largely unknown. Nevertheless, it has been suggested that host factors play a key role, as the overall picture is that in principle all *S. aureus* strains can be tolerated as a human commensal given the proper circumstances [68]. However, these primarily host-defined circumstances are still largely unknown. On the other hand, bacterial factors may decide which strain is carried rather than the carriage status as Peacock et al demonstrated that most mothers carry the same strain as their infants [69].

Mechanisms for establishment and maintenance of nasal carriage need further elucidation [1, 46]. There is a constant shedding of squamous epithelial cells and mucus from the nose that leads to a constant clearance of *S. aureus* cells. To compensate for the mechanical removal, the bacterium needs to be able to adhere to the nasal squamous

epithelium and to proliferate [70]. In addition, the host's immune defences must be evaded for *S. aureus* to become a persistent colonizer.



Staphylococcacea. *S. aureus* was discovered in 1880 by the surgeon Sir Alexander Ogston. He systematically viewed stained slide preparation of pus from patients with postoperative wound suppuration and abscesses under a microscope and observed grape-like clusters of bacteria, which he therefore named Staphylococcus from the Greek expression staphyle ("a bunch of grapes") [72]. In 1884, Rosenbach was able to isolate and grow these bacteria from abscesses and called them *Staphylococcus aureus* because of the yellow-orange or "gold" pigmented appearance of the colonies, "aureus" meaning golden in Latin [73] . *S. aureus* is part of the genus Staphylococcus, which currently contains 47 species and 24 subspecies species (http://www.bacterio.cict.fr/s/staphylococcus.html, accessed 21. Sept. 2012). *S. aureus* is by far the species most pathogenic to humans within the genus.

Traditional identification of *S. aureus* is based on morphological characteristics and biochemical tests. Staphylococci have a Gram positive cell wall with a diameter of 0.7-1.2 µm. *S. aureus* is a facultative anaerobe that grows most rapidly under aerobic conditions and

in the presence of CO₂. Colonies of *S. aureus* are β-hemolytic due to the production of several hemolysins: α -toxin, β -toxin, γ -toxin, and δ - toxin. The species is catalase positive, coagulase positive, and produces pigments (carotenoids) under aerobic conditions. *S. aureus* contains free coagulase enzyme (staphylocoagulase) and bound coagulase (clumping factor), a cell surface-associated fibrinogen-binding protein [74, 75]. Staphylocoagulase (free coagulase) is encoded by the coa gene and causes fibrinogen polymerization and clotting of plasma. Clumping factor encoded by the clfA gene, can directly convert fibrinogen to insoluble fibrin and cause the staphylococci to clump together. Staphylococci can grow in a wide pH range (4.8-9.4), resist desiccation for several weeks, and can survive at temperature extremes as high as 60°C for 30 min. In addition, *S. aureus* grows in high-salt medium due to the production of osmoprotectants [76], and can tolerate 7.5–10% NaCl.

Polymerase chain reaction (PCR) testing is not yet routine practice for daily characterization of *S. aureus* isolates, but its use is becoming more widely available in clinical settings as well as in research. One of the most reliable PCR tests for identification of *S. aureus* detects the presence of the thermonuclease gene, nuc [77]. PCR can also be used to determine the presence of the methicillin-resistance genes *mecA* and others.

Host specificity and host range

In addition to human colonization, *S. aureus* is also known to colonize and infect both pets and livestock, including dogs, cats, rabbits, horses, cattle and pigs [78]. A major concern is the presence of MRSA in pets and livestock, as these may serve as reservoirs for human colonization, exemplified by ST398 from pigs [4, 79]. Various genetic analyses have shown that lineages of *S. aureus* are not so commonly found in animals, and vica-versa. However, there is also an exchange of strains between the reservoirs. Furthermore, livestock-associated and human-associated strains share virulence factors, but have also distinct virulence factors that appear to be important in host adaptation, supporting that there is an exchange of genes encoding virulence factors between strains from livestock and humans. These factors may expand the host range and thereby threaten public health [4].

The genome

Genome sequencing of *S. aureus* has enabled investigators to explore questions of virulence, resistance, physiology, host interactions, and the microbe's success as a bacterial pathogen. The genome size of *S. aureus* typically varies from 2.5 to 3.1 megabase (Mb), and contains

~2,500 open reading frames. The first *S. aureus* genome sequences were published in 2001 by Hiramatsu's group comparing the genomes of two methicillin-resistant strains, N315, Mu50 [80]. Today, full genome sequencing has become commonly used in research, and the number of sequenced genome drafts has exploded in recent years, however only a subset of these are fully annotated and completed [81]. The *S. aureus* genome consists of 1) 80% core genes, conserved between different lineages, and 2) 20% accessory genes with mobile genetic elements (MGEs).

The core genome contains genes vital to cell survival, including genes for surface proteins involved in adhesion and surface architecture as well as genes encoding essential metabolic and regulatory functions. Within the core genome are core variable (CV) regions containing genes with a higher nucleotide substitution rate than the more stable core genes and often showing variation associated with lineage [82]. The core variable regions often encode regulators of virulence genes or surface proteins involved in host interactions during nasal carriage, such as global virulence regulations (accessory gene regulator [agr], the target of RNAIII-activating protein [trap] and staphylococcal accessory regulator T gene [sarT]) known to regulate expression of surface proteins including Staphylococcal protein A (spa) [82].

The accessory genome is assembled from mobile genetic elements (MGEs) that are integrated throughout the genome and carry about 50% of known *S. aureus* virulence factors. These elements include plasmids, bacteriophages, pathogeniticy islands, transposons and insertion sequences, and they are capable of horizontal transfer between strains. There is an exchange of virulence factors between strains contributing to adaption of clones specialized for infection of selected hosts or environments [83, 84].

Molecular typing

By use of molecular typing techniques, the spread of clones in hospitals and in the community can be identified and kept under surveillance. In outbreak situations, typing of bacteria is important for resolving transmission routes and thus for infection control. For epidemiologic surveillance, typing systems reveal the prevalence of different clones in the population in different geographical areas [85]. Today, a range of techniques are in use for typing of staphylococci, with different strengths and weaknesses. One of these methods uses the *spa* gene which encodes the *S. aureus* specific surface protein A in the core variable genome. Sequence-based typing of the *spa* gene has a relatively high discriminatory power and can be

used both for outbreak investigations as well as population studies due to slow accumulation of point mutations and relatively fast changes in repeat numbers [86]. The typing method uses the region X of the *spa* gene containing a variable number of mainly 24–27bp tandem repeats. Repeats are assigned a numerical code according to the actual sequence and the *spa* type is deduced from the repeat succession (Figure 3). The *spa* repeat and *spa* type annotations are mediated at a central localized internet server that ensures the maintenance of unique annotations by the RidomStaphType software [87]. The method has been demonstrated to be highly reproducible between laboratories. The recognized *spa* types may be grouped into clusters, *spa* CC groups, using the Based Upon Repeat BURP algorithm [88].

Other methods in use are the pulsed field gel electrophoresis (PFGE) that became the "gold standard" in typing of MRSA through the 1990s. PFGE has a high discriminatory power that makes it excellent for investigation of person-to-person transmission in a restricted time-frame; e.g. outbreaks [89]. However, PFGE has disadvantages regarding long-term analysis as genetic variations are expected. Furthermore, the comparison of inter-laboratory results are difficult and no common nomenclature exists [90].

Multilocus sequence typing (MLST) is another important typing method using the sequences of internal fragments of seven house-keeping genes in S. aureus (arcC, aroE, glpF, gmk, pta, tpi, and ygiL). Sequence variation within these genes, which occurs primarily as a result of point mutations [91], provides an allelic profile that defines the sequence type (ST) and a determination of long-term genetic variation and evolution. Related sequence types can be grouped into clonal complexes (CC) using the eBURST analysis (www.MLST:net) [92-94]. MLST is today one of the most frequently used molecular typing methods in evolutionary epidemiology, but MLST does not have the discriminatory power to be used in S. aureus outbreak situations [95]. Several other methods have been applied for typing of S. aureus, including variable number of tandem repeat (VNTR) methods and amplified fragment length polymorphism (AFLP). Microarrays can also be used for population analysis and smaller DNA microarrays have been developed focusing on detection of genes associated with virulence, antimicrobial resistance or adhesion [96-98]. Whole genome sequencing has an extremely large discriminatory power, and has been proven to be a valuable research tool [99]. The main challenge is the need for data interpretation. A recent method is the matrixassisted laser desorption/ionisation time of flight mass spectrometry (MALDI-TOF-MS), which analyses surface-associated proteins by mass spectral analysis and can be used on intact bacterial cells [100]. However, it is so far not clear which role this method will have when it comes to bacterial typing.

The various typing methods differ with respect to discriminatory power, accuracy, reproducibility, costs and technical challenges. The choice of method will depend on the study design and hypothesis. For local studies of population structure and short-term outbreaks, it is advantageous to use a method with relatively high discriminatory power, such as *spa* typing, whereas for global population surveys and long-term studies, methods based on stable housekeeping genes (such as MLST) may be preferred [95].

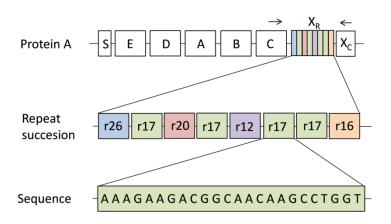


Figure 3. The principle of *spa* typing. The VNTR repeat region X_R of Protein A is the basis for *spa* typing. This region consists of a number of short repeats, and the number of repeats as well as their order determines the *spa* type. The particular repeat succession in the figure represents *spa* type t003. Arrows indicate the primers used in *spa* typing (PhD thesis, ISBN 978-82-7589-370-1, Jan 2013 of Sangvik M). Used with permission.

Population structure and invasiveness of S. aureus

Most *S. aureus* isolates (colonizing as well as invasive isolates of MSSA and MRSA) have been placed in five major, universally occurring clusters: clonal complex(CC) 8, CC30, CC5, CC22 and CC45 using the MLST typing method [91, 93, 101-103]. A similar analysis was performed using AFLP and this demonstrated that all strains fell into three major and two minor clusters, much like the MLST analysis [104]. Also, different studies have shown no distinction between colonizing and invasive isolates [82, 91, 104]. However, subclusters of strains with different degrees of pathogenicity have been observed, suggesting that the presence of accessory genes apart from the core genome of *S. aureus* may enhance or reduce the pathogenic potential of a given clone [104, 105]. Important examples of this phenomenon are genes giving rise to antimicrobial resistance, since the ability to overcome antimicrobial therapy gives a microorganism a selective advantage in hospitals or other settings where antibiotics are frequently used [68, 106]. Furthermore, MRSA has been found in all the major

clusters, suggesting that acquisition of the *mecA* gene has occurred across distinct phylogenetic subpopulations [68, 103, 107].

Geographic diversity

One may question whether the distribution of *S. aureus* genotypes from various geographical locales differs significantly or, alternatively, is rather similar. A study of Melles et al of nonclinical isolates showed that the same genotypes were identified both among individuals from USA (N = 391) and from The Netherlands (N = 829) [108]. The AFLP clusters II and III, which represent MLST CC30 and CC45, respectively, accounted for 46.6% of all carriage MSSA isolates, which underlines that these two clonal complexes have evolved to be very successful in colonizing humans. These findings are also supported by Grundmann et al [109]. They collected 2,890 clinical MSSA and MRSA isolates from blood cultures from 357 laboratories in 26 countries. The MSSA *spa* types showed a high degree of diversity with extensive geographic distribution compared with the MRSA *spa* types which displayed relatively more geographical clustering. Nevertheless, data from Indonesia have shown that one of the major AFLP classes, AFLP cluster II, as identified both in the USA and in Europe was non-existent in that East region [110]. Whether these findings are caused by a certain level of host resistance needs to be addressed in further studies [68].

Bacterial factors possibly influencing nasal carriage.

Many microbial features have been implicated in the host microbe interaction. *S. aureus* lineages have individual combinations of surface proteins involved in adhesion as well as secreted proteins involved in immune response evasion [111] (Figure 4). During *S. aureus* colonization, the individual combinations of adhesion and immune evasion factors as well as their expression levels may be of importance.

Adhesion factors

Wall teichoic acid (WTA) of *S. aureus* is suggested to play an important role in attachment, both in the early stages and for continued colonization [112, 113]. In addition, a class of cell wall-associated proteins termed microbial surface components recognizing adhesive matrix molecules (MSCRAMMs) [114] may have critical roles at a later stage of colonization of the nose [115]. In vitro experiments show that *S. aureus* can directly adhere to the keratinized squamous epithelial cells in the anterior nares via cell wall anchored clumping factor B

(ClfB). S. aureus ClfB binds to cytokeratin 10 which is a component of the squamous cell [116]. Also, the iron-regulated surface determinant A (IsdA) protein of S. aureus can bind to cytokeratin 10, loricrin and involucrin, important proteins of the matrix surrounding the upper anucleated layers of the epithelium [117]. ClfB and IsdA have both been demonstrated to promote colonization of the nares of rodents in in vivo models [118, 119] and were expressed during nasal colonization in humans [120]. S. aureus surface protein G (SasG) and serine-aspartic acid repeat proteins SdrC and SdrD are other bacterial surface proteins probably contributing to adhesion to nasal epithelial cells [121, 122].

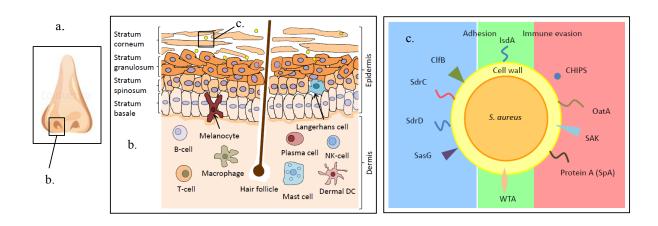


Figure 4. a. The nose with the vestibulum nasi **b.** The epidermis with the layers of keratinocytes: stratum corneum, stratum granulosum, stratum spinosum and stratum basale. During *S. aureus* colonization, *S. aureus* can be found in the epidermis. The immune cell type, Langerhans cell, is found in the epidermis. The dermis includes several immune cells such as natural killer (NK) cells, macrophages, T-cells, B-cells, mast cells, dermal dendritic cells (DC) and plasma cells. **c.** *S. aureus* exhibits adhesion factors (blue background), factors involved in immune evasion (pink background), and factors influencing both adhesion and immune evasion (green background) during nasal colonization.

Abbreviations: ClfB, clumping factor B; SdrC, serine-aspartic acid repeat protein C; SdrD, serine-aspartic acid repeat protein D; SasG, *S. aureus* surface protein G; CHIPS, chemotaxis inhibitory protein of *S. aureus*; OatA: O-acetyl transferase, SAK, staphylokinase; *spa*, protein A; WTA, wall teichoic acid, (Modified from PhD thesis, ISBN 978-82-7589-370-1, Jan 2013 of Sangvik M). Used with permission.

Immune evasion factors

S. aureus can produce a large variety of secreted proteins involved in immune evasion. Some of the proteins target immunoglobulins, complement or neutrophil recruitment, whereas others counteract the effects of antimicrobial molecules such as lysozyme and defensins. Nasal secretion is the first line of host defence against inhaled bacteria. The nasal secrete is a complex mixture of proteins, sugars and salts, containing e.g. lysozyme and immunoglobulins IgA and IgG [123], as well as defensins [124] and complement proteins [125]. *S. aureus* is

resistant to lysozyme due to the cell wall modifying enzyme O-acetyltransferase (OatA) in combination with WTA [126]. In a study of *S. aureus* isolated from persistent nasal carriers, several factors, including staphylococcal protein A (*spa*), staphylokinase (SAK) and chemotaxis inhibitory protein of *S. aureus* (CHIPS) were expressed [120]. *Spa* is able to limit opsonisation by binding to the Fc-region of IgG in a conformation that inhibit recognition by the neutophils [127]. Through this IgG-binding, *spa* also interferes with binding to the complement system [128]. SAK and CHIPS are suggested to inhibit immune response in different ways [129-131].

Nasal microflora

The availability of resources (e.g. nutrients and attachment sites), the presence of harmful substances, and the host's immune responses can be influenced by the presence of established bacterial communities in the nose [132] and determine the colonization success of different bacteria.

The microbial ecology of the vestibulum nasi is complex. The nares are colonized by a temporally stable microbiota that by culture-independent approaches in healthy adults consists primarily of the phylum Actinobacteria (e.g., *Propionebacterium* spp, and *Corynebacterium* spp), Firmicutes (*Lactobacillae spp* and *Staphylococcus* spp) and Proteobacteria (*Enterobater* spp) [133].

A persistent carrier seem to be protected from acquiring new strains of *S. aureus*, e.g. during hospitalization, also known as colonization resistance. This was exploited in the 1960s, to protect infants of hospital acquisition of virulent strains of *S. aureus* [134, 135]. The colonization resistance is reduced when carriers are treated with antibiotics [135, 136]. It has also been shown that MSSA nasal carriage interferes with and hence may protect against MRSA acquisition [137].

The prevalence of *S. aureus* carriage has previously been found to be lower among those colonized with corynebacteria, but the underlying mechanism is not known [138, 139].

Frank et al observed negative associations between *S. aureus* and *S. epidermidis* and suggested microbial competition as a cause [133]. Mechanisms of bacterial interference applied by *S. epidermidis* may also involve production of phenol-soluble modulins (PSMs) [140, 141] that induces antimicrobial effects against *S. aureus*, peptide pheromones [142, 143] and induction of human β-defensins [144]. Furthermore, a serine protease (Esp, 27kDa) secreted by a subset of *S. epidermidis* has been reported to inhibit *S. aureus* nasal colonization

through reduced biofilm formation [145]. As *S. aureus* does not form a typical biofilm in the nasal cavity this mechanism has been questioned [146]. However, the Esp protease may inhibit *S. aureus* nasal colonization by removing adhesion or immune evasion factors essential for colonization [70].

Bogaert et al noted a negative correlation for co-colonization of *S. aureus* and vaccine-type of pneumococci but not for *S. aureus* and non-vaccine type pneumococci in the nasopharynx of children [147]. However, a study in children did not reveal an increase in prevalence of *S. aureus* colonization after introduction of the 7-valent pneumococcal-conjugate vaccine (PCV7) [148]. It has been proposed that the displacement of *S. aureus* by *S. pneumonia* in nasopharynx may be explained by H₂O₂-mediated bacterial interference [149], but this has not been confirmed by others [150].

Another study has concluded that nasal microbiomes may be grouped into 12 supergroups, with *S. aureus* present in 2 but absent in the others [151]. In contrast, Frisoni ED et al [152] found in nasal metagenome analyses that the microbial diversity was similar in both *S. aureus* carriers and non-carriers which may imply that *S. aureus* seem to come in addition to the other normal flora, and that large-scale carriage eradication is discouraged [152].

1.4.2 Host factors

Although the role of host factors in nasal carriage of *S. aureus* has been extensively studied, the host-defined circumstances are still somewhat unclear.

The results from studies of host genetic factors on nasal colonization are not consistent, suggesting that the role of heritability is modest [153].

S. aureus nasal carriage rates vary by ethnic groups, with higher rates among Caucasians [66, 154]. Previous studies have consistently found increased carriage rates associated with male gender [154, 155], younger age [47, 68, 69, 154], and oral contraceptive use [156]. Children have higher persistent carriage rates than adults [47, 157]. The colonization rate declines from approximately 45% during the first 8 weeks to about 21 % by 6 months [69]. A transition zone from persistent carriage to intermittent carriage or non-carriage has been proposed, which implies development of the immune response during maturation from childhood to adolescence [3, 47]. Patients with chronic skin diseases [68, 158, 159], HIV/AIDS [36, 38], end stage renal disease [31, 160] and end stage liver disease

[32, 34] are at increased risk of *S. aureus* nasal carriage supporting the view that impaired immune responses may increase the risk of carriage. There are only a few reports on the association between measured biomarkers, as metabolic and hormonal factors, and *S. aureus* nasal carriage in the general population; diabetes mellitus (DM) [161, 162], obesity [154, 155], and vitamin D deficiency [163], have been positively associated with *S. aureus* nasal carriage. Also, polymorphisms in the glucocorticoid receptor gene increasing the sensitivity and endogenous levels of glucocorticoid hormones were positively associated with *S. aureus* nasal carriage [164]. However, long-term cortisol levels determined in hair segments of 72 healthy individuals were not associated with the carrier state [165].

Host immunity and S. aureus carriage

S. aureus predominantly colonizes the anterior human nares in an area covered with ordinary skin supplemented with nasal secretion. Nasal secretions are a part of the host defence against microbes, and it has been shown that nasal fluids from non-carriers were bacteriostatic or bactericidal, whereas the nasal fluids from carriers allowed growth of S. aureus [166], and it has been proposed that the presence of haemoglobin in nasal secretions promotes S. aureus carriage through inhibition of the agr system [167]. The epidermis contains several antimicrobial lipids, peptides and proteins provided by keratinocytes, sebocytes, mast cells, and eccrine sweat glands and also by circulating neutrophils or natural killer cells that are recruited to the skin [168]. The individual or combined expression patterns of the various antimicrobial molecules may influence the colonization status of S. aureus [169]. Lipids in epidermidis have antimicrobial activity against S. aureus, and a reduction of synthesized fatty acids has been found to be associated with S. aureus carriage in atopic dermatitis patients [170, 171].

Epidermal keratinocytes shape the physical barrier of the skin and contribute to innate immunity. Professional innate immune cells, such as dendritic cells (DCs) and macrophages reside in the skin ready to respond to bacterial invasion [172]. The keratinocytes and cells involved in the innate immune system sense pathogens by expressing pattern recognition receptors (PRRs) that recognize pathogen-associated molecular patterns (PAMPs) [173]. PAMPs are evolutionary conserved microbial components, including lipopolysaccharide (LPS), peptidoglycan, flaggelin and nucleic acids [172]. The bacterial cell wall of *S. aureus* is composed of multiple peptidoglycan layers in combination with WTAs, LTA, and various MSCRAMMs or other substances that can be recognized as PAMPs [174, 175]. The most

important PRR known to be involved in recognizing *S. aureus* is the Toll-like receptor (TLR) 2 [174, 176] as well as the intracellular nucleotide-binding and oligomerization domain (NOD)-like receptors (NLRs) NOD2 and NLRP3 [174, 176]. Activation of PRRs induce intracellular signalling resulting in altered gene expression with the final result of increased expression and secretion of various antimicrobial peptides (AMPs), cytokines and chemokines with initiation of innate and adaptive immune responses which promote killing of *S. aureus* [177, 178].

AMPs have the capability to kill various pathogens and modulate innate and adaptive immune functions and are secreted from keratinocytes, other resident cells in the skin (e.g cells in eccrine glands, mast cells, sebocytes) and invading immune cells (e.g. neutrophils, NK cells) [179-181]. Two important and well studied AMP gene families in the skin are the defensins and the cathelicidin [182, 183]. The human β -defensins expressed in mucosa and epithelial cells, have been compared in their antimicrobial activity against *S. aureus*, and the most potent is the β -defensin 3 (HBD3) followed by β -defensin 2 (HBD2) and 1 [184-186]. A higher induction of β -defensin 3 is associated with a better clinical course and outcome of *S. aureus* skin infection and the level of both constitutive and induced β -defensin 3 is lower in persistent *S. aureus* carriers than non-carriers [187, 188]. Also, the Cathelicidin and its active peptides LL-37 and others, have all antimicrobial activity against *S. aureus* [189].

Vitamin D

Vitamin D was discovered in 1922 and has, because of its effect on bone metabolism, for decades been used in prevention and treatment of rickets in children and osteoporosis in adults [190, 191]. Vitamin D is a fat soluble vitamin, which exists in two forms, ergocalciferol (vitamin D₂) and cholecalciferol (vitamin D₃) [190]. The two sources of vitamin D are diet and sun exposure [191]. Dietary sources include fatty fish, cod liver oil, egg yolk, mushrooms, fortified products and supplements [191]. Solar ultraviolet B (UVB) radiation at wavelength 290-320 nm induces conversion of 7-dehydrocholesterol to pre-vitamin D₃ in skin, which under normal temperature conditions isomerizes to vitamin D₃ [192]. The relative importance of vitamin D from UVB exposure and vitamin D from dietary sources varies between populations and is affected by ethnicity/skin pigmentation, climate conditions, dietary habits and cultural practices [193, 194].

Although vitamin D is a vitamin by name, the molecular structure is that of a secosteroid [195]. Physiologically it acts more like a pre-hormone as it is not biologically

active until it has been metabolized [195]. In the liver vitamin D is converted to $25(OH)D_2$ or $25(OH)D_3$ by 25-hydroxylase [190]. In the following these will be referred to as 25(OH)D. In the kidneys 25(OH)D is further converted to 1,25-dihydrovitamin D (1,25(OH)₂D) by 1α -hydroxylase [190]. This renal conversion is tightly regulated by calcium and phosphate levels. However, the 1α -hydroxylase is also present in many other tissues throughout the body [196]. This extra-renal conversion of 25(OH)D to $1,25(OH)_2D$, which is thought to be regulated by local growth factors and cytokines, is dependent on the amount of substrate available and works locally in an autocrine or paracrine fashion [190].

Ligand binding to the vitamin D receptor (VDR), a member of the superfamily of nuclear receptors for steroid hormones, is necessary for the biological activities of 1,25(OH)₂D and 25(OH)D [190]. The most potent metabolite 1,25 (OH)₂D binds to the VDR with high affinity while 25(OH)D binds to the receptor nearly 100 times less avidly [190]. The VDR regulates the transcription of several target genes in a variety of vitamin D target cells.

Polymorphisms of the VDR have been associated with insulin resistance, decreased bone density, infections, *S. aureus* nasal colonization among type 1 diabetes patients, autoimmune diseases, cancer and resistance to vitamin D therapy [197-200]. Several functional VDR polymorphisms have been found [201], but the field is still under exploration [190].

Measurement of vitamin D

The preferred biomarker for an individual's vitamin D status is 25(OH)D, the major form in the circulation. This is due to its high stability in stored serum and plasma samples, a characteristic that makes accurate, long-term epidemiological studies possible [202]. Serum 25(OH)D reflects the amount of vitamin D ingested from food and produced in the skin during UVB exposure. 1,25(OH)₂D does not enter the circulation in large amounts, but as the local conversion is dependent on the 25(OH)D available, measuring serum 25(OH)D is a good alternative [190]. 25(OH)D is measured by use of immunoassays or chromatographic methods. High throughput automated immunoassay methods are the most commonly used in large population and clinical studies [203]. However, the immunoassay methods are prone to performance change over time and have varying ability to distinguish between 25(OH)D₂ and 25(OH)D₃ in contrast to the chromatographic methods [203, 204].

The variability of 25(OH)D levels between individuals is explained by both environmental factors and heritability [205], and several polymorphisms have been identified

as important determinants of serum 25(OH)D levels [201]. There is no consensus as to what is the optimal serum concentration of 25(OH)D, but a serum concentration <50 nmol/l is considered as deficient as this level is associated with an increase in Parathyroid hormone (PTH) level [206] and decrease in physical performance among elderly [207]. Concentrations between 50–75 nmol/l are considered as insufficient [191] and a recent consensus panel recommended that a serum concentration 75–100 nmol/l for different health outcomes should be targeted [208, 209]. Very high levels of serum 25(OH)D also seem to be disadvantageous. Data from the National Health and Nutrition Examination Survey (NHANES) III showed a lower risk of mortality at levels of 75–125 nmol/l, but a higher risk of mortality among women at levels higher than 125 nmol/l [210]. On the other hand, serum 25(OH)D levels >150 nmol/l have been found in healthy populations living in areas close to equator and spending much time outdoors [211, 212], and currently 250 nmol/l is considered the upper physiological limit [213]. In the Nordic countries the recommended intakes of vitamin D supplementation are 400 IU daily for infants, elderly, pregnant and lactating women, and 300 IU daily for all others 2–60 years [214].

Vitamin D and risk of infection and bacterial colonization

As the VDR and the 1α hydroxylase are found in many tissues and cells in the body [196], serum 25(OH)D levels have been proposed to influence risk of several common diseases including infections and *S. aureus* nasal colonization [163, 191, 196].

Several studies suggest that vitamin D has a protective role in respiratory tract infections where viruses represent the most common pathogens. Seasonality in the occurrence of influenza and respiratory tract infections has been attributed to low wintertime vitamin D levels [215-217]. Recently, inverse associations between 25(OH)D concentrations and incidence of respiratory tract infections with thresholds of 25(OH)D \geq 75 nmol/l [218] and \geq 40 nmol/l [219] has been observed.

The associations between vitamin D and bacterial infections have been addressed in different studies. Epidemiological data have established that vitamin D deficiency is associated with increased *Mycobacterium tuberculosis* (TB) prevalence and susceptibility to active TB disease [220-224]. *In vitro* studies have also proved that vitamin D₃ has inhibitory activity on strains of *S. aureus*, *Streptococcus pneumonia*, *Klebsiella pneumonia* and *Escherichia coli*. In the presence of 50,000–90,000 IU/mL of vitamin D₃, the organisms were killed or demonstrated marked growth inhibition [225]. Furthermore, pneumococcal

infections have been shown to increase each winter and extended periods of low UV radiation have been related to invasive pneumococcal disease [226, 227]. Supplementation with oral vitamin D₃ has been observed reducing the risk of a repeated episode of pneumonia among children in Kabul [228].

In the US National Health and Nutrition Examination Survey (NHANES) 2000–04 including 14,000 women and men, vitamin D deficiency was associated with an increased risk of nasal carriage of MRSA but not MSSA [163]. Furthermore, a study from a diabetic clinic of Heraklion, Crete, Greece, reported possible associations between VDR polymorphisms and nasal carriage of *S. aureus* among 93 type I diabetes patients aged 3–25 years [200], but a population-based cohort study from Rotterdam, Netherlands, including more than 2000 healthy elderly individuals did not observe any associations with VDR polymorphisms [229]. Moreover, vitamin D deficiency has been linked to adverse outcomes in veterans with *Clostridium difficile* and MSSA infections [230]. However, there is still limited knowledge of the possible relationships between serum vitamin D levels and *S. aureus* nasal carriage in an adult general population, also considering possible age and gender interactions.

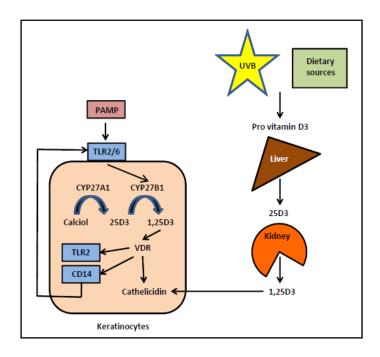


Figure 5. Keratinocyte. Mechanisms of vitamin D₃ activation and cathelicidin response. Extrarenal metabolism of vitamin D₃ by keratinocytes provides a system for rapid control of cathelicidin expression. Activation of calciol to 25D₃ and 1,25D₃ requires two hydroxylations steps that occur sequentially in liver and kidney as well as in keratinocytes who expresses CYP27A1 and CYP27B1.1,25D₃ binds to and activates the vitamin D receptor (VDR) which subsequently activates transcription of cathelicidin. Based on [231].

Vitamin D and immune response

Vitamin D has modulatory effects on both innate and adaptive immunity, which may influence susceptibility to infection and bacterial colonization [163, 232]. Nearly all immune cells display a specific vitamin D receptor (VDR) including B and T lymphocytes, monocytes and dendritic cells [233]. The capacity of 1,25(OH)₂D₃ to modulate cytokine responses to a Th2 signalling pattern and induce AMP production are important biological effects possibly protecting individuals against microbial infections and colonization. In a distinct immune regulatory role, vitamin D₃ affects the innate antimicrobial defense at epithelial barriers, such as the airway epithelium or the skin.

Keratinocytes can also activate vitamin D₃ independent of renal and hepatic hydroxylation steps [233] with the final result of increased expression and secretion of various AMPs, cytokines and chemokines leading to initiation of innate and adaptive immune responses which promote killing of *S. aureus* and other microbial agents (Figure 5).

The Cathelicidin, often referred to by its peptide form hCAP18, is stored in keratinocytes and secreted into different layers in the epidermis prossessing the protein to active peptides LL-37 and others, which all have antimicrobial activity against *S. aureus* [189]. Furthermore, LL-37 influences TLR2 signalling and CD14 expression in keratinocytes [231], which may result in increased ability to detect pathogens. All these studies support that vitamin D protects against different infections and also against *S. aureus* nasal carriage (Figure 5).

Obesity

The incidence of obesity worldwide has increased dramatically during recent decades. Obesity and associated disorders now constitute a serious threat to the current and future health of all human populations on earth. The World Health Organization (WHO) estimates that more than 1 billion adults worldwide are overweight (body mass index [BMI] 25.0–<30.0 kg/m²), 300 million of whom are clinically obese with a BMI ≥30.0 kg/m² [234, 235]. Obesity has been associated with numerous health problems and chronic diseases, including increased risk of insulin resistance, type 2 diabetes, fatty liver disease, atherosclerosis, degenerative disorders such as dementia, airway diseases and some cancers [235-237]. These comorbidities have been attributed to hormonal and metabolic changes related to increased adipose tissue mass [235, 238].

Measurement of body fat

There are various anthropometric methods for estimating body fat. Underwater weighing based on Archimedes' principle has long been considered to be the gold standard for estimation of body fat percentage, but this is a labour-intensive method that is not feasible in larger studies [239].

Dual energy X-ray absorptiometry, or DXA (formerly DEXA), is a practical and newer method for estimating body fat percentage, and determining body composition and bone mineral density [240]. Percent visceral and subcutaneous body fat assessed by DXA has been associated with metabolic syndrome and through it DM as well as cardiovascular disease [241].

The most widely used methods for measuring adipose tissue depots are the estimation of BMI from an individual's height and weight, and the use of waist circumference (WC), hip circumference, and the waist/hip ratio. BMI is calculated as weight (kg) divided by height (m) squared (kg/m²). There are different methods for measuring WC. WHO STEPS protocol for measuring WC instructs that the measurement be made at the midpoint between the lower margin of the last palpable rib and the top of the iliac crest [242]. The United State (US) National Institutes of Health (NIH) protocol and the protocol used by the US National Health and Nutrition Examination Survey (NHANES) III indicate that the WC measurement should be made at the top of the iliac crest [243]. The NIH also provided a protocol for measurement of WC for the Multi-Ethnic Study Atherosclerosis (MESA) study. This protocol indicates that the WC measurement should be made at the level of the umbilicus [243]. Some studies have assessed the WC at the point of minimal waist [244]. A review of 120 studies concluded that different WC measurement protocols had no substantial influence on the association between WC, all-cause mortality and cardiovascular disease (CVD)-specific mortality, and risk of CVD and diabetes [244].

Regarding measurements of hip circumference, all the protocols mentioned indicate that the hip circumference measurement should be taken around the widest portion of the buttocks.

Obesity and immune response

Adipose tissue represents an endocrine organ from which a high number of proteins and hormones, so called adipokines, are synthesized and secreted [245, 246]. One of the adipokines, leptin, produced predominantly by subcutaneous adipose tissue [247], is probably

one of the best characterized links between the obesity-induced chronic low-grade inflammation and modulation of immune function [248]. In malnourished infants who have low plasma leptin, impairment of both the innate and adaptive immune response has been observed [249]. Obesity may promote a chronic low-grade inflammation that attenuates leptin signalling [248]. Along this line, it has been observed an underreactivity of the innate immune response to clear *S. aureus* invasive infection and survive sepsis in diet-induced obese mice and genetically obese Ob/Ob mice on low fat diet [250]. These findings are further supported by the observation of chronically increased leptin levels in obese mice associated with a state of leptin resistance in the central nervous system, a hallmark of the obesity-induced impaired immune response [248]. Also, leptin seems to modulate the expression of antimicrobial peptides observed for Human β -defensin-2 (HBD2) in keratinocytes [251]. Taken together, these results suggests that leptin may play a key role in antimicrobial defense, and allows us to hypothesize that leptin has a role in *S. aureus* nasal carriage as well as infections [248, 251, 252].

Obesity is often linked to elevated serum glucose concentration and type 2 diabetes [253]. Both insulin dependent- and independent DM have been associated with *S. aureus* nasal carriage and infections including also other types of microbial pathogens [68, 161, 254]. The pathophysiological basis for this association remains to be discovered. Increased blood and mucosal glucose levels may influence bacterial adherence, promote staphylococcal growth, reduce neutrophil chemotaxis, and phagocyte activation in neutrophils and macrophages, as well as impair killing of intracellular micro-organisms (including *S. aureus*) [162, 254, 255].

Obesity and insulin resistance have also been linked to changes in circulating levels of reproductive hormones [256-258]. Estrogens generally exert immune enhancing activities while androgens exert suppressive effects on both innate and adaptive immune responses [259, 260]. One may thus hypothesize that in premenopausal women, obesity may be linked to anovulatory cycles and a lower estrogen/androgen ratio with increased susceptibility to colonization [257, 261] whereas in postmenopausal women and men obesity may be linked to higher estrogen/androgen ratio and lower susceptibility to colonization [256, 258].

Obesity and S. aureus infection and carriage

Recent clinical findings indicate that obesity may be linked to increased susceptibility to infections. The various infections include community- acquired pneumonia and wound

infections, as well as nosocomial infections such as sepsis, pneumonia, surgical site infections, catheter-related infections and respiratory related hospitalizations during influenza seasons [252, 262-266]. These associations may partly reflect a mechanical dysfunction in the respiratory tract due to obesity, as well as prolonged surgery and hospitalization with increased risk of nosocomial infections.

Interestingly, former studies have observed obesity as a risk factor for *S. aureus* nasal colonization [154, 155]. However, epidemiological studies in a general population investigating the role of obesity on *S. aureus* colonization are limited, and the role of obesity per se, independent of DM, is of particular interest.

1.4.3 Environmental factors

An important determinant of *S. aureus* nasal carriage is exposure to the bacterium. *S. aureus* is acquired from sources in the environment, with human carriers as the most important source. Hands are the main vector for transmitting *S aureus* from other humans or surfaces to the nasal niche; e.g. nose picking [267]. A typical transmission route of *S. aureus* is from the nose to the hand of a person, then to a surface (e.g. a door knob), and/or via the hand to the nose of a second person. *S. aureus* may also reach the nose directly through the air, but this probably occurs less frequently [268]. However, airborne transmission is important for the dispersal of staphylococci to many different reservoirs, from where, via the hands, the microbe can reach the nose [268]. *S. aureus* nasal carriers with rhinitis can disperse high loads of *S aureus* into the environment, and may be the source of outbreaks of *S. aureus* infections [269].

Environmental factors such as crowding, transmission between household members and family size seem to be risk factors for carriage [69, 147, 270-272]. Cross-sectional surveys of healthy adults have reported a decline in *S. aureus* nasal carriage rates from 35% in 1934 to 27% in the year 2000 [1]. Explanations for this decline include improved personal hygiene, changes in socioeconomic class and smaller families [147, 273].

Activities involving close physical contact and the risk of minor injuries, such as sports, are positively correlated with *S. aureus* spread and acquisition [147, 274]. In contrast, current smoking seems to reduce the risk of *S. aureus* carriage probably due to the bactericidal activity of cigarette smoke [162, 275], and the increased immune and

inflammatory responses in smokers [162]. As pets also may be colonized with *S. aureus*, they may serve as vehicles for transmission to humans [276].

Healthcare - associated environmental exposure determinants for S. aureus nasal carriage.

Healthcare-associated environmental exposure is defined as an environmental exposure that occurs in any healthcare facility as a result of medical care.

The levels of crowding and hygiene in both hospital and household settings are important for the rate of transmission [3]. Hospitalization implying breaching of skin and/or mucosal barriers in patients has been observed to increase the risk of *S. aureus* carriage [1]. Despite various infection control strategies receiving increased attention, about 20% of patients undergoing surgery still acquire at least one nosocomial infection [1].

HCWs have been reported to have rates of *S. aureus* nasal carriage comparable to the general population in different cross-sectional studies [46, 277] but the range of carriage rates is large possibly due to differences in the quality of sampling and culture techniques as summarized by Kluytmans et al [46]. Recent reports have revealed higher *S. aureus* nasal carriage rates among surgeons than among high-risk patients groups [278], among physicians compared with other professionals in the society [279], and among nurses compared to other HCWs [280, 281]. This supports the view that working in healthcare services with substantial patient contact may be a risk factor for carriage.

Nearly 20% of *S. aureus* nosocomial infections have an exogenous origin [25, 26, 43], where HCWs may serve as an important vector. Colonized HCWs are capable of transmitting *S. aureus* to patients [1, 282, 283], and of introducing *S. aureus* into their families [270-272]. The bacterium can also be reintroduced into the hospital by intrafamilial spread from and to healthcare workers [270, 272]. The typical transmission route of *S. aureus* from HCWs to patients appears to be transiently contaminated hands of HCWs, who have acquired the microorganism from their nose, by direct patient contact, by direct contact with their family members or by handling contaminated materials [272, 282, 283].

In general, many of the studies in healthcare settings may have been biased by a lack of information on background prevalence in the relevant general population as well as in households. Thus, the role of working in healthcare settings as well as exposures in households as independent risk factors for *S. aureus* colonization in a general population may need further research.

2. AIMS OF THE THESIS

The main objectives of this thesis were to investigate whether sex, metabolic and hormonal profiles such as serum 25(OH)D levels and obesity, and environmental factors such as current daily smoking, being a healthcare worker and residing with children are associated with *S. aureus* nasal colonization and carriage in an adult general population. Also, host susceptibility and-environmental factors and their associations with *S. aureus spa* types were explored to contribute to the knowledge on possible interactions between *S. aureus* strains, the host and the environment for nasal carriage.

The aims of this thesis were:

- to examine the relationship between serum 25(OH)D concentration and *S. aureus* nasal colonization and carriage among men and women in a unselected general population.
- to investigate whether excess body weight and abdominal adiposity are associated with *S. aureus* nasal colonization independent of pre-diabetes and diabetes among women and men in an unselected general population.
- to study whether being a healthcare worker is associated with *S. aureus n*asal carriage overall and certain *S. aureus spa* types compared with non-healthcare workers and if residing with children influences the odds of *S. aureus* nasal carriage and *spa* types among women and men in an unselected general population.

3. MATERIAL AND METHODS

3.1 The Study population-The Tromsø Staph and Skin Study (TSSS)

The Tromsø Study is a longitudinal population-based multipurpose study with five previous surveys undertaken between 1974–2001. These were all focused on lifestyle-related diseases [284, 285]. The sixth survey (Tromsø 6) was carried out from October 2007 to December 2008 in the municipality of Tromsø. Based on the official population registry, residents of the municipality of Tromsø were invited to take part in the survey. The subjects who were invited included all residents in Tromsø aged 40–42 and 60–87 years (N=12,578), a 10% random sample of individuals aged 30–39 years (N = 1,056), a 40% random sample of individuals aged 43–59 years (N = 5,787), and all subjects who had attended the second visit in Tromsø 4, if not already included in the three groups mentioned above (N = 341; in **Paper III**, the number N = 295 is incorrect). Of the total of 19,762 invited, 12,984 men and women aged 30–87 years attended Tromsø 6 (65.7%). Women constituted 51.3% of the invited and 53.4% of the participants [284, 285].

The Tromsø Staph and Skin Study (TSSS) was part of the Tromsø 6. For the study of *S. aureus* nasal colonization in the TSSS, a more evenly distributed sampling across age groups was considered to be suitable and the inclusion of 4,000 observations to be sufficient for subgroup analysis of host-microbe relationships. Thus, TSSS collected nasal swab cultures during October 2007 to June 2008. The eligible group was all participants in Tromsø 6 aged 30–49 years (N = 1,730) and random samples of older participants aged 50–87 years (N = 2,629, relative distribution of birth cohorts as in the municipality). A total of 4,026 men and women aged 30–87 years (30–49 years, N = 1,597; 50–87 years, N = 2,429) had a nasal swab culture taken and were included in the TSSS at the first visit. Of these, 2,997 subjects (30–49 years, N = 1,118; 50-87 years, N = 1,879) had a repeated culture taken (Figure 6). The median interval between cultures was 28 days and for 90% of the observations the interval was \geq 12 days.

Ethics

The study was approved by the Regional Committee of Medical and Health Research Ethics, North Norway, and all attendees signed an informed consent form. Interviews, clinical examinations, nasal swab cultures, and blood samples were performed according to standardized procedures by trained healthcare personnel at the screening centre. Two self-

administered structured questionnaires covered a broad range of issues related to socioeconomic status, lifestyle, health and disease. Further information about the Tromsø Study, including invitation letter, consent form and questionnaires is available at the study web pages www.tromsoundersokelsen.no, for questionnaires, please, also see Appendices A – D.

3.1.1 Study population - Paper I-III

The study population in **Paper I** consisted of participants with either a single nasal swab culture (taken at the first visit) or double nasal swab cultures (taken at first and second visit). Participants with missing data on serum 25(OH)D (N = 60) or smoking status (N = 48) and those without valid swab cultures at the first and the second visit (first swab N = 129, second swab N = 49), were excluded. Thus, we included 3,789 participants with a minimum of one nasal swab culture for analysis of *S. aureus* nasal colonization, and 2,780 participants with two nasal swab cultures for the analysis of *S. aureus* nasal carriage (Figure 6).

The study population in **Paper II** consisted of participants who attended the first visit. Thus, the S. aureus colonization state was determined by a single nasal swab culture taken at the first visit. This decision was based on the evaluation of the agreement between culturing results in a sub-cohort of the TSSS including 2,868 participants with two valid swab cultures. These participants had made a second visit to the screening centre and had a second nasal swab culture taken after a median time of 28 days. In 90% of these participants the interval was ≥ 12 days, and only 113 of the 2,868 participants (3.9%) were misclassified as colonized from the culturing results of the first nasal swab (i.e. first swab culture positive and second swab culture negative). Among those with two positive swab cultures (N = 727), 669 participants (92%) had the same spa type in both samples. Among the 4,026 participants aged 30-87 years who attended the first visit, 129 nasal swab cultures were considered invalid due to the use of antibiotics within the last 24 hours (N = 27) or no bacterial growth in cultures (N = 27)= 102). Pregnant women (N = 15) and participants with missing height and/or weight data (N = 4) were excluded, leaving 3,878 participants for analysis of BMI. In addition, 103 participants with missing WC data were excluded, leaving 3,775 participants for analysis of WC (Figure 6).

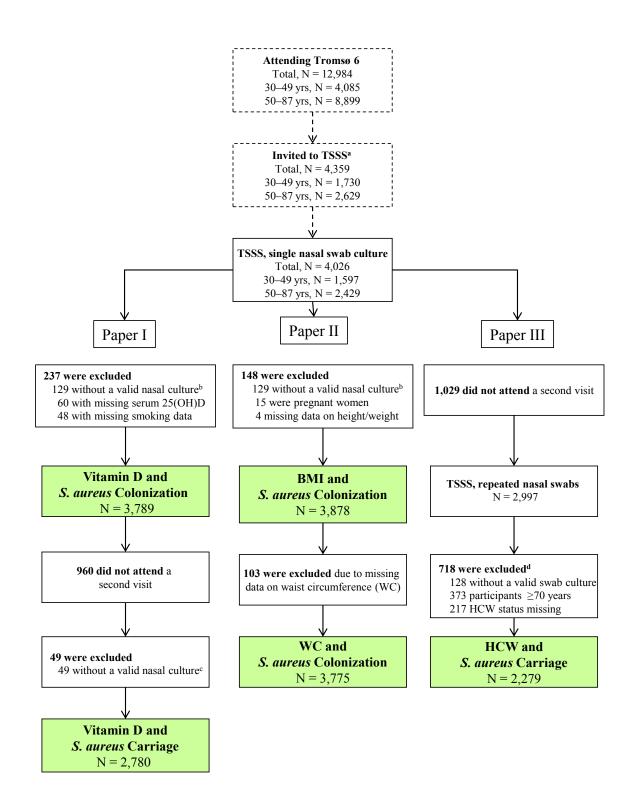


Figure 6. The study populations of The Tromsø 6 and The Tromsø Staph and Skin Study, Papers I-III.

^a Invited to The Tromsø Staph and Skin Study. Age group <50 years: all subjects. Age group 50–87 years: random samples of subjects. ^b Not valid swab culture at the first visit: 102 had no growth in swab culture and 27 had taken antibiotics last 24 hours before visit (systemic or eye drops/ointments). ^c Not valid swab culture at the second visit: 35 had not growth in swab culture and 14 had taken antibiotics last 24 hours before visit (systemic or eye drops/ointments). ^d Of the 2,997 participants with a repeated nasal swab culture, 718 participants were excluded. Abbrevations: TSSS, The Tromsø Staph and Skin Study; BMI, body mass index; WC, waist circumference; HCW, Healthcare worker.

The study population in **Paper III** consisted of participants with a double set of nasal swab cultures taken (first and second visit). Of the 2,997 participants 30–87 years who had repeated nasal swab cultures taken, a total of 373 subjects aged 70 years or more were excluded due to normal age of retirement in Norway, 217 subjects had missing data on HCW status, and 128 had no valid swab cultures. Thus, for the present analysis 2,279 participants aged 30–69 were included (Figure 6).

3.2 Measurements

3.2.1 Assessment of S. aureus nasal colonization/carriage

Both vestibuli nasi were sampled by the same NaCl-moistened sterile rayon-tipped swab and placed in Amies charcoal transport medium (Copan, Brescia, Italy). Within three days, all specimens were cultured on blood agar (Oxoid, Cambridge, UK), chromIdTM *S. aureus* and chromIdTM MRSA agars (bioMérieux, Marcy I'Etoile, France) and incubated for 42–48 hours at 37°C. If positive (green) colonies were found on the chromId plates, the most dominating colony was selected and confirmed as *S. aureus* by the Staphaurex Plus (Remel, Lenexa, KS, USA) agglutination test. All *S. aureus* isolates were frozen at –70°C in glycerol-containing liquid media. No MRSA was detected. In **Paper I** and **II** the *S. aureus* nasal colonization state was defined as positive or negative for *S. aureus* in the first nasal sample. In **Paper I** and **III**, the carrier state was based on the culturing results of two consecutive samples; carrier = two positive samples and non-carrier (intermittent carrier) = one or none positive sample [64].

3.2.2 spa typing

S. aureus isolates from frozen cultures (-70°C) in glycerol-containing medium were inoculated on blood agar (Oxoid) and incubated overnight at 37°C. 2–3 colonies were transferred to 200 μl dH₂O and vortexed. The isolates were *spa* typed using the primers *spa*-1113f and *spa*-1514r with the following cycling conditions: 95°C for 10 min; 35 cycles of 95°C for 30 s, 60°C for 15 s, and 72°C for 1 min; and 72°C for 10 min and then kept at 4°C as described previously [286, 287]. PCR products were sequenced on both strands by Macrogen Korea or Macrogen Europe, and *spa* types were assigned using Ridom StaphType software (Ridom GmbH, Würzburg, Germany) [87]. *spa*

types were obtained from 99% of the isolates, and 364 unique *spa* types were assigned according to the Ridom StaphType software in the first visit. Six isolates were not typed due to repeated negative *spa* PCR amplification or deviating repeat length.

3.2.3 Questionnaires

Healthcare workers

Self-reported information on work in the healthcare services was obtained by the interview question 'Do you work in healthcare services (hospitals, nursing home, senior care service, general practitioner (GP)'s office, and public health centre)?' (Yes/No) (Appendices C and D).

Two self-administered questionnaires were filled out and checked for any errors at the screening center by the interviewers (Appendices A and B).

Smoking

Smoking status was determined from the question 'Do you/Did you smoke daily?' (Yes, now/Yes, previously/Never) and recoded into 'Current daily smoking' (Yes/No). If you currently smoke, how many cigarettes do you usually smoke per day? and if you currently smoke, how many years in all have you smoked daily?

Other variables

Alcohol intake was determined from the question 'How often do you usually drink alcohol?' (Never/Monthly or more infrequently/2–4 times a month/2–3 times a week/4 or more times a week) and dichotomized into two categories (< or ≥2–3 times/week). Physical activity was obtained through the following: 'Exercise and physical exertion in leisure time the last twelve months was divided in four levels' (Reading, watching TV, or other sedentary activity?/Walking, cycling, or other forms of exercise at least 4 hours a week including walking or cycling to place of work, Sunday-walking, etc.?/ Participation in recreational sports, heavy gardening etc. duration of activity at least 4 hours a week?/Participation in hard training or sports competitions, regularly several times a week?). The response categories were recoded to three levels with the two upper categories merged.

Residing with children was determined from the question 'Do you live with people younger than 18 years of age?'(Yes/No). Education level was determined from the question

'What is the highest level of education you have completed?' (Primary, Secondary/Secondary, O-level (GCSE)/A-level/college, university) recoded into two levels (< or \ge college, university). Total household income was determined from 'What is the household's total taxable income last year? Include income from work, social benefits and similar', and dichotomized into < or \ge level of the lowest income quintile (37,000 Euro/years).

Use of hormonal contraceptives was determined from the question 'Do you currently use any prescription drug that influences the menstruation? Including oral or dermal contraceptives, intra uterine device with hormones or similar' (Yes/No). Diabetes status was determined from the question 'Do you have or have you had diabetes?' (Yes/No). The information available did not permit subclassification of type 1 or type 2 diabetes mellitus (DM). Atopic eczema was determined from the question' Have you ever been diagnosed with atopic eczema by a physician?' (Yes/No). Psoriasis was determined from the question 'Have you ever been diagnosed with psoriasis by a physician?' (Yes/No). Recent hospitalization was determined from the question 'Have you during the past year been admitted to a hospital?' (Yes/No).

3.2.4 Clinical examiniation

Body height in centimetres (cm) and weight in kilograms (kg) were electronically measured to the nearest 0.1 unit wearing light clothing and no shoes (Jenix DS 102 stadiometer, Dong Sahn Jenix, Seoul, Korea). BMI was calculated as weight divided by height squared (kg/m²) [284]. WC was measured without outerwear by using a measuring tape. WC was measured at the umbilical line to the nearest cm [284]. The World Health Organization (WHO) defines $BMI \ge 30.0 \text{ kg/m}^2$ as obesity and WC values >88 cm and >102 cm in women and men, respectively, as high risk abdominal obesity [234, 243].

3.2.5 Blood samples

Non-fasting blood samples were collected from an antecubal vein and were taken throughout the survey opening hours. Serum was prepared by centrifugation after 30 minutes respite at room temperature and analysed at the Department of Medical Biochemistry at the University Hospital of North Norway.

Vitamin D

The sera were consecutively analysed for 25(OH)D by immunometry (electrochemiluminescence immunoassay), using an automated clinical chemistry analyser (Modular E170; Roche Diagnostics) [288, 289]. The total analytical coefficient of variation (CV) was 7.3%. This was in accordance with the total analytic precision reported from the producer, where the CV was ≤7.8% as judged in any of the three different concentrations (48.6–73.8–177.0 nmol/l). The cross-reactivity with 25(OH)D₂ was <10% and the detection limit was 10 nmol/l. This analysis has been approved by the Norwegian Accreditation Authority. It has recently been found that smokers have 15–20% higher serum 25(OH)D levels than non-smokers when this method was used. The same effect of smoking was not detected when measuring serum 25(OH)D with other immunological methods or the liquid chromatography mass spectrometry (LC-MS) method [290]. We presently do not have an explanation for this discrepancy. Thus, in **Paper I** non-smokers and smokers are analysed separately.

HbA1c

HbA1c was measured from EDTA-blood samples and determined by high performance liquid chromatography (HPLC) using an automated analyzer (Variant II, Bio-Rad Laboratories INC., Hercules, CA, USA) the day after blood sampling. The reference interval was 4.3–6.1% and the total analytical coefficient of variation (CV) was <3.0%. This analysis has been certified by the National Glycohemoglobin Standardization Program (NGSP) as having documented traceability to the Diabetes Control and Complication Trial (DCCT) reference method [291]. The laboratory analysis was approved by the Norwegian Accreditation Authority. HbA1c has recently been recommended by the World Health Organization (WHO) and the American Diabetes Association (ADA) as an alternative to measurements of glucose levels in diagnosing pre-diabetes and diabetes, and an international expert committee has proposed cut-off values for pre-diabetes (HbA1c 6.0–6.4%) and diabetes (HbA1c ≥6.5%) [292].

3.2.6 Statistical analysis

Logistic regression models were used to test whether metabolic and hormonal profiles and environmental factors were associated with *S. aureus* nasal colonization and carriage with estimation of Odds Ratios (ORs) and 95% confidence intervals (CIs). Multiple logistic

regression models to evaluate the joint statistical significance of several independent variables in relation to the outcome variables were used. As studies have identified higher *S. aureus* nasal carriage rates among men as compared to women [68, 154, 155, 162] and previous reports have shown that predictors of *S. aureus* nasal carriage may vary by sex [154], the regression models were stratified by sex. Tests for interaction were performed by inclusion of multiplicative terms in the models. Tests of reliability of the final analyses were done by the Hosmer-Lemeshow goodness of fit test.

In **Paper I**, logistic regression models were used to study the association between serum 25(OH)D and *S. aureus* colonization and carriage (single and double set of nasal swab cultures). The models were stratified by sex and smoking status [290]. As established thresholds for the associations between serum 25(OH)D and *S. aureus* nasal colonization and carriage are lacking, serum 25(OH)D tertiles were selected as suitable for the samples; non-smokers: <44.9 nmol/l, 44.9–58.6 nmol/l, >58.6 nmol/l; and smokers: <59.6 nmol/l, 59.6–75.3 nmol/l, >75.3 nmol/l. Also, proposed cut-points for vitamin D deficiency/insufficiency were examined (i.e. <50.0, 50.0– <75.0, ≥75.0 nmol/l) among non-smokers [208]. Selected characteristics of men and women in the different serum 25(OH)D tertiles were compared by oneway ANOVA and Kruskal-Wallis test for continuous variables and two-sided Pearson chisquared test for categorical variables. We evaluated model fit and biological plausibility of several covariates and the final multivariable models included age, BMI, DM (yes/no), and calendar month (2 months categories), and in smokers also number of cigarettes smoked per day and total years smoked [154, 161, 162, 290, 293-295].

In **Paper II**, as established thresholds for the associations between BMI, WC and *S. aureus* nasal colonization are lacking, BMI categories (<22.5, 22.5 –<25.0, 25.0 –<27.5, 27.5 –<30.0, 30.0 –<32.5, ≥ 32.5 kg/m²), WC quintiles among women (<80, 80–86, 87–92, 93–100, ≥ 101 cm) and WC quintiles among men (<91, 91–95, 96–101, 102–107, ≥ 108 cm) were defined. Selected characteristics of women and men were compared using age adjusted regression analysis with linear *Ptrend* across all BMI categories. On the basis of biological plausibility and model fit, the variables age (continuous), DM, current daily smoking, education level, and total household income were included as covariates in the multivariable model [154, 155, 161, 162, 296]. Possible interactions with age in logistic regression models stratified by age tertiles (30–43, 44–59, and 60–87 years) for both sexes and by proposed pre-/postmenopausal age ranges (<55 and ≥ 55 years) among women were explored by inclusion of multiplicative terms in the models. To control for possible confounding by pre-diabetes and undiagnosed diabetes, sensitivity analysis restricted to those with HbA1c <6.0% (N = 3,207)

was performed. To control for possible confounding by exogenous reproductive hormones, additional restriction analysis, including only non-users of hormonal contraceptives, was performed among young and premenopausal women [156].

Tests for linear trend in **Paper I** and **II** were performed by assigning consecutive integers to each serum 25(OH)D tertiles- and categories, BMI categories and WC quintiles, and testing whether the slope coefficient differed from zero by using the Wald chi-square test. Tests of statistical significance of the interaction terms were done by the likelihood ratio test comparing models with and without the multiplicative interaction terms.

In **Paper III**, selected characteristics of HCW and non-HCW were compared by two-sided Student's t-test for continuous variables and two-sided Pearson chi-squared/Fisher's exact test for categorical variables. Multivariable logistic regression models were used with adjustments of possible confounders. We tested whether residing with children modified the association between HCW status and *S. aureus nasal* carriage in stratified logistic regression analysis. Test for interaction was done by inclusion of the multiplicative terms of the two predictor variables in the model.

The analysis of *spa* types was restricted to the first visit's nasal swab culture as persistent *S. aureus* nasal carriage is not defined as concordance of *spa* types in repeated samples but as overall growth of *S. aureus* in repeated samples. The six predominant *spa* types were chosen and the logistic regression models where age- and sex adjusted [287]. In general, all tests were done two sided, and significance level was set at 0.05. Normal distribution of the continuous variables was assessed by visual inspection of histograms and where non-normality were found, ANOVA with the non-parametric Kruskal Wallis test were used for the continuous variables.

In all the three papers, subjects with missing data were excluded from the analyses.

Power calculations were performed for some of the predictors before the data collection and 4000 participants were considered to be sufficient for subgroup analysis of host-microbe relationships. Further power calculations for the same predictors were also carried out to test whether the number of participants achieved with a double set of nasal swab cultures were sufficient for subgroup analysis (See Table 1 and 2).

Table 1. Power calculations for continuous predictor variables. Tromsø Staph and Skin Study N = 2,986. Persistent *S. aureus* nasal carriers: N = 747 (25%). Nonpersistent *S. aureus* nasal carriers: N = 2,239.

Predictor variable	Mean*	SD*	Statistical Power % Difference in mean between the two carrier groups				
			2.5%	5%	10%	15%	
Serum Vit 25-OH-D3 (mmol/l)	60.9	19.2	48	96	100	100	
Serum Glucose (mmol/l)	5.2	1.2	73	99	100		
Serum HbA1c (%)	5.6	0.7	99	100			
Waist Circumference (cm)	94.9	12.2	99	100			

^{*} Total study population in the Sixth Tromsø Study. SD = standard deviation

Table 2. Power calculations for categorical predictor variables. Tromsø Staph and Skin Study N = 2,986. Persistent *S. aureus* nasal carriers: N = 747 (25%). Non-persistent *S. aureus* nasal carriers: N = 2,239.

Predictor variable	Prevalence in carriers*	Statistical Power % Absolute increase in prevalence in non-persistent carriers				
		2%	4%	6%	8%	
Ex. Current Smoking Low level of leisure physical activity	20%	22	64	93	99	

^{*} Estimate based on the total study population in the Sixth Tromsø Study.

4. SUMMARY OF MAIN RESULTS

4.1 Paper I

Staphylococcus aureus nasal carriage is associated with serum 25-hydroxyvitamin D levels, gender and smoking status. The Tromsø Staph and Skin Study.

- The prevalence of *S. aureus* nasal colonization and carriage was 37.5% (506/1,351) and 34.1% (338/992) among non-smoking men, and 24.4% (403/1,655) and 21.3% (264/1,239) among non-smoking women, respectively.
- In non-smoking men, we observed a 6.6% and 6.7% decrease in the probability of *S. aureus* colonization and carriage, respectively, for each 5 nmol/l increase in serum 25(OH)D concentration (*P*<0.001 and *P*=0.001; unadjusted).
- There was a 35% and 33% reduction in odds of colonization and carriage in upper *versus* bottom tertile of serum 25(OH)D among non-smoking men, (OR, 0.65; 95% CI, 0.49–0.87; *Ptrend* = 0.004, and OR, 0.67; 95% CI, 0.48–0.95; *Ptrend* = 0.03, respectively) and those with serum 25(OH)D concentration ≥75 nmol/l versus <50 nmol/l had almost half the odds of *S. aureus* colonization and carriage (OR, 0.54; 95% CI, 0.35–0.84; *Ptrend* = 0.004, and OR, 0.52; 95% CI, 0.31–0.90; *Ptrend* = 0.02, respectively).
- In non-smoking men aged 44–60 years, the odds ratio for *S. aureus* colonization and carriage was 0.44 and 0.51, (95% confidence interval, 0.28–0.69; *Ptrend* <0.001 and 95% CI, 0.30-0.88; *P* for trend, 0.02, respectively) in the top tertile versus the bottom tertile of serum 25(OH)D, while in younger and older adult men no association was observed (*P* for interaction = 0.10 and 0.45, respectively).
- In the smoking population, average vitamin D concentration was higher than in non-smokers; mean serum 25(OH)D concentration was 66.8 nmol/l among men and 71.3 nmol/l among women.
- The prevalence of *S. aureus* nasal colonization and carriage was 29.1% (94/323) and 24.5% (57/233) among smoking men, and 18.3% (84/460) and 15.2% (48/316) among smoking women, respectively. All the prevalence rates were significantly lower than in non-smokers (all *P*-values < 0.05).

• We did not observe any associations between serum 25(OH)D concentration and *S. aureus* nasal colonization or carriage rates either among non-smoking and smoking women or among smoking men.

4.2 Paper II

Obesity and Staphylococcus aureus nasal colonization among women and men in a general population.

- There was a positive relationship between BMI and *S. aureus* nasal colonization among women. For each 2.5 kg/m² increase in BMI a 7% increase in the odds of *S. aureus* nasal colonization was observed (multivariable model; OR 1.07, 95% CI 1.01–1.14).
- Among women, the odds of *S. aureus* nasal colonization was 67% higher in those with BMI ≥32.5 versus <22.5 kg/m² (OR, 1.67; 95% CI 1.11–2.52).
- When comparing obese and lean women aged 30–43 years, we observed that BMI ≥ 32.5 versus <22.5 kg/m² and WC ≥101 versus <80 cm were associated with a 2.60 and 2.12 times higher odds of *S. aureus* colonization, respectively (95% confidence intervals [CI] 1.35–4.98 and 95% CI 1.17–3.85).
- When comparing obese and lean women aged 30–54 years, we observed that BMI ≥ 32.5 versus <22.5 kg/m² and WC ≥101 versus <80 cm were essentially unchanged compared with women aged 30–43 years.
- When restricting the analysis to those with HbA_{1c} <6.0%, the estimated ORs for the relationships between BMI and S. *aureus* nasal colonization for women 30–43 years (P for interaction = 0.03), and 30–54 years (P for interaction = 0.67), remained essentially unchanged. Also, regarding the analysis of WC restricted to those with HbA_{1c} <6.0%, the estimated ORs among women and men remained essentially unchanged.
- Among women aged 30–43 and 30–54 years, further sensitivity analyses to non-users of hormonal contraceptives did not change the results significantly.
- BMI was not associated with *S. aureus* nasal colonization among men, but for men aged 30–43 years, being in the 5^{th} and 1^{st} WC quintiles (≥ 108 and < 91cm) were both

associated with a 1.88 times higher odds of *S. aureus* nasal colonization, compared to being in the 4th WC quintile (95% CI 1.01–3.49 and 95% CI 1.08–3.28).

4.3 Paper III

Prevalence and population structure of Staphylococcus aureus nasal carriage among healthcare workers in a general population. The Tromsø Staph and Skin Study.

- HCWs comprised 25.7% (334/1,302) women and 7.3% 71/977 men. HCWs were younger than non-HCWs among both women and men (both *P*-values < 0.05). The overall prevalence of *S. aureus* nasal carriage was 26.2 % in HCWs and 26.0 % in non-HCWs. The corresponding sex-specific rates were 22.5% and 18.4 % in women (*P* = 0.11), and 43.7% and 34.1% in men (*P* = 0.10), respectively.
- Although HCW status in the total population was not associated with *S. aureus* nasal carriage in multivariable analysis, among women, HCWs had 54% higher odds of *S. aureus* nasal carriage versus non-HCWs (OR 1.54, 95% CI 1.09–2.19). In men, no such differences were observed.
- Among women residing with children, HCWs had an 86% higher odds of *S. aureus* nasal carriage compared with non-HCWs (multivariable analysis: OR 1.86, 95% CI 1.14–3.04), whereas among women not residing with children there was no difference in odds by HCW status (*P* for interaction = 0.42), and for men, there was no pattern of effect modification by family status.
- The majority of *spa* types were observed in both HCWs and non-HCW.
- Among *S. aureus* nasal carriers, HCWs had 2.17 and 3.16 times higher odds of *spa* types t012 and t015 in the first sample, respectively, compared with non-HCWs (multivariable analysis: OR 2.17, 95% CI 1.16–4.08 and OR 3.16, 95% CI 1.13–8.87).
- For nasal carriers residing with children, HCWs had a 2.42 times higher odds of *spa* type t012 compared with non-HCWs (age- and sex-adjusted analysis: OR 2.42, 95% CI 1.03–5.70), and this association was particularly strong in male nasal carriers (age-adjusted analysis: OR 4.61, 95% CI 1.36–15.61).

5. DISCUSSION

5.1 Discussion-methodology

5.1.1 Considerations of internal validity

Epidemiological observational studies are prone to limitations regarding the validity of the findings. Internal validity refers to whether the findings are true for the population studied, while external validity refers to whether the findings also apply to populations not studied. The cross-sectional study design of this study is prone to the important limitation regarding temporality. The temporal relationship between the exposure and outcome is not clear, thus, the study design is useful for finding associations and creating hypotheses, but will not give clear answers with regard to causal relationship between exposure and outcome.

5.1.1.1 Study design

The Tromsø Staph and Skin Study is the first attempt in Norway to address the prevalence as well as the bacterial, host and environmental determinants of *S. aureus* nasal carriage in a large population-based study. Our results were in concordance with a recent study of 348 individuals who visited two shopping centers in Southern Norway during the period 2001–2005, regarding overall *S. aureus* nasal carriage rates, sex differences and predominating *S. aureus* clones [287, 297]. Furthermore, two other previous smaller Norwegian studies regarding *S. aureus* carriage in hospital settings have been performed [298, 299].

However, we aimed to describe a large unselected population of 4,026 participants of community-living adult women and men as distinct from hospital in-patients in order to better understand the interaction between microbial, host and environmental determinants for *S.aureus* carriage. The chosen population-based study design was important for the generalizability of the results in relation to the aims of our study.

5.1.1.2 Study population-Selection bias

The random population selection, high attendance rate (65.7%) and the age range 30–87 years in the sixth Tromsø Study, may reduce the risk of selection bias and increase both the internal and external validity, thus, the findings may be true for the population studied and the study

population may represent the general population of the Tromsø area [284]. However, the attendance rate varied substantially between the age groups, being 47% in the age group 30–39 years, 60% in the age group 40–49 years, 71% in the age group 50–59 years, 74% in the age group 60–79 years, and 40% in the age group 80–87 years [284]. Thus, in summary, the younger and very old subjects were underrepresented in the survey due to low attendance rates. Furthermore, the population in age group <30 years was not invited at all [284, 285].

For the study of *S. aureus* nasal colonization in the TSSS, a more evenly distributed sampling across age groups was considered to be suitable and the inclusion of 4,000 observations to be sufficient for subgroup analysis of host-microbe relationships. Of the eligible group included (N = 4,359), the attendance rate for the TSSS was 92.4% (Figure 6). The lack of participants in the age group <30 years, implies that the findings in the study are only generalizable to the age group 30–87 years, and in Paper III, in particular, the lack of participant <30 years may represent a limitation of the study as the prevalence of *S. aureus* carriage is higher in younger compared with older adults. Thus, the role of work in healthcare services for carriage among the working population below 30 years should be addressed in further studies.

Differences in motivation (little interest in health screening, which could give lower participation rates among younger adults-a trend in the Tromsø Study as well as in other population studies both in Norway and internationally) [284, 285] or other obstacles (e.g. nursing home residence, chronically ill or bedridden patients and homeless people) to participate can produce a non-response bias among certain subgroups. Thus, participants in this study may represent a healthier population compared to non-participants and thus have lower rates of *S. aureus* nasal carriage [1, 284, 285]. Also, women had an overall higher attendance rate than men (68.4 vs. 62.9%) increasing the generalizability for the female part of the population [285].

5.1.1.3 Misclassification bias

Misclassification bias results from a systematic tendency for individuals selected for inclusion in the study to be erroneously placed in different exposure/outcome categories, thus leading to misclassification [300]. This misclassification can be differential or non-differential, depending on whether the degree of misclassification of exposure is dependent on the outcome or not, respectively. Whereas non-differential misclassification tends to weaken true

associations, the direction of the bias when differential misclassification occurs is difficult to predict [300].

The outcome variable-S. aureus nasal colonization and carriage

The sampling procedure and frequency as well as the culturing methods gives possibilities for misclassification of the *S. aureus* nasal colonization and carriage state in the current study. The nasal swabs were collected with well established methods [57] by a team of technicians, mostly nurses. Before the screening, technicians were trained in the sampling techniques and the standard procedures were specified in a protocol. In spite of this, we recognized that growth of any bacteria on the control blood agar plates in some of the nasal cultures was poor or even totally lacking, possibly caused by inadequately performed sampling. Repeated quality control activities were performed including training and observation of the technicians who did the sampling, and the swab cultures with no growth of any bacteria on both the control blood agar- and the chromIdTM *S. aureus* plates were excluded from the analyses. Also, a validation study among 108 participants of the TSSS after the quality control activity, revealed that the inter-rater reliability for nasal swab cultures taken by different technicians, was excellent (simple kappa = 0.94; 95% CI = 0.87–1.00) [301].

The number of samplings required to identify persistent carriers has been debated over time, and the more samplings that are performed and the longer time interval the carrier carries the same strain, the more accurate the classification [57, 64, 302]. We used two nasal swab cultures with a time period between the first and the second swab of 0–124 days (median 28 days and in 90% of the observations the interval was \geq 12 days). According to the "culture rule" proposed by Nouwen et al [57], in the validation study among 108 participants in the TSSS, we also assessed the concordance between *S. aureus* culturing results comparing samplings a 1-week apart with a flexible interval of 2–6 weeks. The inter-method reliability or validity was excellent for nasal carriage (weighted kappa = 0.85; 95 % CI= 0.77–0.92) [301]. However, a recent work showed that the transiently colonized subjects carried the organism for a median of 14 days [64]. The large scale screening setting in the TSSS did not allow us to strictly follow the most optimal interval of 14 days or more. Consequently the inclusion of a second sample after a shorter interval than 14 days, may have increased the risk of misclassifying some of the participants (N = 104 with two positive cultures) into the persistent carrier group.

The rationale for using the result of a single nasal swab culture to assess *S. aureus* nasal colonization state as "a proxy" for *S. aureus* nasal carriage in **Paper II** was developed from the analysis of the same sub-cohort of 2,868 participants in the TSSS who had a valid nasal swab culture taken at two different time points. We observed that only 113 of the 2,868 participants (3.9%) were misclassified as colonized from the culturing results from the first nasal swab (i.e. first swab culture positive and second swab culture negative). Furthermore, when using results of both swab cultures, 2,141 of 2,868 participants were classified as non-or transiently colonized, whereas 2,028 participants were classified as non-colonized when only the results from the first sampling were used. The concordance was thus very high with a specificity of 94.7% (2,028/2,141). Moreover, among those with two positive nasal swab cultures, 92% had the same *spa* type in both samples supporting some degree of persistency. Taken together, the high concordance of the culture results in first and second samples taken with a median time interval of 28 days implies that the use of a single nasal swab culture to assess *S. aureus* nasal colonization state as "a proxy" for *S. aureus* nasal carriage in our study may be justified.

Another important issue that could have affected the data quality of the outcome variable is the laboratory method without use of enrichment broths for analysing the swab cultures. In a recent survey of about 990 high-school students (FitFutures) which is part of the Tromsø Study, we found the sensitivity of direct plating of nasal swab cultures to be 77.2% compared with using enrichment broths (unpublished data). A number of studies have estimated the prevalence of *S. aureus* nasal colonization in the general population to be approximately 20–32% [1, 59, 154, 157]. In the present study, we observed a prevalence of *S. aureus* nasal colonization (first culture positive) and carriage (both cultures positive) of 28.7% and 25.4%, respectively) which shows good concordance with previous reports. Thus, based on a cost-effectiveness analysis, we chose to omit the enrichment step. The culturing method may have led to reduced sensitivity resulting in false-negatives by misclassifying some of the study participants as non-persistent nasal carriers when their true status should have been persistent carriers. On the other hand, the chosen culture method may have increased the specificity of the *S. aureus* nasal carriage state reducing the amount of intermittent carriers and those persistent carriers with low numbers of colony forming units (CFUs).

The nares were the only body site sampled, whereas colonization may occur also in other sites such as the throat, skin, perineum and gastrointestinal tract [1, 3, 46-49]. However, as decolonization of the nose usually has a decolonizing effect on skin, the nose is assumed to be the major site of *S. aureus* colonization [42, 54, 55]. Also, nasal carriage of the microbe

has been identified as an important risk factor for the development of *S. aureus* nosocomial infections [25, 26]. When considering the main aims of this study, nasal carriage therefore seems to play a key role.

Overall, we would expect the possible misclassifications of the outcome to be independent of the exposures, i.e. non-differential, as the study design was cross-sectional implying that neither the participants nor screening personnel would know the culturing results during the data collection. The possible misclassifications would thus attenuate the associations between the predictors and outcome.

The exposure variable–Serum 25(OH)D

In a validation study, based on data and material in the Tromsø Study, it was recently observed that smokers had 15–20% higher serum 25(OH)D levels than non-smokers when using the ECLIA (Roche) method but not when using other immunological and liquid-chromatography mass spectrometry methods [290]. Also, there was a clear dose-response relationship between the serum 25(OH)D levels and the amount of smoking and the number of years of smoking as well as overall current smoking status [290]. We do not at present have any explanation for this discrepancy, but the measurement error has led to a differential misclassification of the serum 25(OH)D levels; i.e. relatively more smokers have been classified in the higher serum 25(OH)D tertiles and categories as compared to non-smokers (**Paper I**). Furthermore, as current smoking is a known determinant reducing the risk of *S. aureus* nasal colonization and carriage, these relationships may have led to an overestimation of the association between serum 25(OH)D and *S. aureus* nasal colonization and carriage if we had analysed non-smokers and smokers together. These factors were important reasons for stratifying by smoking status in **Paper I**. Also, non-respondent participants regarding smoking status were excluded from the analysis.

The exposure variable-Healthcare worker status

In **Paper III**, self-reported information on current healthcare-associated environmental exposure was obtained by interviews (Appendices C and D). There was no information about the actual healthcare professions or in which part of the healthcare services the participants' worked in. This has probably resulted in a less precise exposure variable. Previous studies may support that the risk of *S. aureus* carriage could differ depending on the type of healthcare profession and in which part of the healthcare services the study participants work

[280, 281]. Thus, more precise information about actual jobs and working tasks would have improved the validity of this exposure variable. The imperfect definition of this exposure variable may have obscured/diluted the associations between being a "high risk healthcare worker" in hospitals and the outcome variable. Taking into account the cross-sectional study design, this misclassification would be expected to be non-differential, possibly attenuating our findings.

Furthermore, changes in the data collection regarding this variable led to missing information on healthcare worker status (N = 217) until the fifth week of the survey. However, the 217 subjects with missing data on the healthcare-associated environmental exposure were unselected as invitation letters were sent randomly, avoiding selection bias during the sampling period.

The exposure variables BMI and WC

Cardiovascular diseases and their risk factors, including overweight, obesity and diabetes, are the main focus of the Tromsø Study cohort [284, 285]. This has ensured high quality of clinical and anthropometric measurements. However, high BMI may represent muscle mass [234]. Therefore, BMI as an estimate of total body fat may be biased. As shown in **Paper II**, this may partly be supported by abdominal adiposity found to be a risk factor for *S. aureus* nasal colonization among men in contrast to high BMI.

There are measurement errors regarding **WC**. Previous studies have observed that both the intra-observer and inter-observer variability for WC were higher than for BMI [303]. Nevertheless, the differences in repeated measurements of WC were relatively small and thus, the reliability of WC should be considered in clinical practice. Also, WC has been evaluated on the prediction or estimation of specific adipose tissue depot (by DXA-derived abdominal fat mass) and found satisfactorily [304]. Taken together, this supports using WC as a predictor for abdominal fat mass in **Paper II**.

5.1.1.4 Confounding and interaction

The term confounding refers to situations when a non-causal association between a given exposure and an outcome is observed as a result of a third variable associated with both the exposure and the outcome [300]. There are no formal tests for confounding but in

observational studies, multivariable- and restriction analyses or stratification are the analytic tools that are used to control for confounding effects [300].

Paper I

Serum 25(OH)D levels have a seasonal variation in the Tromsø survey population [289] and *S. aureus* carriage has also in other studies been found to vary by season [305]. Thus, adjustment for season seems to be required in the multivariable model. This could have been solved in different ways, from dichotomizing into summer and winter, to splitting the years into three, four, six parts or by each month. In **Paper I**, we chose to adjust for season by dividing into two month periods as the two consecutive months had relatively similar serum 25(OH)D levels. However, a simulated model has shown that simple adjustments for season in observational studies, might result in a bias away from nil [306]. The authors suggested stratification of serum 25(OH)D within each month, with subsequent pooling of strata. When applying this method to our data, the associations between serum 25(OH)D levels and *S. aureus* nasal colonization still reached statistical significance levels (results not shown).

Other possible confounders that were adjusted for in **Paper I** included age, diabetes and BMI as these covariates were associated with serum 25(OH)D and *S. aureus* nasal colonization and carriage or changed the effect estimates of serum 25(OH)D on the outcome by 10% or more.

Paper II

On the basis of model fit, biological plausibility, and literature review the multivariable model included age, current daily smoking, self-reported diabetes, education level and household income as risk factors for *S. aureus* nasal colonization when evaluating the associations with BMI and WC [154, 155, 161, 162, 296, 307]. Age was included as a continuous variable.

As age is inversely related to *S. aureus* nasal carriage rates [154, 155, 162], and may also modulate lean and fat body mass and reproductive hormonal profiles in women, we explored possible interactions with age in logistic regression models stratified by age tertiles (30–43, 44–59, and 60–87 years) for both sexes and by proposed pre-/postmenopausal age ranges (30–54 and 55–87 years) among women. We were able to find an interaction between young (30–43 years) and middle aged (44–59 years) women with increasing BMI and a pattern of significant interaction between young (30–43 years) and older (60–87 years) women with increasing WC. This supports possible differences between younger (30–43) and older (44–59 years and 60–87 years) women in the effects of general and abdominal obesity

on *S. aureus* nasal colonization (**Paper II**, Table 1S and 2S). The reason for the differences is not clear but may include biological and/or environmental factors, please, see 5.2 Discussion of main results.

Serum HbA1c was used in sensitivity analyses to control for possible influences of pre-diabetes and undiagnosed diabetes by restriction of the logistic regression analysis to those with HbA1c <6.0% (N=3,207). Use of hormonal contraceptives has been found to increase the risk of *S. aureus* nasal carriage among young women [156]. To eliminate possible residual confounding by use of certain types of these exogenous hormones, restriction analysis including only non-users of hormonal contraceptives was performed among young and premenopausal women. We observed that BMI and WC as significant predictors of *S. aureus* nasal colonization among younger and premenopausal women remained or became even stronger among non-users of hormonal contraceptives.

Paper III

We adjusted for age, smoking status, BMI, education level, household income and residing with children <18 years as these factors seemed to be associated with *S. aureus* carriage and HCW status or changed the effect estimate with 10% or more. It is more disputable whether it was appropriate to adjust for recreational physical activity and alcohol intake, as these two variables did not seem to be important covariates in the model. The stratification analysis of residing with children, showed that female HCWs residing with children had an even higher odds of *S. aureus* nasal carriage than female HCWs in general, and that the female HCWs not residing with children had no increased odds of *S. aureus* nasal carriage compared with female non-HCWs.

5.1.1.5 Bias in analysis

Finding associations by chance

A statistical type 1 error denotes reporting a difference which is not real. To avoid such errors, strict statistical criteria are predefined to assess when a finding should be regarded significant. We have in the papers included in this thesis, set the significance level at 0.05, which means that p<0.05 leads to rejection of the null hypothesis. According to the normal distribution, one in 20 non-significant findings will by chance turn out significant. Hence, the more statistical comparisons performed, the greater the chance will be for reporting a false significant finding.

Since multiple testing was performed in **Paper II**, one may question if chance is an alternative explanation for the presented associations. However, the primary hypothesis was tested in all the statistical models, thus reducing the risk of chance findings. Furthermore, subgroup analysis was done as a result of formal tests for interaction. Importantly, we performed sensitivity analyses to minimize the effect of confounding by diabetes and use of hormonal contraceptives, though increasing the numbers of tests, but possibly also increasing the validity of the results.

The sample size of the study

A type 2 error denotes a situation with failure to detect a difference which is real. This might happen if the number of participants included in the study is too small or the drop-out rate is higher than expected. Adequately sized study samples will prevent this type of error.

All the papers were based on data from the TSSS which included a relatively large number of participants attending the first (N = 4,026) and second (N = 2,997) visit, minimizing the risk for type 2 error. However, the relatively extensive stratifications with subgroup analyses performed in **Paper II** and also in **Paper I**, required large sample sizes. When the initial analysis showed high concordance between two repeated nasal swab cultures with a median interval of 28 days, and that about 92% had the same *spa* type in both samples [287], the decision to use the data from the first visit in **Paper I** and **II** was made. In **Paper I**, the power was further restricted due to the need of stratifying by current daily smoking [290].

In **Paper III** we used a study population based on participants from first and second visit, as the information on current healthcare-associated environmental exposure was collected during the second visit. This study population included only 71 male HCWs which in the analyses may have caused a type 2 error, as the difference in prevalence of *S. aureus* nasal carriage among male HCWs compared with non-HCWs was relatively large, 43.7% and 34.1%, respectively. Stratifying the analysis on 'residing with children' enhanced the risk of type 2 errors in the analysis among men. Thus, all together, the interpretation of the results among men has to be done with caution.

In **Paper III**, we analyzed the associations between HCW status and the distribution of different *spa* types. We used the six predominant *spa* types to minimize the risk of type 2 errors as about 65% of the *spa* types were only observed in single individuals, implying a large genetic diversity.

5.1.2 Considerations of external validity

In spite of the limitations discussed above, the overall internal validity may be satisfactory, which is a requirement of external validity. The major threat to external validity is the representativeness of the study population. The large scale study with high numbers of participants in the age range 30–87 years, random population selection and high attendance rate (65.7%) of the sixth Tromsø Study, may secure external validity. Thus, the study population represents an ethnically homogenous, healthy general population in a society with high living standards and relatively small differences in socio-economic status. On this basis our results should be held true and allow for generalization to other populations.

5.2 Discussion of main results

Epidemiological research seeks to provide a broad perceptive on causes of disease. With the many complex known and unknown mechanisms and risk factors behind the development of *S. aureus* nasal colonization and carriage, in addition to the cross-sectional study design of the current study, we are investigating associations, not causality [308]. However, there might exist causal relationships between the variables presented here as defined by the criteria for causality published by Hill [308]. Hill's criteria of causation such as strength of the association, dose-response, consistency with other studies, specificity, biological plausibility and adjustments for important confounders will be discussed where they are relevant to evaluate our findings [308].

Persistent nasal carriage of *S. aureus*, a major risk factor for invasive *S. aureus* serious infections with high mortality [25, 26], is a complex condition determined by host, bacterial and environmental factors. The relative importance of these factors is largely unknown, but it has been suggested that host factors are the most significant [68]. It is still unclear what these host-defined circumstances are. However, in a recent Danish study of 617 middle-aged and elderly twin pairs it was concluded that the host genetic contribution to nasal carriage with *S. aureus*, is at most, very limited [153]. Nevertheless, candidate gene case-control studies have observed associations with polymorphisms in genes encoding the glucocorticoid receptor [164], interleukin 4, complement factor H, C-reactive protein [309], and HLA [310], as well as polymorphisms in vitamin D responsive genes among type 1 diabetes patients but not among healthy elderly [200, 229]. None of the studies pointing to these candidate genes have

so far been confirmed in other populations. Furthermore, bacterial factors may determine which strain is carried rather than the overall carriage status [69]. As host genetic- and bacterial factors seems to be modest determinants for overall carriage, gene-environment interactions have been proposed as alternative determinants for the *S. aureus* carriage status [153]. The microbial community consisting of *S. epidemidis*, other *S. aureus* strains, *S. pneumonia* and Corynebacterium spp. may also be of relevance [133, 134, 137, 139, 145, 147] as well as repeated antibiotic exposure [68]. These factors reflect the complexity of the determinants of *S. aureus* nasal carriage

5.2.1 The host-microbe-environment interplay

Sex and S. aureus nasal colonization and carriage

We observed that S. aureus nasal colonization and carriage rates varied by the non-modifiable host attribute sex, being highest among men (Paper I). Variation in S. aureus nasal carriage rates by sex, age and hormonal contraceptive use, with lower carriage rates among women, elderly and non-users of hormonal contraceptives, is well-known [154-156, 162]. The sexdifference may suggest a biological mechanism between reproductive hormones and carriage. Differences in behavioral and environmental factors between genders may also play a role [260, 311]. It has been proposed that women are inherently better protected to infections due to estrogens, which enhance immune functions [259, 260, 312]. In general, females may generate more robust and potentially protective innate and adaptive immune responses than their male counterparts, thereby reducing females' susceptibility to infections but increasing their risk of developing autoimmune diseases as estrogens generally exert immune enhancing activities while androgens exert suppressive effects on both innate and adaptive immune responses [259, 260]. Interestingly, the expression of antimicrobial peptides (AMPs), some of which are associated with S. aureus skin infections and nasal colonization [187, 188, 313], are modified by reproductive hormones in other body sites, e.g. the genital tract [314, 315]. Furthermore, sex steroid hormones may regulate behaviours possibly resulting in differential exposure and contact with pathogens between sexes [311]. Thus, we hypothesize that the stable, low prevalence of S. aureus carriage in women may be explained by endogenous estrogens.

The interrelationships of serum 25(OH)D, obesity and S. aureus nasal colonization and carriage

Since about 20% of healthy adults are persistent nasal carriers [1], and persistent nasal carriage of *S. aureus* is a major risk factor for infection with the microbe [25, 26], prevention or elimination of the carrier state may contribute substantially in reducing the *S. aureus* disease burden. However, there is still limited evidence in relation to modifiable risk factors for the carrier state [68].

In our study, we identified various modifiable host factors as determinants for *S. aureus* nasal colonization and carriage. Low serum 25(OH)D and obesity were associated with increased odds of *S. aureus* nasal colonization and carriage among men and women, respectively (**Paper I and II**). The inverse dose-response association between serum 25(OH)D and *S. aureus* nasal colonization and carriage in adjusted models were observed among non-smoking men with no such clear association among smoking men and non-smoking and smoking women.

Various findings in the research literature render a causal relationship between serum 25(OH)D levels and *S. aureus* carriage biologically plausible. Vitamin D appears to promote both innate and adaptive immune responses [233]. 1,25(OH)₂D₃ has been found to enhance the antimicrobial peptide function against *S. aureus* in vivo [231, 316]. The promoter of the antimicrobial protein cathelicidin and human β-defensin 2 have vitamin D responsive elements (VDRs) [317], and 1,25(OH)₂D₃ can induce cathelicidin and/or β-defensin 2 in isolated keratinocytes, monocytes, neutrophils, and myeloid cells as well as in human skin biopsies [317-319]. In addition to induce AMP expression, 1,25(OH)₂D₃ has been found to induce CD14 and TLR2 in keratinocytes [231], which may result in increased ability to detect pathogens. All these studies are suggestive of biological mechanisms that may partly explain how serum concentrations of vitamin D may protect against carriage of *S. aureus*.

The observed findings in **Paper I** are partly in line with others [163]. However, the data in **Paper I**, are to our knowledge, the first to report an association between serum vitamin D levels and MSSA carriage in a general population. In a recent study including single nasal swab cultures from 14,000 children and adults across USA, an inverse- and doseresponse association between vitamin D levels and odds of MRSA but not MSSA was observed [163]. Matheson et al [163] suggested that the microbe-dependent association could be due to the increased resistance of MRSA to natural antimicrobial peptides (i.e. cathelicidin) induced by vitamin D in host defence against *S. aureus* [320]. The apparent discrepancy with

our MSSA results may be explained by several factors such as lack of more detailed subgroup analysis as well as geographical and ethnical heterogeneity which may have influenced the findings by Matheson et al [163].

Studies have also suggested that polymorphisms in the vitamin D responsive genes, might be associated with *S. aureus* carriage among type 1 diabetes patients, but not among healthy elderly [200, 229]. A recent retrospective study including 52 subjects with *Clostridium difficile* and *S. aureus* infections showed a link between low vitamin D status and adverse outcome [230]. This may support vitamin D's immune enhancing activities.

Previous studies of other infectious disease outcomes than *S. aureus* suggest that higher vitamin D status is protective against respiratory tract infections [218, 219] and that seasonal influenza may be linked to the wintertime deficiency of vitamin D [215]. Furthermore, vitamin D deficiency has been associated with increased risk of tuberculosis (TB) [221], and immunomodulatory effects of vitamin D and sunlight in TB therapy have been observed [321].

We observed that *S. aureus* nasal colonization and carriage was significantly less frequent among smokers than among non-smokers (**Paper I**). This is in line with previous reports observing current smoking as an important determinant reducing *S. aureus* nasal carriage [68, 275]. Furthermore, serum 25(OH)D levels did not vary by *S. aureus* colonization or carriage states among smokers. The reason for the lack of association among smokers is unclear. However, we hypothesize that these findings may be caused by smoking masking the inverse association between vitamin D and *S. aureus* colonization and carriage, a possible example of gene-environment interactions [153].

Although we adjusted our analysis for important risk factors for nasal colonization, we cannot exclude that residual confounding may account for the presented associations. High serum 25(OH)D concentrations may be a proxy for a healthier lifestyle in general. More outdoor physical activity may lead to lower BMI as well as increased sun exposure. Higher socioeconomic status may be linked to a healthier lifestyle and fewer hospital admissions, all factors that may be associated with higher serum 25(OH)D and reduced risk of *S. aureus* nasal carriage. There are also several other possible confounders that may have influenced the current results such as microbial interference (*S. epidemidis*, *S. pneumonia*, *Corynebacterium* spp) [133, 139, 145, 147], repeated antibiotic use [68], and host genetic factors such as polymorphisms in the VDR gene [200, 229] but information of these variables were not included in our data.

Paper II, is to our knowledge, the first report which shows that women with higher BMI and WC have increased odds of *S. aureus* nasal colonization independent of pre-diabetes and diabetes, suggesting that excess body weight may be a marker of increased susceptibility to colonization. The association seemed to be restricted to young and premenopausal women and the associations remained stable or became even stronger among non-users of hormonal contraceptives. There was no association among older and postmenopausal women. The current study indicates that a threshold effect of fat mass may be more important than a doseresponse effect on *S. aureus* nasal colonization.

Previous studies have observed that obese patients are more likely to develop community acquired pneumonia and wound infections, as well as nosocomial sepsis, bacteremia, surgical site infections, and catheter-related infections [252, 262-265]. *S. aureus* is a frequent causative agent in several of these infections.

Our results are partly in consistency with others, i.e. obesity was associated with S. aureus nasal colonization among both women and men in the US National Health and Nutrition Examination Survey (NHANES) 2000–04 [154]. The associations in NHANES were not adjusted for DM as this covariate was not significantly associated with the outcome. Furthermore, obesity has also been identified as an independent risk factor for preoperative S. aureus nasal colonization among 4,039 surgical patients when adjusting for age, sex, current smoking, and previous antimicrobial therapy [155]. As elevated serum glucose concentration has been associated with S. aureus nasal colonization and carriage [161, 162], and obesity is linked to elevated serum glucose levels [253], one may assume that altered glucose metabolism may mediate obesity-related effects on immune responses [162, 252, 254, 255]. Importantly, Paper II shows associations between BMI and WC and S. aureus nasal colonization independent of pre-diabetes or diabetes, and thus, extends previous findings. The reasons for these associations are unclear, but may include physical, biochemical, hormonal, or environmental factors. Studies in humans and animals have suggested that adiposity in itself may cause impaired immune responses through immunomodulatory effects of changes in reproductive hormones [256-259, 312] and that obesity may cause a chronic low-grade inflammation which may attenuate leptin signalling and thus, reducing the immune responses [248, 322].

The reason for the restriction of the associations to young and premenopausal women in the current study is not clear, but we hypothesize that the possible effects of adiposity increasing *S. aureus* nasal colonization in young and premenopausal women is caused by

impaired immune responses through immunomodulatory effects of changes in reproductive hormones in this group [257, 259-261] as opposed to older and postmenopausal women [256].

In the current study, we observed that the association between BMI and *S. aureus* nasal colonization was modified by sex, which is in contrast to others who have observed increased odds of colonization among both obese women and men [154]. Nevertheless, the current study also suggests a U-shaped relationship between WC and *S. aureus* nasal colonization among young men. This may reflect non-causal relationships or sex-associated differences in lean and fat body mass. A possible lack of association between obesity and *S. aureus* nasal colonization among men may be caused by the immunomodulatory effects of changes in reproductive hormones due to obesity [258]. Also, we hypothesize that obese women may be more susceptible to leptin resistance with reduced immune responses and increased *S. aureus* colonization compared to obese men, as extra fat mass is generally accumulated subcutaneously in women and as intraabdominal visceral fat in men [323]. Lower estrogen levels may also increase the risk of leptin resistance [324].

Important risk factors of nasal colonization were included as covariates in the models, but there may still be residual confounding factors, which may account for the presented associations. Residing with children may be one of these. Environmental factors such as transmission between household members and residing with children have been shown to be risk factors for colonization [69, 147]. However, adjustments for residing with children in our analysis did not change the effect of the associations. Genetic—environment interactions as determinants of obesity and risk of carriage and infection may also be considered. Altered methylations and histone modifications of gene transcripts of cells in the immune system and leptin production, adapted in response to intrinsic and environmental stimuli, may be of importance [248]. Further studies are needed addressing the effects of diet-induced obesity on epigenetic modification of cells in the immune system and leptin production.

In contrast to the positive associations between BMI, WC and *S. aureus* nasal colonization among women, low serum 25(OH)D concentrations were associated with *S. aureus* nasal colonization and carriage among non-smoking men. The reason for the variation in determinants for *S. aureus* nasal colonization and carriage by sex in our study is unclear, but previous reports have shown that predictors of *S. aureus* nasal colonization may vary by sex [154]. However, spurious associations in our study cannot be excluded. Nevertheless, we hypothesize that the low prevalence of *S. aureus* carriage in women is mainly explained by endogenous estrogens that may overwhelm the protective effect of vitamin D. The observed

sex difference is in accordance with studies of other outcomes; type 2 diabetes and insulin resistance have been associated with low vitamin D status in men only [325, 326].

Given that causality can be established, the inverse dose-response relationship between vitamin D status and S. aureus nasal colonization and carriage observed among nonsmoking men in our study may point to targets for reducing the reservoir of S. aureus in the population. Although there is no agreement on the optimal serum concentration of 25(OH)D, a recent consensus panel recommended that a serum concentration 75-100 nmol/l for different health outcomes should be targeted [208, 209]. In our study, mean serum 25(OH)D concentration was 53.3 nmol/l and 52.4 nmol/l in non-smoking men and women, respectively, and among the 3,006 non-smoking participants, only 286 (9.5%) had serum levels of \geq 75 nmol/l. The Tromsø area is situated at 69°N and the UVB radiation at this latitude is below the threshold for dermal vitamin D production five months of the year. The arctic climate with its low temperatures may limit the degree of skin exposure in the remaining months of the year. The relatively low rates of participants having vitamin D levels ≥75.0 nmol/l, underlines that vitamin D insufficiency may be of significance in the Tromsø area as well as globally. Given the high risk of S. aureus infection in combination with malnutrition in specific patient populations (i.e. surgical, dialysis, ICU, HIV) [27-29, 35, 36, 38, 327], and the fact that most of the infections are caused by the patient's nasal strain [25, 26], it would be of interest in further studies to evaluate if vitamin D repletion \geq 75 nmol/l might reduce the prevalence of S. aureus nasal carriage and thus reduce the risk of nosocomial infections. However, there have been divergent results of vitamin D supplementation to subjects for preventing different health outcomes [219, 228, 328, 329]. One of the reasons for a possible lack of effect of vitamin D supplementation might be too high baseline serum 25(OH)D concentrations [328]. Another reason may be unrecognized confounding. Large randomized control trials (RCT) are needed to determine a possible role of vitamin D supplementation and repletion in relation to S. aureus colonization, carriage and infection. Importantly, a U-shaped association between serum 25(OH)D concentration and risk of active TB was recently observed [330], indicating that vitamin D supplementation may have detrimental effects on the immune function among individuals with normal or high serum vitamin D levels. The low proportion of participants with higher serum 25(OH)D concentrations \geq 100 nmol/l (N = 33) in our study gave us less statistical power to evaluate the odds of S. aureus nasal colonization and carriage in those with high compared with low serum 25(OH)D levels.

As obesity has become endemic worldwide, even small increases in risk may have major impact on the overall *S. aureus* disease burden in a population. Thus, given that

causality can be established, the positive relationship between adiposity and *S. aureus* nasal colonization observed in the current study may points to important targets for intervention. For example, adipose young women in child bearing age undergoing caesarean delivery and men in need of surgery may be at particular risk for surgical site infections [252, 265]. However, future prospective studies of long term effects of obesity and weight change on the risk of *S. aureus* colonization and subsequent infections are needed.

Healthcare worker status and S. aureus nasal carriage and spa types

In **Paper III**, we observed that work in healthcare services was associated with a 54% increased odds of *S. aureus* nasal carriage among women. The odds were even higher among female HCWs residing with children, whereas in female HCWs not residing with children there was no difference in odds by HCW status. Working in the healthcare services was associated with increased odds of *spa* type t012 and t015. For men, work in healthcare services and residing with children was associated with increased prevalence of the common *spa* type t012.

To our knowledge, this is the first study which reports that female HCWs have higher odds of *S. aureus* nasal carriage than female non-HCWs in a general working-age population. Several studies have investigated whether working in healthcare services may be an environmental risk factor for MSSA colonization, but the results are not consistent [46, 277, 278, 280]. In general, many of the studies in healthcare settings may have been biased by a lack of information on background prevalence in the relevant general population as well as in households. However, in consistency with our findings, other studies confined to HCWs observed higher nasal carriage rates of MRSA among nurses than among other healthcare professionals, and cited increased patient contact as a cause [280, 281]. Our screening study did not include information about the HCWs' profession. However, the majority of the Norwegian healthcare workforce is nurses and auxiliary nurses, and about 90% of these are women [331, 332].

Our findings are in agreement with current literature supporting the view that contact transmission within HCWs' families may affect the burden of MRSA infections in hospitals [270, 272]. Households have been suggested to serve as a reservoir for *S. aureus* in the community [333] and children have been found to have a higher prevalence of *S. aureus* colonization than adults [154, 334, 335]. *S. aureus* nasal carriers may impose their carrier status upon other family members [69, 271, 272] and family size of more than 5 persons has

been found to be correlated with a higher risk of *S. aureus* colonization in children [147]. Importantly, residing with children *per se* was not associated with *S. aureus* nasal carriage in our study, and the number of children <18 years did not differ between HCWs and non-HCWs (results not shown). Thus, we hypothesize that the environmental pressure caused by high rates of contact transmission of *S. aureus* from both patients and children may exceed female HCWs' ability (i.e through hand hygiene, immune responses) to defend themselves against colonization.

The increased odds of *spa* type t012 and t015 among healthcare workers in our study are in agreement with the findings of others [109]. Grundmann et al observed that *spa* types t012 and t015 were among the top 5 predominant *spa* types that causes invasive infections in Europe [109]. Also, different studies have shown no distinction between colonizing and invasive isolates [82, 104, 287]. Thus, we may expect *spa* types t012 and t015 to be frequent causes of auto-infections among patients in the community and in hospitals, with an increased potential for transmission, which partly may explain our findings.

Among men, HCWs status and residing with children predicted S. aureus carriage by common spa types but not overall carriage rates. Men are generally at increased risk of S. aureus nasal carriage [154, 155, 162]. Thus, we hypothesize that the sex-associated susceptibility to S. aureus among men may overwhelm the effect of environmental risk factors (i.e. work and family status) for nasal carriage in our study, particularly considering the imperfect definition of the exposure variable "Do you work in healthcare services" that may have obscured/diluted the association between being a "high-risk" healthcare worker in hospitals and the outcome. However, the actual strain acquired may be determined by the surroundings [69]. Interestingly, among women who are generally at low risk of S. aureus nasal carriage, the opposite picture was observed; HCW status predicted nasal carriage rates overall and not by spa types. However, the uncertainty of this interpretation is considerable due to small numbers of male HCWs and lack of statistically significant interactions between the environmental risk factors (HCW status and residing with children). The external validity may therefore be limited. The female preference of spa type t012 in our study population may also possibly confound our results in women [287]. Another important limitation of this study is the lack of information of S. aureus nasal carriage and spa types among patients and household members, and the degree of direct contact between HCWs, their patients and household members. Future studies containing more detailed information on work and home environmental risk factors are therefore needed to increase our knowledge about the link

between HCWs and *S. aureus* nasal carriage, and to suggest novel targets for improved infection control strategies against exogenous MSSA and MRSA infections in patients.

Nearly 20% of nosocomial *S. aureus* infections have an exogenous origin [26], where HCWs may serve as an important vector. Furthermore, as colonized HCWs are capable of introducing *S. aureus* into their families [270-272], a possible clinical implication of our findings is the need for good adherence to infection control guidelines of HCWs at work [336]. Adherence to standard and- isolation precautions wherever needed at work as well as adequate routines for hand hygiene at home are of importance. Also, during outbreak investigation of MRSA in the healthcare setting, screening of patients, healthcare workers as well as their family members in question has to be considered to successfully eradicate MRSA and to reduce the risk of reintroducing MRSA into the healthcare services [272].

6. MAIN CONCLUSIONS

In summary, our cross-sectional study may support the view that there is a complex interplay between host-, microbial-, and environmental factors during *S. aureus* colonization and carriage.

- Our study suggests higher S. aureus nasal colonization and carriage rates among men compared with women and that predictors of colonization and carriage may vary by sex.
- The current study indicates an inverse dose-response association between serum 25(OH)D concentration and the odds of *S. aureus* nasal colonization and carriage in non-smoking men. Furthermore, we hypothesize that the relative importance of vitamin D in this context is particularly high in the male population, and that the inverse association between vitamin D and *S. aureus* colonization and carriage may be masked by smoking. Current smoking also reduced the odds of nasal colonization and carriage among both women and men.
- We observed that young and premenopausal women with higher BMI and WC have increased odds of *S. aureus* nasal colonization independent of pre-diabetes and diabetes and that the association remained significant among non-users of hormonal contraceptives. The results indicate that a threshold effect of fat mass may be more important than a dose-response effect on *S. aureus* nasal colonization. There was no association among postmenopausal women. High WC may also be a determinant for *S. aureus* nasal colonization among young men. The present findings imply support for additional biological mechanisms other than the glucose-insulin pathway.
- Work in healthcare services was associated with increased odds of *S. aureus* nasal carriage among women. Odds were even higher among women residing with children. Among men, work in healthcare services and residing with children were associated with increased odds of common *spa* types. Our study suggests that a synergism between environmental risk factors (work and household) is of importance for the overall *S. aureus* carrier state in HCWs. The *spa* types carried may be dictated by the surroundings.

7. FUTURE RESEARCH

- The cross-sectional study design of this work implies that further studies trying to establish a causal relationship between the main predictors and *S. aureus* nasal carriage are essential.
- The observed inverse dose-response association between serum 25(OH)D concentration and odds of *S. aureus* nasal colonization and carriage in non-smoking men needs further investigations. Prospective randomized controlled trials are needed to assess whether increase in circulating vitamin D concentration can effectively decrease the risk of *S. aureus* carriage and subsequent infection.
- The positive association between obesity and *S. aureus* nasal carriage among premenopausal women needs to be explored further. Thus, future prospective studies of long term effects of obesity, and weight loss on the risk of *S. aureus* colonization and subsequent infections are needed. Addressing serum levels of sex hormones as well as leptin among women and men in different age-strata as determinants of *S. aureus* nasal carriage may also be relevant.
- Previous studies on the relationships between the intestinal microbial flora and obesity have uncovered profound changes in the composition and metabolic function of the gut microbiota in obese individuals, which appear to enable an "obese microbiota" to extract more energy from the diet. There is increasing evidence that obese mice and humans exhibit a major reduction in the abundance of *Bacteroidetes* (e.g. Enterobacter spp.) and a proportional increase in Firmicutes (e.g. Lactobacilli and Staphylococcus spp) of the intestinal microbiota. Future studies addressing these questions are warranted.
- A seventh Tromsø Study is in the planning phase, where the 4,026 participants in the Tromsø 6 may be invited for a follow-up of nasal- and throat samples for culturing of *S. aureus*. This will give us the opportunity to further address the important question of persistent carriage, concordance of *spa* types between samples taken at Tromsø 6 and 7, and the relationships between different determinants and *S. aureus* carriage.
- Thus, as for the future, we will continue our search for determinants among hosts, microbes, and environmental factors that may be involved in *S. aureus* carriage of healthy individuals.

8. REFERENCES

- 1. Wertheim HF, Melles DC, Vos MC, van Leeuwen W, van Belkum A, Verbrugh HA, Nouwen JL: The role of nasal carriage in Staphylococcus aureus infections. *Lancet Infect Dis* 2005, **5**(12):751-762.
- 2. NORM/NORM-VET 2011. Usage of antimicrobial agents and occurance of antimicrobial resistance in Norway. Tromsø/Oslo. 2012. ISSN:1502-2307 (print)/1890-9965 (electronic). Available at: www.antibiotikaresistens.no. Accessed 2013 April 12.
- 3. Williams RE: **Healthy carriage of Staphylococcus aureus: its prevalence and importance**. *Bacteriol Rev* 1963, **27**:56-71.
- 4. Fluit AC: Livestock-associated Staphylococcus aureus. *Clin Microbiol Infect* 2012, **18**(8):735-744.
- 5. Tong SY, Chen LF, Fowler VG, Jr.: Colonization, pathogenicity, host susceptibility, and therapeutics for Staphylococcus aureus: what is the clinical relevance? *Semin Immunopathol* 2012, **34**(2):185-200.
- 6. Lowy FD: **Staphylococcus aureus infections**. *N Engl J Med* 1998, **339**(8):520-532.
- 7. Diekema DJ, Pfaller MA, Schmitz FJ, Smayevsky J, Bell J, Jones RN, Beach M: Survey of infections due to Staphylococcus species: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, Latin America, Europe, and the Western Pacific region for the SENTRY Antimicrobial Surveillance Program, 1997-1999. Clin Infect Dis 2001, 32 Suppl 2:S114-S132.
- 8. van Hal SJ, Jensen SO, Vaska VL, Espedido BA, Paterson DL, Gosbell IB: **Predictors of mortality in Staphylococcus aureus Bacteremia**. *Clin Microbiol Rev* 2012, **25**(2):362-386.
- 9. Fridkin SK, Hageman JC, Morrison M, Sanza LT, Como-Sabetti K, Jernigan JA, Harriman K, Harrison LH, Lynfield R, Farley MM *et al*: **Methicillin-resistant Staphylococcus aureus disease in three communities**. *N Engl J Med* 2005, **352**(14):1436-1444.
- 10. Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB: Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. Clin Infect Dis 2004, 39(3):309-317.
- 11. VandenBergh MF, Kluytmans JA, van Hout BA, Maat AP, Seerden RJ, McDonnel J, Verbrugh HA: Cost-effectiveness of perioperative mupirocin nasal ointment in cardiothoracic surgery. *Infect Control Hosp Epidemiol* 1996, **17**(12):786-792.
- 12. Pittet D, Wenzel RP: Nosocomial bloodstream infections. Secular trends in rates, mortality, and contribution to total hospital deaths. *Arch Intern Med* 1995, **155**(11):1177-1184.
- 13. Kirkland KB, Briggs JP, Trivette SL, Wilkinson WE, Sexton DJ: **The impact of surgical-site infections in the 1990s: attributable mortality, excess length of hospitalization, and extra costs**. *Infect Control Hosp Epidemiol* 1999, **20**(11):725-730.
- 14. de Kraker ME, Davey PG, Grundmann H: Mortality and hospital stay associated with resistant Staphylococcus aureus and Escherichia coli bacteremia: estimating the burden of antibiotic resistance in Europe. *PLoS Med* 2011, **8**(10):e1001104.
- 15. van Rijen MM, Bode LG, Baak DA, Kluytmans JA, Vos MC: Reduced costs for Staphylococcus aureus carriers treated prophylactically with mupirocin and chlorhexidine in cardiothoracic and orthopaedic surgery. *PLoS One* 2012, **7**(8):e43065.
- 16. Kuehnert MJ, Hill HA, Kupronis BA, Tokars JI, Solomon SL, Jernigan DB: **Methicillin-resistant-Staphylococcus aureus hospitalizations, United States**. *Emerg Infect Dis* 2005, **11**(6):868-872.
- 17. Laupland KB, Church DL, Mucenski M, Sutherland LR, Davies HD: **Population-based study of the epidemiology of and the risk factors for invasive Staphylococcus aureus infections**. *J Infect Dis* 2003, **187**(9):1452-1459.

- 18. Cosgrove SE, Sakoulas G, Perencevich EN, Schwaber MJ, Karchmer AW, Carmeli Y:

 Comparison of mortality associated with methicillin-resistant and methicillin-susceptible

 Staphylococcus aureus bacteremia: a meta-analysis. Clin Infect Dis 2003, 36(1):53-59.
- 19. Cosgrove SE, Qi Y, Kaye KS, Harbarth S, Karchmer AW, Carmeli Y: **The impact of methicillin** resistance in Staphylococcus aureus bacteremia on patient outcomes: mortality, length of stay, and hospital charges. *Infect Control Hosp Epidemiol* 2005, **26**(2):166-174.
- 20. Whitby M, McLaws ML, Berry G: **Risk of death from methicillin-resistant Staphylococcus aureus bacteraemia: a meta-analysis**. *Med J Aust* 2001, **175**(5):264-267.
- 21. Grundmann H, Aires-de-Sousa M, Boyce J, Tiemersma E: **Emergence and resurgence of meticillin-resistant Staphylococcus aureus as a public-health threat**. *Lancet* 2006, **368**(9538):874-885.
- 22. European Centre for Disease Prevention and Control (ECDC). Proportion of Methicillin-Resistant Staphylococcus aureus (MRSA) isolates in Participating Countries in 2011.
 Stockholm. 2013. Available at: http://ecdc.europa.eu/en/activities/surveillance/EARS-Net/database/Pages/maps report.aspx. Accessed 2013 Feb 12.
- 23. Gagliotti C, Balode A, Baquero F, Degener J, Grundmann H, Gur D, Jarlier V, Kahlmeter G, Monen J, Monnet DL *et al*: Escherichia coli and Staphylococcus aureus: bad news and good news from the European Antimicrobial Resistance Surveillance Network (EARS-Net, formerly EARSS), 2002 to 2009. *Euro Surveill* 2011, 16(11).
- 24. Skov R: MRSA infections increasing in the Nordic countries. *Euro Surveill* 2005, **10**(8):E050804 050802.
- von Eiff C, Becker K, Machka K, Stammer H, Peters G: **Nasal carriage as a source of Staphylococcus aureus bacteremia. Study Group**. *N Engl J Med* 2001, **344**(1):11-16.
- Wertheim HF, Vos MC, Ott A, van Belkum A, Voss A, Kluytmans JA, van Keulen PH, Vandenbroucke-Grauls CM, Meester MH, Verbrugh HA: Risk and outcome of nosocomial Staphylococcus aureus bacteraemia in nasal carriers versus non-carriers. *Lancet* 2004, 364(9435):703-705.
- 27. Bode LG, Kluytmans JA, Wertheim HF, Bogaers D, Vandenbroucke-Grauls CM, Roosendaal R, Troelstra A, Box AT, Voss A, van dT, I et al: Preventing surgical-site infections in nasal carriers of Staphylococcus aureus. N Engl J Med 2010, 362(1):9-17.
- 28. Kalmeijer MD, van Nieuwland-Bollen E, Bogaers-Hofman D, de Baere GA: Nasal carriage of Staphylococcus aureus is a major risk factor for surgical-site infections in orthopedic surgery. *Infect Control Hosp Epidemiol* 2000, **21**(5):319-323.
- 29. Kaplowitz LG, Comstock JA, Landwehr DM, Dalton HP, Mayhall CG: **Prospective study of microbial colonization of the nose and skin and infection of the vascular access site in hemodialysis patients**. *J Clin Microbiol* 1988, **26**(7):1257-1262.
- 30. Rebel MH, Van Furth R, Stevens P, Bosscher-Zonderman L, Noble WC: **The flora of renal** haemodialysis shunt sites. *J Clin Pathol* 1975, **28**(1):29-32.
- 31. Luzar MA, Coles GA, Faller B, Slingeneyer A, Dah GD, Briat C, Wone C, Knefati Y, Kessler M, Peluso F: **Staphylococcus aureus nasal carriage and infection in patients on continuous ambulatory peritoneal dialysis**. *N Engl J Med* 1990, **322**(8):505-509.
- 32. Chang FY, Singh N, Gayowski T, Drenning SD, Wagener MM, Marino IR: **Staphylococcus** aureus nasal colonization and association with infections in liver transplant recipients. *Transplantation* 1998, **65**(9):1169-1172.
- 33. Desai D, Desai N, Nightingale P, Elliott T, Neuberger J: Carriage of methicillin-resistant Staphylococcus aureus is associated with an increased risk of infection after liver transplantation. *Liver Transpl* 2003, **9**(7):754-759.
- 34. Chapoutot C, Pageaux GP, Perrigault PF, Joomaye Z, Perney P, Jean-Pierre H, Jonquet O, Blanc P, Larrey D: **Staphylococcus aureus nasal carriage in 104 cirrhotic and control patients.** A prospective study. *J Hepatol* 1999, **30**(2):249-253.

- 35. Corbella X, Dominguez MA, Pujol M, Ayats J, Sendra M, Pallares R, Ariza J, Gudiol F: Staphylococcus aureus nasal carriage as a marker for subsequent staphylococcal infections in intensive care unit patients. *Eur J Clin Microbiol Infect Dis* 1997, **16**(5):351-357.
- 36. Nguyen MH, Kauffman CA, Goodman RP, Squier C, Arbeit RD, Singh N, Wagener MM, Yu VL: Nasal carriage of and infection with Staphylococcus aureus in HIV-infected patients. *Ann Intern Med* 1999, **130**(3):221-225.
- 37. Garrouste-Orgeas M, Timsit JF, Kallel H, Ben Ali A, Dumay MF, Paoli B, Misset B, Carlet J: Colonization with methicillin-resistant Staphylococcus aureus in ICU patients: morbidity, mortality, and glycopeptide use. *Infect Control Hosp Epidemiol* 2001, **22**(11):687-692.
- 38. Melles DC, Pauw E, van den Boogaard L, Boelens HA, Peters J, Peeters JK, Witsenboer H, van Leeuwen WB, Verbrugh HA, van Belkum A *et al*: **Host-microbe interplay in persistent Staphylococcus aureus nasal carriage in HIV patients**. *Microbes Infect* 2008, **10**(2):151-158.
- 39. Kluytmans JA, Mouton JW, Ijzerman EP, Vandenbroucke-Grauls CM, Maat AW, Wagenvoort JH, Verbrugh HA: Nasal carriage of Staphylococcus aureus as a major risk factor for wound infections after cardiac surgery. *J Infect Dis* 1995, **171**(1):216-219.
- 40. Calia FM, Wolinsky E, Mortimer EA, Jr., Abrams JS, Rammelkamp CH, Jr.: **Importance of the carrier state as a source of Staphylococcus aureus in wound sepsis**. *J Hyg (Lond)* 1969, **67**(1):49-57.
- 41. Kluytmans JA, Wertheim HF: Nasal carriage of Staphylococcus aureus and prevention of nosocomial infections. *Infection* 2005, **33**(1):3-8.
- 42. Perez-Fontan M, Garcia-Falcon T, Rosales M, Rodriguez-Carmona A, Adeva M, Rodriguez-Lozano I, Moncalian J: **Treatment of Staphylococcus aureus nasal carriers in continuous ambulatory peritoneal dialysis with mupirocin: long-term results**. *Am J Kidney Dis* 1993, **22**(5):708-712.
- 43. Perl TM, Cullen JJ, Wenzel RP, Zimmerman MB, Pfaller MA, Sheppard D, Twombley J, French PP, Herwaldt LA: Intranasal mupirocin to prevent postoperative Staphylococcus aureus infections. *N Engl J Med* 2002, **346**(24):1871-1877.
- 44. Chow JW, Yu VL: Staphylococcus aureus nasal carriage in hemodialysis patients. Its role in infection and approaches to prophylaxis. *Arch Intern Med* 1989, **149**(6):1258-1262.
- 45. Boelaert JR, De Smedt RA, De Baere YA, Godard CA, Matthys EG, Schurgers ML, Daneels RF, Gordts BZ, Van Landuyt HW: **The influence of calcium mupirocin nasal ointment on the incidence of Staphylococcus aureus infections in haemodialysis patients**. *Nephrol Dial Transplant* 1989, **4**(4):278-281.
- 46. Kluytmans J, van Belkum A, Verbrugh H: **Nasal carriage of Staphylococcus aureus:** epidemiology, underlying mechanisms, and associated risks. *Clin Microbiol Rev* 1997, **10**(3):505-520.
- 47. Armstrong-Esther CA: Carriage patterns of Staphylococcus aureus in a healthy non-hospital population of adults and children. *Ann Hum Biol* 1976, **3**(3):221-227.
- 48. Ridley M: Perineal carriage of Staph. aureus. *Br Med J* 1959, **1**(5117):270-273.
- 49. Wertheim HF, Verveer J, Boelens HA, van Belkum A, Verbrugh HA, Vos MC: **Effect of** mupirocin treatment on nasal, pharyngeal, and perineal carriage of Staphylococcus aureus in healthy adults. *Antimicrob Agents Chemother* 2005, **49**(4):1465-1467.
- 50. Hamdan-Partida A, Sainz-Espunes T, Bustos-Martinez J: Characterization and persistence of Staphylococcus aureus strains isolated from the anterior nares and throats of healthy carriers in a Mexican community. *J Clin Microbiol* 2010, **48**(5):1701-1705.
- 51. Lee CJ, Sankaran S, Mukherjee DV, Apa ZL, Hafer CA, Wright L, Larson EL, Lowy FD: Staphylococcus aureus oropharyngeal carriage in a prison population. *Clin Infect Dis* 2011, **52**(6):775-778.
- 52. Marshall C, Spelman D: Re: is throat screening necessary to detect methicillin-resistant Staphylococcus aureus colonization in patients upon admission to an intensive care unit? *J Clin Microbiol* 2007, **45**(11):3855.

- 53. Nilsson P, Ripa T: **Staphylococcus aureus throat colonization is more frequent than colonization in the anterior nares**. *J Clin Microbiol* 2006, **44**(9):3334-3339.
- 54. Parras F, Guerrero MC, Bouza E, Blazquez MJ, Moreno S, Menarguez MC, Cercenado E: Comparative study of mupirocin and oral co-trimoxazole plus topical fusidic acid in eradication of nasal carriage of methicillin-resistant Staphylococcus aureus. *Antimicrob Agents Chemother* 1995, **39**(1):175-179.
- 55. Varga DT, White A: **Suppression of nasal, skin, and aerial staphylococci by nasal application of methicillin**. *J Clin Invest* 1961, **40**:2209-2214.
- 56. Eriksen NH, Espersen F, Rosdahl VT, Jensen K: **Carriage of Staphylococcus aureus among 104** healthy persons during a **19-month** period. *Epidemiol Infect* 1995, **115**(1):51-60.
- 57. Nouwen JL, Ott A, Kluytmans-Vandenbergh MF, Boelens HA, Hofman A, van Belkum A, Verbrugh HA: **Predicting the Staphylococcus aureus nasal carrier state: derivation and validation of a "culture rule"**. *Clin Infect Dis* 2004, **39**(6):806-811.
- 58. Hu L, Umeda A, Kondo S, Amako K: **Typing of Staphylococcus aureus colonising human nasal** carriers by pulsed-field gel electrophoresis. *J Med Microbiol* 1995, **42**(2):127-132.
- 59. Kuehnert MJ, Kruszon-Moran D, Hill HA, McQuillan G, McAllister SK, Fosheim G, McDougal LK, Chaitram J, Jensen B, Fridkin SK *et al*: **Prevalence of Staphylococcus aureus nasal colonization in the United States, 2001-2002**. *J Infect Dis* 2006, **193**(2):172-179.
- 60. VandenBergh MF, Yzerman EP, van Belkum A, Boelens HA, Sijmons M, Verbrugh HA: **Follow-up of Staphylococcus aureus nasal carriage after 8 years: redefining the persistent carrier state**. *J Clin Microbiol* 1999, **37**(10):3133-3140.
- 61. White A: Quantitative studies of nasal carriers of staphylococci among hospitalized patients. *J Clin Invest* 1961, **40**:23-30.
- 62. Nouwen JL, Fieren MW, Snijders S, Verbrugh HA, van Belkum A: **Persistent (not intermittent)** nasal carriage of Staphylococcus aureus is the determinant of CPD-related infections. *Kidney Int* 2005, **67**(3):1084-1092.
- 63. White A: Relation between quantitative nasal cultures and dissemination of staphylococci. *J Lab Clin Med* 1961, **58**:273-277.
- oan Belkum A, Verkaik NJ, de Vogel CP, Boelens HA, Verveer J, Nouwen JL, Verbrugh HA, Wertheim HF: **Reclassification of Staphylococcus aureus nasal carriage types**. *J Infect Dis* 2009, **199**(12):1820-1826.
- 65. Nouwen J, Boelens H, van Belkum A, Verbrugh H: **Human factor in Staphylococcus aureus nasal carriage**. *Infect Immun* 2004, **72**(11):6685-6688.
- 66. Cole AM, Tahk S, Oren A, Yoshioka D, Kim YH, Park A, Ganz T: **Determinants of Staphylococcus aureus nasal carriage**. *Clin Diagn Lab Immunol* 2001, **8**(6):1064-1069.
- 67. Quinn GA, Cole AM: Suppression of innate immunity by a nasal carriage strain of Staphylococcus aureus increases its colonization on nasal epithelium. *Immunology* 2007, **122**(1):80-89.
- 68. van Belkum A, Melles DC, Nouwen J, van Leeuwen WB, van Wamel W, Vos MC, Wertheim HF, Verbrugh HA: **Co-evolutionary aspects of human colonisation and infection by Staphylococcus aureus**. *Infect Genet Evol* 2009, **9**(1):32-47.
- 69. Peacock SJ, Justice A, Griffiths D, de Silva GD, Kantzanou MN, Crook D, Sleeman K, Day NP: **Determinants of acquisition and carriage of Staphylococcus aureus in infancy**. *J Clin Microbiol* 2003, **41**(12):5718-5725.
- 70. Weidenmaier C, Goerke C, Wolz C: **Staphylococcus aureus determinants for nasal colonization**. *Trends Microbiol* 2012, **20**(5):243-250.
- 71. Olsen K, Simonsen GS, Sundsfjord A, Haukland HH, Sollid JUE, Danielsen K, Haldorsen B, Eggen AE, Furberg A-S: **The epidemiology of** *Staphylococcus aureus* **nasal and throat carriage in a large community-based population in Northern Norway. The Tromsø Staph and Skin Study.** Abstract number 1678. 19th European Congress of Clininical Microbiology and Infectious Diseases (ECCMID). May 2009. Helsinki. Finland.
- 72. Ogston A: Micrococcus Poisoning. J Anat Physiol 1882, 17(Pt 1):24-58.

- 73. Rosenbach FJ: Mikro-organismen bei den Wund-Infections-Krankheiten des Menschen. JF Bergmann's Verlag. 1884:pp. 1-122.
- 74. Engels W, Kamps M, van Boven CP: Influence of cultivation conditions on the production of staphylocoagulase by Staphylococcus aureus 104. *J Gen Microbiol* 1978, 109(2):237-243.
- 75. Zajdel M, Wegrzynowicz Z, Jeljaszewicz J, Pulverer G: **Mechanism of action of staphylocoagulase and clumping factor**. *Contrib Microbiol Immunol* 1973, **1**:364-375.
- 76. Graham JE, Wilkinson BJ: **Staphylococcus aureus osmoregulation: roles for choline, glycine betaine, proline, and taurine**. *J Bacteriol* 1992, **174**(8):2711-2716.
- 77. Becker K, von Eiff C, Keller B, Bruck M, Etienne J, Peters G: Thermonuclease gene as a target for specific identification of Staphylococcus intermedius isolates: use of a PCR-DNA enzyme immunoassay. *Diagn Microbiol Infect Dis* 2005, **51**(4):237-244.
- 78. Morgan M: Methicillin-resistant Staphylococcus aureus and animals: zoonosis or humanosis? *J Antimicrob Chemother* 2008, **62**(6):1181-1187.
- 79. Weese JS: **Methicillin-resistant Staphylococcus aureus in animals**. *ILAR J* 2010, **51**(3):233-244.
- 80. Kuroda M, Ohta T, Uchiyama I, Baba T, Yuzawa H, Kobayashi I, Cui L, Oguchi A, Aoki K, Nagai Y et al: Whole genome sequencing of meticillin-resistant Staphylococcus aureus. *Lancet* 2001, **357**(9264):1225-1240.
- 81. Chain PS, Grafham DV, Fulton RS, Fitzgerald MG, Hostetler J, Muzny D, Ali J, Birren B, Bruce DC, Buhay C *et al*: **Genomics. Genome project standards in a new era of sequencing**. *Science* 2009, **326**(5950):236-237.
- 82. Lindsay JA, Moore CE, Day NP, Peacock SJ, Witney AA, Stabler RA, Husain SE, Butcher PD, Hinds J: Microarrays reveal that each of the ten dominant lineages of Staphylococcus aureus has a unique combination of surface-associated and regulatory genes. *J Bacteriol* 2006, **188**(2):669-676.
- 83. Herron-Olson L, Fitzgerald JR, Musser JM, Kapur V: **Molecular correlates of host specialization in Staphylococcus aureus**. *PLoS One* 2007, **2**(10):e1120.
- 84. Holden MT, Lindsay JA, Corton C, Quail MA, Cockfield JD, Pathak S, Batra R, Parkhill J, Bentley SD, Edgeworth JD: Genome sequence of a recently emerged, highly transmissible, multi-antibiotic- and antiseptic-resistant variant of methicillin-resistant Staphylococcus aureus, sequence type 239 (TW). *J Bacteriol* 2010, 192(3):888-892.
- 85. Struelens MJ: Molecular epidemiologic typing systems of bacterial pathogens: current issues and perspectives. *Mem Inst Oswaldo Cruz* 1998, **93**(5):581-585.
- 86. Koreen L, Ramaswamy SV, Graviss EA, Naidich S, Musser JM, Kreiswirth BN: spa typing method for discriminating among Staphylococcus aureus isolates: implications for use of a single marker to detect genetic micro- and macrovariation. *J Clin Microbiol* 2004, **42**(2):792-799.
- 87. Harmsen D, Claus H, Witte W, Rothganger J, Claus H, Turnwald D, Vogel U: **Typing of methicillin-resistant Staphylococcus aureus in a university hospital setting by using novel software for spa repeat determination and database management**. *J Clin Microbiol* 2003, **41**(12):5442-5448.
- 88. Mellmann A, Weniger T, Berssenbrugge C, Rothganger J, Sammeth M, Stoye J, Harmsen D: Based Upon Repeat Pattern (BURP): an algorithm to characterize the long-term evolution of Staphylococcus aureus populations based on spa polymorphisms. *BMC Microbiol* 2007, 7:98.
- 89. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, Swaminathan B: Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995, **33**(9):2233-2239.
- 90. Murchan S, Kaufmann ME, Deplano A, de Ryck R, Struelens M, Zinn CE, Fussing V, Salmenlinna S, Vuopio-Varkila J, El Solh N *et al*: Harmonization of pulsed-field gel electrophoresis protocols for epidemiological typing of strains of methicillin-resistant Staphylococcus aureus: a single approach developed by consensus in 10 European

- laboratories and its application for tracing the spread of related strains. *J Clin Microbiol* 2003, **41**(4):1574-1585.
- 91. Feil EJ, Cooper JE, Grundmann H, Robinson DA, Enright MC, Berendt T, Peacock SJ, Smith JM, Murphy M, Spratt BG *et al*: **How clonal is Staphylococcus aureus?** *J Bacteriol* 2003, **185**(11):3307-3316.
- 92. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG: Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of Staphylococcus aureus. *J Clin Microbiol* 2000, **38**(3):1008-1015.
- 93. Enright MC, Spratt BG: Multilocus sequence typing. Trends Microbiol 1999, 7(12):482-487.
- 94. Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG: **eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data**. *J Bacteriol* 2004, **186**(5):1518-1530.
- 95. Melles DC, van Leeuwen WB, Snijders SV, Horst-Kreft D, Peeters JK, Verbrugh HA, van Belkum A: Comparison of multilocus sequence typing (MLST), pulsed-field gel electrophoresis (PFGE), and amplified fragment length polymorphism (AFLP) for genetic typing of Staphylococcus aureus. *J Microbiol Methods* 2007, 69(2):371-375.
- 96. El Garch F, Hallin M, De Mendonca R, Denis O, Lefort A, Struelens MJ: **StaphVar-DNA** microarray analysis of accessory genome elements of community-acquired methicillin-resistant **Staphylococcus** aureus. *J Antimicrob Chemother* 2009, **63**(5):877-885.
- 97. Monecke S, Jatzwauk L, Weber S, Slickers P, Ehricht R: **DNA microarray-based genotyping of methicillin-resistant Staphylococcus aureus strains from Eastern Saxony**. *Clin Microbiol Infect* 2008, **14**(6):534-545.
- 98. Saunders NA, Underwood A, Kearns AM, Hallas G: A virulence-associated gene microarray: a tool for investigation of the evolution and pathogenic potential of Staphylococcus aureus. *Microbiology* 2004, **150**(Pt 11):3763-3771.
- 99. Kennedy AD, Otto M, Braughton KR, Whitney AR, Chen L, Mathema B, Mediavilla JR, Byrne KA, Parkins LD, Tenover FC *et al*: **Epidemic community-associated methicillin-resistant Staphylococcus aureus: recent clonal expansion and diversification**. *Proc Natl Acad Sci U S A* 2008, **105**(4):1327-1332.
- 100. Rajakaruna L, Hallas G, Molenaar L, Dare D, Sutton H, Encheva V, Culak R, Innes I, Ball G, Sefton AM *et al*: **High throughput identification of clinical isolates of Staphylococcus aureus using MALDI-TOF-MS of intact cells**. *Infect Genet Evol* 2009, **9**(4):507-513.
- 101. Feil EJ, Enright MC: Analyses of clonality and the evolution of bacterial pathogens. *Curr Opin Microbiol* 2004, **7**(3):308-313.
- 102. Robinson DA, Enright MC: **Multilocus sequence typing and the evolution of methicillin- resistant Staphylococcus aureus**. *Clin Microbiol Infect* 2004, **10**(2):92-97.
- 103. Enright MC, Robinson DA, Randle G, Feil EJ, Grundmann H, Spratt BG: **The evolutionary history of methicillin-resistant Staphylococcus aureus (MRSA)**. *Proc Natl Acad Sci U S A* 2002, **99**(11):7687-7692.
- 104. Melles DC, Gorkink RF, Boelens HA, Snijders SV, Peeters JK, Moorhouse MJ, van der Spek PJ, van Leeuwen WB, Simons G, Verbrugh HA *et al*: **Natural population dynamics and expansion of pathogenic clones of Staphylococcus aureus**. *J Clin Invest* 2004, **114**(12):1732-1740.
- 105. Koning S, van Belkum A, Snijders S, van Leeuwen W, Verbrugh H, Nouwen J, Op 't Veld M, van Suijlekom-Smit LW, van der Wouden JC, Verduin C: Severity of nonbullous Staphylococcus aureus impetigo in children is associated with strains harboring genetic markers for exfoliative toxin B, Panton-Valentine leukocidin, and the multidrug resistance plasmid pSK41. *J Clin Microbiol* 2003, 41(7):3017-3021.
- 106. Muto CA, Jernigan JA, Ostrowsky BE, Richet HM, Jarvis WR, Boyce JM, Farr BM: **SHEA** guideline for preventing nosocomial transmission of multidrug-resistant strains of **Staphylococcus aureus and enterococcus**. *Infect Control Hosp Epidemiol* 2003, **24**(5):362-386.

- 107. Fitzgerald JR, Sturdevant DE, Mackie SM, Gill SR, Musser JM: **Evolutionary genomics of Staphylococcus aureus: insights into the origin of methicillin-resistant strains and the toxic shock syndrome epidemic**. *Proc Natl Acad Sci U S A* 2001, **98**(15):8821-8826.
- 108. Melles DC, Tenover FC, Kuehnert MJ, Witsenboer H, Peeters JK, Verbrugh HA, van Belkum A:
 Overlapping population structures of nasal isolates of Staphylococcus aureus from healthy
 Dutch and American individuals. *J Clin Microbiol* 2008, **46**(1):235-241.
- 109. Grundmann H, Aanensen DM, van den Wijngaard CC, Spratt BG, Harmsen D, Friedrich AW: Geographic distribution of Staphylococcus aureus causing invasive infections in Europe: a molecular-epidemiological analysis. *PLoS Med* 2010, **7**(1):e1000215.
- 110. Severin JA, Lestari ES, Kuntaman K, Melles DC, Pastink M, Peeters JK, Snijders SV, Hadi U, Duerink DO, van Belkum A *et al*: **Unusually high prevalence of panton-valentine leukocidin genes among methicillin-sensitive Staphylococcus aureus strains carried in the Indonesian population**. *J Clin Microbiol* 2008, **46**(6):1989-1995.
- 111. McCarthy AJ, Lindsay JA: **Genetic variation in Staphylococcus aureus surface and immune** evasion genes is lineage associated: implications for vaccine design and host-pathogen interactions. *BMC Microbiol* 2010, **10**:173.
- 112. Weidenmaier C, Kokai-Kun JF, Kristian SA, Chanturiya T, Kalbacher H, Gross M, Nicholson G, Neumeister B, Mond JJ, Peschel A: Role of teichoic acids in Staphylococcus aureus nasal colonization, a major risk factor in nosocomial infections. *Nat Med* 2004, **10**(3):243-245.
- 113. Weidenmaier C, Kokai-Kun JF, Kulauzovic E, Kohler T, Thumm G, Stoll H, Gotz F, Peschel A: Differential roles of sortase-anchored surface proteins and wall teichoic acid in Staphylococcus aureus nasal colonization. *Int J Med Microbiol* 2008, **298**(5-6):505-513.
- 114. Speziale P, Pietrocola G, Rindi S, Provenzano M, Provenza G, Di Poto A, Visai L, Arciola CR: Structural and functional role of Staphylococcus aureus surface components recognizing adhesive matrix molecules of the host. *Future microbiology* 2009, **4**(10):1337-1352.
- 115. Burian M, Rautenberg M, Kohler T, Fritz M, Krismer B, Unger C, Hoffmann WH, Peschel A, Wolz C, Goerke C: **Temporal expression of adhesion factors and activity of global regulators during establishment of Staphylococcus aureus nasal colonization**. *J Infect Dis* 2010, **201**(9):1414-1421.
- 116. O'Brien LM, Walsh EJ, Massey RC, Peacock SJ, Foster TJ: **Staphylococcus aureus clumping** factor B (ClfB) promotes adherence to human type I cytokeratin 10: implications for nasal colonization. *Cell Microbiol* 2002, **4**(11):759-770.
- 117. Clarke SR, Andre G, Walsh EJ, Dufrene YF, Foster TJ, Foster SJ: Iron-regulated surface determinant protein A mediates adhesion of Staphylococcus aureus to human corneocyte envelope proteins. *Infect Immun* 2009, **77**(6):2408-2416.
- 118. Clarke SR, Brummell KJ, Horsburgh MJ, McDowell PW, Mohamad SA, Stapleton MR, Acevedo J, Read RC, Day NP, Peacock SJ *et al*: **Identification of in vivo-expressed antigens of Staphylococcus aureus and their use in vaccinations for protection against nasal carriage**. *J Infect Dis* 2006, **193**(8):1098-1108.
- 119. Schaffer AC, Solinga RM, Cocchiaro J, Portoles M, Kiser KB, Risley A, Randall SM, Valtulina V, Speziale P, Walsh E *et al*: Immunization with Staphylococcus aureus clumping factor B, a major determinant in nasal carriage, reduces nasal colonization in a murine model. *Infect Immun* 2006, **74**(4):2145-2153.
- 120. Burian M, Wolz C, Goerke C: **Regulatory adaptation of Staphylococcus aureus during nasal colonization of humans**. *PLoS One* 2010, **5**(4):e10040.
- 121. Corrigan RM, Miajlovic H, Foster TJ: **Surface proteins that promote adherence of Staphylococcus aureus to human desquamated nasal epithelial cells**. *BMC Microbiol* 2009, **9**:22.
- 122. Roche FM, Meehan M, Foster TJ: **The Staphylococcus aureus surface protein SasG and its homologues promote bacterial adherence to human desquamated nasal epithelial cells**. *Microbiology* 2003, **149**(Pt 10):2759-2767.

- 123. Kaliner MA: **Human nasal respiratory secretions and host defense**. *Am Rev Respir Dis* 1991, **144**(3 Pt 2):S52-56.
- 124. Cole AM, Dewan P, Ganz T: Innate antimicrobial activity of nasal secretions. *Infect Immun* 1999, **67**(7):3267-3275.
- 125. Casado B, Pannell LK, Iadarola P, Baraniuk JN: **Identification of human nasal mucous proteins using proteomics**. *Proteomics* 2005, **5**(11):2949-2959.
- 126. Bera A, Biswas R, Herbert S, Kulauzovic E, Weidenmaier C, Peschel A, Gotz F: **Influence of wall teichoic acid on lysozyme resistance in Staphylococcus aureus**. *J Bacteriol* 2007, **189**(1):280-283.
- 127. Moks T, Abrahmsen L, Nilsson B, Hellman U, Sjoquist J, Uhlen M: **Staphylococcal protein A consists of five IgG-binding domains**. *Eur J Biochem* 1986, **156**(3):637-643.
- 128. Rooijakkers SH, Milder FJ, Bardoel BW, Ruyken M, van Strijp JA, Gros P: **Staphylococcal complement inhibitor: structure and active sites**. *J Immunol* 2007, **179**(5):2989-2998.
- van Wamel WJ, Rooijakkers SH, Ruyken M, van Kessel KP, van Strijp JA: **The innate immune** modulators staphylococcal complement inhibitor and chemotaxis inhibitory protein of Staphylococcus aureus are located on beta-hemolysin-converting bacteriophages. *J Bacteriol* 2006, **188**(4):1310-1315.
- 130. Jin T, Bokarewa M, Foster T, Mitchell J, Higgins J, Tarkowski A: **Staphylococcus aureus resists** human defensins by production of staphylokinase, a novel bacterial evasion mechanism. *J Immunol* 2004, **172**(2):1169-1176.
- 131. Rooijakkers SH, van Wamel WJ, Ruyken M, van Kessel KP, van Strijp JA: **Anti-opsonic properties of staphylokinase**. *Microbes Infect* 2005, **7**(3):476-484.
- 132. Margolis E, Yates A, Levin BR: The ecology of nasal colonization of Streptococcus pneumoniae, Haemophilus influenzae and Staphylococcus aureus: the role of competition and interactions with host's immune response. *BMC Microbiol* 2010, **10**:59.
- 133. Frank DN, Feazel LM, Bessesen MT, Price CS, Janoff EN, Pace NR: **The human nasal microbiota and Staphylococcus aureus carriage**. *PLoS One* 2010, **5**(5):e10598.
- 134. Shinefield HR, Wilsey JD, Ribble JC, Boris M, Eichenwald HF, Dittmar CI: Interactions of staphylococcal colonization. Influence of normal nasal flora and antimicrobials on inoculated Staphylococcus aureus strain 502A. Am J Dis Child 1966, 111(1):11-21.
- Aly R, Maibach HI, Shinefield HR, Mandel A, Strauss WG: **Bacterial interference among strains of Staphylococcus aureus in man**. *J Infect Dis* 1974, **129**(6):720-724.
- 136. Noble WC, Williams RE, Jevons MP, Shooter RA: **Some Aspects of Nasal Carriage of Staphylococci**. *J Clin Pathol* 1964, **17**:79-83.
- 137. Dall'Antonia M, Coen PG, Wilks M, Whiley A, Millar M: Competition between methicillinsensitive and -resistant Staphylococcus aureus in the anterior nares. *J Hosp Infect* 2005, **61**(1):62-67.
- 138. Lina G, Boutite F, Tristan A, Bes M, Etienne J, Vandenesch F: **Bacterial competition for** human nasal cavity colonization: role of Staphylococcal agr alleles. *Appl Environ Microbiol* 2003, **69**(1):18-23.
- 139. Uehara Y, Nakama H, Agematsu K, Uchida M, Kawakami Y, Abdul Fattah AS, Maruchi N: Bacterial interference among nasal inhabitants: eradication of Staphylococcus aureus from nasal cavities by artificial implantation of Corynebacterium sp. *J Hosp Infect* 2000, 44(2):127-133.
- 140. Cogen AL, Yamasaki K, Muto J, Sanchez KM, Crotty Alexander L, Tanios J, Lai Y, Kim JE, Nizet V, Gallo RL: **Staphylococcus epidermidis antimicrobial delta-toxin (phenol-soluble modulingamma) cooperates with host antimicrobial peptides to kill group A Streptococcus**. *PLoS One* 2010, **5**(1):e8557.
- 141. Cogen AL, Yamasaki K, Sanchez KM, Dorschner RA, Lai Y, MacLeod DT, Torpey JW, Otto M, Nizet V, Kim JE *et al*: **Selective antimicrobial action is provided by phenol-soluble modulins derived from Staphylococcus epidermidis, a normal resident of the skin**. *J Invest Dermatol* 2010, **130**(1):192-200.

- 142. Otto M: Staphylococcus aureus and Staphylococcus epidermidis peptide pheromones produced by the accessory gene regulator agr system. *Peptides* 2001, **22**(10):1603-1608.
- 143. Otto M, Echner H, Voelter W, Gotz F: **Pheromone cross-inhibition between Staphylococcus** aureus and **Staphylococcus epidermidis**. *Infect Immun* 2001, **69**(3):1957-1960.
- 144. Lai Y, Cogen AL, Radek KA, Park HJ, Macleod DT, Leichtle A, Ryan AF, Di Nardo A, Gallo RL:

 Activation of TLR2 by a small molecule produced by Staphylococcus epidermidis increases
 antimicrobial defense against bacterial skin infections. *J Invest Dermatol* 2010, **130**(9):2211-2221
- 145. Iwase T, Uehara Y, Shinji H, Tajima A, Seo H, Takada K, Agata T, Mizunoe Y: **Staphylococcus epidermidis Esp inhibits Staphylococcus aureus biofilm formation and nasal colonization**. *Nature* 2010, **465**(7296):346-349.
- 146. Krismer B, Peschel A: **Does Staphylococcus aureus nasal colonization involve biofilm formation?** *Future microbiology* 2011, **6**(5):489-493.
- 147. Bogaert D, van Belkum A, Sluijter M, Luijendijk A, de Groot R, Rumke HC, Verbrugh HA, Hermans PW: Colonisation by Streptococcus pneumoniae and Staphylococcus aureus in healthy children. *Lancet* 2004, **363**(9424):1871-1872.
- 148. Lee GM, Huang SS, Rifas-Shiman SL, Hinrichsen VL, Pelton SI, Kleinman K, Hanage WP, Lipsitch M, McAdam AJ, Finkelstein JA: **Epidemiology and risk factors for Staphylococcus aureus colonization in children in the post-PCV7 era**. *BMC Infect Dis* 2009, **9**:110.
- 149. Regev-Yochay G, Trzcinski K, Thompson CM, Malley R, Lipsitch M: Interference between Streptococcus pneumoniae and Staphylococcus aureus: In vitro hydrogen peroxidemediated killing by Streptococcus pneumoniae. *J Bacteriol* 2006, **188**(13):4996-5001.
- 150. Margolis E: Hydrogen peroxide-mediated interference competition by Streptococcus pneumoniae has no significant effect on Staphylococcus aureus nasal colonization of neonatal rats. *J Bacteriol* 2009, **191**(2):571-575.
- 151. Camarinha-Silva A, Wos-Oxley ML, Jauregui R, Becker K, Pieper DH: Validating T-RFLP as a sensitive and high-throughput approach to assess bacterial diversity patterns in human anterior nares. FEMS microbiology ecology 2012, 79(1):98-108.
- 152. Frisoni E, Sakwinska O, Moreillon M: **Genetic diversity of Staphylococcus aureus in asymptomatic carriers, and nasal metagenome analysis in carriers and non-carriers.**Abstract number 10-42. 15th International Symposium on Staphylococci and Staphylococcal Infections (15th ISSSI 2012). Available at: http://isssi2012.univ-lyon1.fr. Aug. 2012. Lyon. France.
- 153. Andersen PS, Pedersen JK, Fode P, Skov RL, Fowler VG, Jr., Stegger M, Christensen K: Influence of host genetics and environment on nasal carriage of staphylococcus aureus in danish middle-aged and elderly twins. *J Infect Dis* 2012, **206**(8):1178-1184.
- 154. Gorwitz RJ, Kruszon-Moran D, McAllister SK, McQuillan G, McDougal LK, Fosheim GE, Jensen BJ, Killgore G, Tenover FC, Kuehnert MJ: **Changes in the prevalence of nasal colonization**with **Staphylococcus aureus in the United States**, **2001-2004**. *J Infect Dis* 2008, **197**(9):1226-1234.
- 155. Herwaldt LA, Cullen JJ, French P, Hu J, Pfaller MA, Wenzel RP, Perl TM: **Preoperative risk** factors for nasal carriage of Staphylococcus aureus. *Infect Control Hosp Epidemiol* 2004, **25**(6):481-484.
- Tanger P, Nurjadi D, Gaile M, Gabrysch S, Kremsner PG: **Hormonal contraceptive use and persistent Staphylococcus aureus nasal carriage**. *Clin Infect Dis* 2012.
- 157. Noble WC, Valkenburg HA, Wolters CH: **Carriage of Staphylococcus aureus in random samples of a normal population**. *J Hyg (Lond)* 1967, **65**(4):567-573.
- 158. Williams JV, Vowels BR, Honig PJ, Leyden JJ: **S. aureus isolation from the lesions, the hands,** and the anterior nares of patients with atopic dermatitis. *Pediatr Dermatol* 1998, **15**(3):194-198.

- 159. Hoeger PH, Lenz W, Boutonnier A, Fournier JM: **Staphylococcal skin colonization in children** with atopic dermatitis: prevalence, persistence, and transmission of toxigenic and nontoxigenic strains. *J Infect Dis* 1992, **165**(6):1064-1068.
- 160. Yu VL, Goetz A, Wagener M, Smith PB, Rihs JD, Hanchett J, Zuravleff JJ: **Staphylococcus** aureus nasal carriage and infection in patients on hemodialysis. **Efficacy of antibiotic** prophylaxis. *N Engl J Med* 1986, **315**(2):91-96.
- 161. Lipsky BA, Pecoraro RE, Chen MS, Koepsell TD: **Factors affecting staphylococcal colonization among NIDDM outpatients**. *Diabetes Care* 1987, **10**(4):483-486.
- 162. Nouwen J, Ott A, Boelens H, al. e: **Smoking pattern and fasting glucose levels determine Staphylococcus aureus nasal carriage.** PhD thesis. Erasmus University Medical Centre,
 Rotterdam, The Netherlands; 2004: 17-36.
- 163. Matheson EM, Mainous AG, 3rd, Hueston WJ, Diaz VA, Everett CJ: **Vitamin D and methicillin- resistant Staphylococcus aureus nasal carriage**. *Scand J Infect Dis* 2010, **42**(6-7):455-460.
- 164. van den Akker EL, Nouwen JL, Melles DC, van Rossum EF, Koper JW, Uitterlinden AG, Hofman A, Verbrugh HA, Pols HA, Lamberts SW *et al*: **Staphylococcus aureus nasal carriage is associated with glucocorticoid receptor gene polymorphisms**. *J Infect Dis* 2006, **194**(6):814-818.
- 165. Manenschijn L, Jetten AM, van Wamel WJ, Tavakol M, Koper JW, van den Akker EL, van Belkum A, van Rossum EF: Long-term cortisol levels are not associated with nasal carriage of Staphylococcus aureus. Eur J Clin Microbiol Infect Dis 2012, 31(1):97-100.
- 166. Cole AM, Dewan P, Ganz T: Innate antimicrobial activity of nasal secretions. *Infect Immun* 1999, **67**(7):3267-3275.
- Pynnonen M, Stephenson RE, Schwartz K, Hernandez M, Boles BR: **Hemoglobin promotes Staphylococcus aureus nasal colonization**. *PLoS pathogens* 2011, **7**(7):e1002104.
- 168. Schauber J, Gallo RL: **Antimicrobial peptides and the skin immune defense system**. *J Allergy Clin Immunol* 2009, **124**(3 Suppl 2):R13-18.
- 169. Proksch E, Brandner JM, Jensen JM: **The skin: an indispensable barrier**. *Exp Dermatol* 2008, **17**(12):1063-1072.
- 170. Arikawa J, Ishibashi M, Kawashima M, Takagi Y, Ichikawa Y, Imokawa G: **Decreased levels of sphingosine**, a natural antimicrobial agent, may be associated with vulnerability of the stratum corneum from patients with atopic dermatitis to colonization by Staphylococcus aureus. *J Invest Dermatol* 2002, **119**(2):433-439.
- 171. Takigawa H, Nakagawa H, Kuzukawa M, Mori H, Imokawa G: **Deficient production of hexadecenoic acid in the skin is associated in part with the vulnerability of atopic dermatitis patients to colonization by Staphylococcus aureus**. *Dermatology* 2005, **211**(3):240-248.
- 172. Nestle FO, Di Meglio P, Qin JZ, Nickoloff BJ: **Skin immune sentinels in health and disease**. *Nat Rev Immunol* 2009, **9**(10):679-691.
- 173. Takeuchi O, Hoshino K, Akira S: **Cutting edge: TLR2-deficient and MyD88-deficient mice are highly susceptible to Staphylococcus aureus infection**. *J Immunol* 2000, **165**(10):5392-5396.
- 174. Fournier B, Philpott DJ: **Recognition of Staphylococcus aureus by the innate immune system**. *Clin Microbiol Rev* 2005, **18**(3):521-540.
- 175. Craven RR, Gao X, Allen IC, Gris D, Bubeck Wardenburg J, McElvania-Tekippe E, Ting JP, Duncan JA: **Staphylococcus aureus alpha-hemolysin activates the NLRP3-inflammasome in human and mouse monocytic cells**. *PLoS One* 2009, **4**(10):e7446.
- 176. Nilsen NJ, Deininger S, Nonstad U, Skjeldal F, Husebye H, Rodionov D, von Aulock S, Hartung T, Lien E, Bakke O *et al*: **Cellular trafficking of lipoteichoic acid and Toll-like receptor 2 in relation to signaling: role of CD14 and CD36**. *J Leukoc Biol* 2008, **84**(1):280-291.
- 177. Akira S, Takeda K: **Toll-like receptor signalling**. *Nat Rev Immunol* 2004, **4**(7):499-511.
- 178. Krishna S, Miller LS: **Host-pathogen interactions between the skin and Staphylococcus aureus**. *Curr Opin Microbiol* 2012, **15**(1):28-35.

- 179. Braff MH, Zaiou M, Fierer J, Nizet V, Gallo RL: **Keratinocyte production of cathelicidin provides direct activity against bacterial skin pathogens**. *Infect Immun* 2005, **73**(10):6771-6781.
- 180. Murakami M, Ohtake T, Dorschner RA, Schittek B, Garbe C, Gallo RL: **Cathelicidin antimicrobial peptide expression in sweat, an innate defense system for the skin**. *J Invest Dermatol* 2002, **119**(5):1090-1095.
- 181. Schauber J, Gallo RL: **Expanding the roles of antimicrobial peptides in skin: alarming and arming keratinocytes**. *J Invest Dermatol* 2007, **127**(3):510-512.
- 182. Selsted ME, Ouellette AJ: **Mammalian defensins in the antimicrobial immune response**. *Nat Immunol* 2005, **6**(6):551-557.
- 183. Zaiou M, Gallo RL: Cathelicidins, essential gene-encoded mammalian antibiotics. *J Mol Med* (*Berl*) 2002, **80**(9):549-561.
- 184. Midorikawa K, Ouhara K, Komatsuzawa H, Kawai T, Yamada S, Fujiwara T, Yamazaki K, Sayama K, Taubman MA, Kurihara H *et al*: **Staphylococcus aureus susceptibility to innate antimicrobial peptides, beta-defensins and CAP18, expressed by human keratinocytes**. *Infect Immun* 2003, **71**(7):3730-3739.
- 185. Chen X, Niyonsaba F, Ushio H, Okuda D, Nagaoka I, Ikeda S, Okumura K, Ogawa H: Synergistic effect of antibacterial agents human beta-defensins, cathelicidin LL-37 and lysozyme against Staphylococcus aureus and Escherichia coli. *J Dermatol Sci* 2005, **40**(2):123-132.
- 186. Kisich KO, Howell MD, Boguniewicz M, Heizer HR, Watson NU, Leung DY: **The constitutive** capacity of human keratinocytes to kill Staphylococcus aureus is dependent on betadefensin **3**. *J Invest Dermatol* 2007, **127**(10):2368-2380.
- 187. Zanger P, Holzer J, Schleucher R, Scherbaum H, Schittek B, Gabrysch S: **Severity of Staphylococcus aureus infection of the skin is associated with inducibility of human beta-defensin 3 but not human beta-defensin 2**. *Infect Immun* 2010, **78**(7):3112-3117.
- 188. Zanger P, Nurjadi D, Vath B, Kremsner PG: **Persistent nasal carriage of Staphylococcus** aureus is associated with deficient induction of human beta-defensin 3 after sterile wounding of healthy skin in vivo. *Infect Immun* 2011, **79**(7):2658-2662.
- 189. Murakami M, Lopez-Garcia B, Braff M, Dorschner RA, Gallo RL: **Postsecretory processing generates multiple cathelicidins for enhanced topical antimicrobial defense**. *J Immunol* 2004, **172**(5):3070-3077.
- 190. Dusso AS, Brown AJ, Slatopolsky E: **Vitamin D**. *Am J Physiol Renal Physiol* 2005, **289**(1):F8-28.
- 191. Holick MF: **Vitamin D deficiency**. *N Engl J Med* 2007, **357**(3):266-281.
- 192. Holick MF: The cutaneous photosynthesis of previtamin D3: a unique photoendocrine system. *J Invest Dermatol* 1981, **77**(1):51-58.
- 193. Mithal A, Wahl DA, Bonjour JP, Burckhardt P, Dawson-Hughes B, Eisman JA, El-Hajj Fuleihan G, Josse RG, Lips P, Morales-Torres J: **Global vitamin D status and determinants of hypovitaminosis D**. *Osteoporos Int* 2009, **20**(11):1807-1820.
- 194. Norman AW: Sunlight, season, skin pigmentation, vitamin D, and 25-hydroxyvitamin D: integral components of the vitamin D endocrine system. *Am J Clin Nutr* 1998, **67**(6):1108-1110.
- 195. Norman AW: From vitamin D to hormone D: fundamentals of the vitamin D endocrine system essential for good health. *Am J Clin Nutr* 2008, **88**(2):491S-499S.
- 196. Maalouf NM: **The noncalciotropic actions of vitamin D: recent clinical developments**. *Curr Opin Nephrol Hypertens* 2008, **17**(4):408-415.
- 197. Kostner K, Denzer N, Muller CS, Klein R, Tilgen W, Reichrath J: **The relevance of vitamin D** receptor (VDR) gene polymorphisms for cancer: a review of the literature. *Anticancer Res* 2009, **29**(9):3511-3536.
- 198. Reis AF, Hauache OM, Velho G: Vitamin D endocrine system and the genetic susceptibility to diabetes, obesity and vascular disease. A review of evidence. *Diabetes Metab* 2005, **31**(4 Pt 1):318-325.

- 199. Valdivielso JM, Fernandez E: **Vitamin D receptor polymorphisms and diseases**. *Clin Chim Acta* 2006, **371**(1-2):1-12.
- 200. Panierakis C, Goulielmos G, Mamoulakis D, Maraki S, Papavasiliou E, Galanakis E: Staphylococcus aureus nasal carriage might be associated with vitamin D receptor polymorphisms in type 1 diabetes. *Int J Infect Dis* 2009, **13**(6):e437-e443.
- 201. Wang TJ, Zhang F, Richards JB, Kestenbaum B, van Meurs JB, Berry D, Kiel DP, Streeten EA, Ohlsson C, Koller DL *et al*: **Common genetic determinants of vitamin D insufficiency: a genome-wide association study**. *Lancet* 2010, **376**(9736):180-188.
- 202. Hollis BW: Measuring 25-hydroxyvitamin D in a clinical environment: challenges and needs. *Am J Clin Nutr* 2008, **88**(2):507S-510S.
- 203. Hollis BW: Assessment of vitamin D status and definition of a normal circulating range of **25-hydroxyvitamin** D. *Curr Opin Endocrinol Diabetes Obes* 2008, **15**(6):489-494.
- 204. Carter GD: **25-Hydroxyvitamin D assays: the quest for accuracy**. *Clin Chem* 2009, **55**(7):1300-1302.
- 205. Hunter D, De Lange M, Snieder H, MacGregor AJ, Swaminathan R, Thakker RV, Spector TD: Genetic contribution to bone metabolism, calcium excretion, and vitamin D and parathyroid hormone regulation. *J Bone Miner Res* 2001, **16**(2):371-378.
- 206. Lips P: Vitamin D deficiency and secondary hyperparathyroidism in the elderly: consequences for bone loss and fractures and therapeutic implications. *Endocr Rev* 2001, 22(4):477-501.
- 207. Wicherts IS, van Schoor NM, Boeke AJ, Visser M, Deeg DJ, Smit J, Knol DL, Lips P: **Vitamin D** status predicts physical performance and its decline in older persons. *J Clin Endocrinol Metab* 2007, **92**(6):2058-2065.
- 208. Bischoff-Ferrari HA, Giovannucci E, Willett WC, Dietrich T, Dawson-Hughes B: **Estimation of optimal serum concentrations of 25-hydroxyvitamin D for multiple health outcomes**. *Am J Clin Nutr* 2006, **84**(1):18-28.
- 209. Dawson-Hughes B, Mithal A, Bonjour JP, Boonen S, Burckhardt P, Fuleihan GE, Josse RG, Lips P, Morales-Torres J, Yoshimura N: **IOF position statement: vitamin D recommendations for older adults**. *Osteoporos Int* 2010, **21**(7):1151-1154.
- 210. Melamed ML, Michos ED, Post W, Astor B: **25-hydroxyvitamin D levels and the risk of mortality in the general population**. *Arch Intern Med* 2008, **168**(15):1629-1637.
- 211. Hollis BW, Wagner CL, Drezner MK, Binkley NC: Circulating vitamin D3 and 25-hydroxyvitamin D in humans: An important tool to define adequate nutritional vitamin D status. *J Steroid Biochem Mol Biol* 2007, **103**(3-5):631-634.
- 212. Luxwolda MF, Kuipers RS, Kema IP, Janneke Dijck-Brouwer DA, Muskiet FA: **Traditionally living populations in East Africa have a mean serum 25-hydroxyvitamin D concentration of 115 nmol/l**. *Br J Nutr* 2012, **108**(9):1557-1561.
- 213. Jones G: Pharmacokinetics of vitamin D toxicity. Am J Clin Nutr 2008, 88(2):582S-586S.
- 214. Becker W, Lyhne N, Pedersen A, Aro A, Fogelholm M, Thorsdottir P, Alexander J, Anderssen S, Meltzer H, Pedersen J: **Nordic Nutrition Recommendations. Integrating nutrition and physical activity.** *Scand J Nutr* 2004; 48:178-87.
- 215. Cannell JJ, Vieth R, Umhau JC, Holick MF, Grant WB, Madronich S, Garland CF, Giovannucci E: **Epidemic influenza and vitamin D**. *Epidemiol Infect* 2006, **134**(6):1129-1140.
- 216. Grant WB: Variations in vitamin D production could possibly explain the seasonality of childhood respiratory infections in Hawaii. *Pediatr Infect Dis J* 2008, **27**(9):853.
- 217. Sabetta JR, DePetrillo P, Cipriani RJ, Smardin J, Burns LA, Landry ML: **Serum 25-hydroxyvitamin d and the incidence of acute viral respiratory tract infections in healthy adults**. *PLoS One* 2010, **5**(6):e11088.
- 218. Ginde AA, Mansbach JM, Camargo CA, Jr.: **Association between serum 25-hydroxyvitamin D** level and upper respiratory tract infection in the Third National Health and Nutrition **Examination Survey**. *Arch Intern Med* 2009, **169**(4):384-390.

- 219. Laaksi I, Ruohola JP, Tuohimaa P, Auvinen A, Haataja R, Pihlajamaki H, Ylikomi T: **An** association of serum vitamin D concentrations < **40** nmol/L with acute respiratory tract infection in young Finnish men. *Am J Clin Nutr* 2007, **86**(3):714-717.
- 220. Gibney KB, MacGregor L, Leder K, Torresi J, Marshall C, Ebeling PR, Biggs BA: Vitamin D deficiency is associated with tuberculosis and latent tuberculosis infection in immigrants from sub-Saharan Africa. Clin Infect Dis 2008, 46(3):443-446.
- 221. Nnoaham KE, Clarke A: Low serum vitamin D levels and tuberculosis: a systematic review and meta-analysis. *Int J Epidemiol* 2008, **37**(1):113-119.
- Talat N, Perry S, Parsonnet J, Dawood G, Hussain R: **Vitamin d deficiency and tuberculosis progression**. *Emerg Infect Dis* 2010, **16**(5):853-855.
- 223. Williams B, Williams AJ, Anderson ST: **Vitamin D deficiency and insufficiency in children with tuberculosis**. *Pediatr Infect Dis J* 2008, **27**(10):941-942.
- 224. Sita-Lumsden A, Lapthorn G, Swaminathan R, Milburn HJ: **Reactivation of tuberculosis and vitamin D deficiency: the contribution of diet and exposure to sunlight**. *Thorax* 2007, **62**(11):1003-1007.
- 225. Feindt E, Stroder J: [Studies on the antimicrobial effect of vitamin D (author's transl)]. *Klin Wochenschr* 1977, **55**(10):507-508.
- Dowell SF, Whitney CG, Wright C, Rose CE, Jr., Schuchat A: **Seasonal patterns of invasive pneumococcal disease**. *Emerg Infect Dis* 2003, **9**(5):573-579.
- 227. White AN, Ng V, Spain CV, Johnson CC, Kinlin LM, Fisman DN: Let the sun shine in: effects of ultraviolet radiation on invasive pneumococcal disease risk in Philadelphia, Pennsylvania. *BMC Infect Dis* 2009, **9**:196.
- 228. Manaseki-Holland S, Qader G, Isaq Masher M, Bruce J, Zulf Mughal M, Chandramohan D, Walraven G: Effects of vitamin D supplementation to children diagnosed with pneumonia in Kabul: a randomised controlled trial. *Trop Med Int Health* 2010, **15**(10):1148-1155.
- 229. Claassen M, Nouwen J, Fang Y, Ott A, Verbrugh H, Hofman A, van Belkum A, Uitterlinden A: Staphylococcus aureus nasal carriage is not associated with known polymorphism in the Vitamin D receptor gene. FEMS Immunol Med Microbiol 2005, 43(2):173-176.
- 230. Youssef D, Bailey B, El Abbassi A, Copeland R, Adebonojo L, Manning T, Peiris AN: **Healthcare** costs of Staphylococcus aureus and Clostridium difficile infections in veterans: role of vitamin D deficiency. *Epidemiol Infect* 2010, **138**(9):1322-1327.
- 231. Schauber J, Dorschner RA, Coda AB, Buchau AS, Liu PT, Kiken D, Helfrich YR, Kang S, Elalieh HZ, Steinmeyer A *et al*: Injury enhances TLR2 function and antimicrobial peptide expression through a vitamin D-dependent mechanism. *J Clin Invest* 2007, **117**(3):803-811.
- 232. Bikle DD: What is new in vitamin D: 2006-2007. Curr Opin Rheumatol 2007, 19(4):383-388.
- 233. Dombrowski Y, Peric M, Koglin S, Ruzicka T, Schauber J: **Control of cutaneous antimicrobial peptides by vitamin D3**. *Archives of dermatological research* 2010, **302**(6):401-408.
- 234. **Global Database on Body Mass Index-World Health Organization.** Geneva. Available: http://apps.who.int/bmi/index.jsp?introPage=intro 1.html. Accessed 2013 April 12.
- 235. Hotamisligil GS: Inflammation and metabolic disorders. Nature 2006, 444(7121):860-867.
- 236. Semenkovich CF: **Insulin resistance and atherosclerosis**. *J Clin Invest* 2006, **116**(7):1813-1822.
- 237. Joseph J, Svartberg J, Njolstad I, Schirmer H: **Change in cardiovascular risk factors in relation to diabetes status: the Tromso Study**. *European journal of preventive cardiology* 2012, **19**(3):551-557.
- 238. Fantuzzi G: Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* 2005, **115**(5):911-919; quiz 920.
- 239. Siri W: **Body composition from fluid spaces and density: Analysis of methods**. In: *Techniques for measuring body composition*. edn. Edited by Physics DLoBaM. University of California Berkeley; 1959: 223-244.

- 240. Sheahan NF, Dowling A, O'Reilly G, Malone JF: Commissioning and quality assurance protocol for dual energy X-ray absorptiometry (DEXA) systems. *Radiation protection dosimetry* 2005, **117**(1-3):288-290.
- 241. Taylor AE, Kuper H, Varma RD, Wells JC, Bell JD, K VR, Kulkarni B, Kinra S, Timpson NJ, Ebrahim S *et al*: Validation of dual energy X-ray absorptiometry measures of abdominal fat by comparison with magnetic resonance imaging in an Indian population. *PLoS One* 2012, 7(12):e51042.
- 242. WHO STEPwise approach to surveillance (STEPS). World Health Organization (WHO). Geneva. 2008. http://www.who.int/chp/steps/en/. Accessed 2013 April 12.
- 243. Waist Circumference and Waist-Hip Ratio: Report of a WHO Expert Concultation. World Health Organization. Geneva, 8-11 December 2008. Available: http://whqlibdoc.who.int/publications/2011/9789241501491 eng.pdf. Accessed 2013 April 12
- 244. Ross R, Berentzen T, Bradshaw AJ, Janssen I, Kahn HS, Katzmarzyk PT, Kuk JL, Seidell JC, Snijder MB, Sorensen TI *et al*: **Does the relationship between waist circumference, morbidity and mortality depend on measurement protocol for waist circumference?**Obesity reviews: an official journal of the International Association for the Study of Obesity 2008, **9**(4):312-325.
- 245. Ahima RS, Flier JS: **Adipose tissue as an endocrine organ**. *Trends Endocrinol Metab* 2000, **11**(8):327-332.
- 246. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW: Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004, 145(5):2273-2282.
- 247. Phillips LK, Prins JB: **The link between abdominal obesity and the metabolic syndrome**. *Curr Hypertens Rep* 2008, **10**(2):156-164.
- 248. Karlsson EA, Beck MA: **The burden of obesity on infectious disease**. *Exp Biol Med (Maywood)* 2010, **235**(12):1412-1424.
- 249. Palacio A, Lopez M, Perez-Bravo F, Monkeberg F, Schlesinger L: **Leptin levels are associated** with immune response in malnourished infants. *J Clin Endocrinol Metab* 2002, **87**(7):3040-3046.
- 250. Strandberg L, Verdrengh M, Enge M, Andersson N, Amu S, Onnheim K, Benrick A, Brisslert M, Bylund J, Bokarewa M *et al*: **Mice chronically fed high-fat diet have increased mortality and disturbed immune response in sepsis**. *PLoS One* 2009, **4**(10):e7605-.
- 251. Kanda N, Watanabe S: **Leptin enhances human beta-defensin-2 production in human keratinocytes**. *Endocrinology* 2008, **149**(10):5189-5198.
- 252. Falagas ME, Kompoti M: **Obesity and infection**. *Lancet InfectDis* 2006, **6**(7):438-446.
- 253. Kahn SE, Hull RL, Utzschneider KM: **Mechanisms linking obesity to insulin resistance and type 2 diabetes**. *Nature* 2006, **444**(7121):840-846.
- 254. Knapp S: Diabetes and Infection: Is There a Link? A Mini-Review. Gerontology 2012.
- 255. Pickkers P, Hoedemaekers A, Netea MG, de Galan BE, Smits P, van der Hoeven JG, van Deuren M: Hypothesis: Normalisation of cytokine dysbalance explains the favourable effects of strict glucose regulation in the critically ill. Neth J Med 2004, 62(5):143-150.
- 256. Liedtke S, Schmidt ME, Vrieling A, Lukanova A, Becker S, Kaaks R, Zaineddin AK, Buck K, Benner A, Chang-Claude J *et al*: **Postmenopausal sex hormones in relation to body fat distribution**. *Obesity (Silver Spring)* 2012, **20**(5):1088-1095.
- 257. Pedersen SB, Borglum JD, Brixen K, Richelsen B: **Relationship between sex hormones, body composition and metabolic risk parameters in premenopausal women**. *Eur J Endocrinol* 1995, **133**(2):200-206.
- 258. Rohrmann S, Shiels MS, Lopez DS, Rifai N, Nelson WG, Kanarek N, Guallar E, Menke A, Joshu CE, Feinleib M *et al*: **Body fatness and sex steroid hormone concentrations in US men:** results from NHANES III. *Cancer Causes Control* 2011, **22**(8):1141-1151.

- 259. Marriott I, Huet-Hudson YM: **Sexual dimorphism in innate immune responses to infectious organisms**. *Immunol Res* 2006, **34**(3):177-192.
- 260. Pennell LM, Galligan CL, Fish EN: Sex affects immunity. J Autoimmun 2012, 38(2-3):J282-291.
- 261. Pandey S, Maheshwari A, Bhattacharya S: **The impact of female obesity on the outcome of fertility treatment**. *J Hum Reprod Sci* 2010, **3**(2):62-67.
- 262. Baik I, Curhan GC, Rimm EB, Bendich A, Willett WC, Fawzi WW: **A prospective study of age** and lifestyle factors in relation to community-acquired pneumonia in US men and women. *Arch Intern Med* 2000, **160**(20):3082-3088.
- 263. Choban PS, Flancbaum L: **The impact of obesity on surgical outcomes: a review**. *J Am Coll Surg* 1997, **185**(6):593-603.
- 264. Choban PS, Heckler R, Burge JC, Flancbaum L: Increased incidence of nosocomial infections in obese surgical patients. *Am Surg* 1995, **61**(11):1001-1005.
- 265. Myles TD, Gooch J, Santolaya J: **Obesity as an independent risk factor for infectious morbidity in patients who undergo cesarean delivery**. *Obstet Gynecol* 2002, **100**(5 Pt 1):959-964.
- 266. Kwong JC, Campitelli MA, Rosella LC: **Obesity and respiratory hospitalizations during** influenza seasons in ontario, Canada: a cohort study. *Clin Infect Dis* 2011, **53**(5):413-421.
- Wertheim HF, van Kleef M, Vos MC, Ott A, Verbrugh HA, Fokkens W: **Nose picking and nasal carriage of Staphylococcus aureus**. *Infect Control Hosp Epidemiol* 2006, **27**(8):863-867.
- 268. Solberg CO: **Spread of Staphylococcus aureus in hospitals: causes and prevention**. *Scand J Infect Dis* 2000, **32**(6):587-595.
- 269. Sherertz RJ, Bassetti S, Bassetti-Wyss B: "Cloud" health-care workers. *Emerg Infect Dis* 2001, **7**(2):241-244.
- 270. Lu PL, Tsai JC, Chiu YW, Chang FY, Chen YW, Hsiao CF, Siu LK: **Methicillin-resistant**Staphylococcus aureus carriage, infection and transmission in dialysis patients, healthcare workers and their family members. *Nephrol Dial Transplant* 2008, **23**(5):1659-1665.
- 271. Mollema FP, Richardus JH, Behrendt M, Vaessen N, Lodder W, Hendriks W, Verbrugh HA, Vos MC: **Transmission of methicillin-resistant Staphylococcus aureus to household contacts**. *J Clin Microbiol* 2010, **48**(1):202-207.
- 272. Wagenvoort JH, De Brauwer EI, Sijstermans ML, Toenbreker HM: **Risk of re-introduction of methicillin-resistant Staphylococcus aureus into the hospital by intrafamilial spread from and to healthcare workers**. *J Hosp Infect* 2005, **59**(1):67-68.
- 273. Bagger JP, Zindrou D, Taylor KM: **Postoperative infection with meticillin-resistant Staphylococcus aureus and socioeconomic background**. *Lancet* 2004, **363**(9410):706-708.
- 274. Kazakova SV, Hageman JC, Matava M, Srinivasan A, Phelan L, Garfinkel B, Boo T, McAllister S, Anderson J, Jensen B *et al*: A clone of methicillin-resistant Staphylococcus aureus among professional football players. *N Engl J Med* 2005, **352**(5):468-475.
- 275. Wang JT, Liao CH, Fang CT, Chie WC, Lai MS, Lauderdale TL, Lee WS, Huang JH, Chang SC: Prevalence of and risk factors for colonization by methicillin-resistant Staphylococcus aureus among adults in community settings in Taiwan. *J Clin Microbiol* 2009, **47**(9):2957-2963.
- 276. Simoons-Smit AM, Savelkoul PH, Stoof J, Starink TM, Vandenbroucke-Grauls CM: Transmission of Staphylococcus aureus between humans and domestic animals in a household. Eur J Clin Microbiol Infect Dis 2000, 19(2):150-152.
- 277. Johnston CP, Stokes AK, Ross T, Cai M, Carroll KC, Cosgrove SE, Perl TM: **Staphylococcus aureus colonization among healthcare workers at a tertiary care hospital**. *Infect Control Hosp Epidemiol* 2007, **28**(12):1404-1407.
- 278. Schwarzkopf R, Takemoto RC, Immerman I, Slover JD, Bosco JA: **Prevalence of Staphylococcus aureus colonization in orthopaedic surgeons and their patients: a prospective cohort controlled study**. *J Bone Joint Surg Am* 2010, **92**(9):1815-1819.
- 279. Nulens E, Gould I, MacKenzie F, Deplano A, Cookson B, Alp E, Bouza E, Voss A:

 Staphylococcus aureus carriage among participants at the 13th European Congress of

- Clinical Microbiology and Infectious Diseases. *Eur J Clin Microbiol Infect Dis* 2005, **24**(2):145-148.
- 280. Elie-Turenne MC, Fernandes H, Mediavilla JR, Rosenthal M, Mathema B, Singh A, Cohen TR, Pawar KA, Shahidi H, Kreiswirth BN *et al*: **Prevalence and characteristics of Staphylococcus aureus colonization among healthcare professionals in an urban teaching hospital**. *Infect Control Hosp Epidemiol* 2010, **31**(6):574-580.
- 281. Suffoletto BP, Cannon EH, Ilkhanipour K, Yealy DM: **Prevalence of Staphylococcus aureus** nasal colonization in emergency department personnel. *Ann Emerg Med* 2008, **52**(5):529-533.
- 282. Blok HE, Troelstra A, Kamp-Hopmans TE, Gigengack-Baars AC, Vandenbroucke-Grauls CM, Weersink AJ, Verhoef J, Mascini EM: Role of healthcare workers in outbreaks of methicillin-resistant Staphylococcus aureus: a 10-year evaluation from a Dutch university hospital.

 Infect Control Hosp Epidemiol 2003, 24(9):679-685.
- 283. Tammelin A, Klotz F, Hambraeus A, Stahle E, Ransjo U: Nasal and hand carriage of Staphylococcus aureus in staff at a Department for Thoracic and Cardiovascular Surgery: endogenous or exogenous source? *Infect Control Hosp Epidemiol* 2003, **24**(9):686-689.
- 284. Eggen AE, Mathiesen EB, Wilsgaard T, Jacobsen BK, Njolstad I: **The sixth survey of the**Tromso Study (Tromso 6) in 2007-08: Collaborative research in the interface between
 clinical medicine and epidemiology: Study objectives, design, data collection procedures,
 and attendance in a multipurpose population-based health survey. *Scand J Public Health*2013, **41**(1):65-80.
- 285. Jacobsen BK, Eggen AE, Mathiesen EB, Wilsgaard T, Njolstad I: **Cohort profile: the Tromso Study**. *Int J Epidemiol* 2012, **41**(4):961-967.
- 286. Strommenger B, Braulke C, Heuck D, Schmidt C, Pasemann B, Nubel U, Witte W: **spa Typing of Staphylococcus aureus as a frontline tool in epidemiological typing**. *J Clin Microbiol* 2008, **46**(2):574-581.
- 287. Sangvik M, Olsen RS, Olsen K, Simonsen GS, Furberg AS, Sollid JU: **Age- and Gender- Associated Staphylococcus aureus spa Types Found among Nasal Carriers in a General Population: the Tromso Staph and Skin Study**. *J Clin Microbiol* 2011, **49**(12):4213-4218.
- 288. Leino A, Turpeinen U, Koskinen P: **Automated measurement of 25-OH vitamin D3 on the Roche Modular E170 analyzer**. *Clin Chem* 2008, **54**(12):2059-2062.
- 289. Jorde R, Sneve M, Hutchinson M, Emaus N, Figenschau Y, Grimnes G: **Tracking of serum 25-hydroxyvitamin D levels during 14 years in a population-based study and during 12 months in an intervention study**. *Am J Epidemiol* 2010, **171**(8):903-908.
- 290. Grimnes G, Almaas B, Eggen AE, Emaus N, Figenschau Y, Hopstock LA, Hutchinson MS, Methlie P, Mihailova A, Sneve M *et al*: **Effect of smoking on the serum levels of 25-hydroxyvitamin D depends on the assay employed**. *Eur J Endocrinol* 2010, **163**(2):339-348.
- 291. Little RR, Sacks DB: **HbA(1c)**: what do the numbers really mean? *Lancet* 2011, **378**(9796):1068-1069; author reply 1069-1070.
- 292. Nathan DM: International Expert Committee report on the role of the A1C assay in the diagnosis of diabetes. *Diabetes Care* 2009, **32**(7):1327-1334.
- 293. Scragg R, Holdaway I, Singh V, Metcalf P, Baker J, Dryson E: **Serum 25-hydroxyvitamin D3** levels decreased in impaired glucose tolerance and diabetes mellitus. *Diabetes Res Clin Pract* 1995, **27**(3):181-188.
- 294. Grimnes G, Emaus N, Joakimsen RM, Figenschau Y, Jenssen T, Njolstad I, Schirmer H, Jorde R: Baseline serum 25-hydroxyvitamin D concentrations in the Tromso Study 1994-95 and risk of developing type 2 diabetes mellitus during 11 years of follow-up. *Diabet Med* 2010, 27(10):1107-1115.
- 295. Konradsen S, Ag H, Lindberg F, Hexeberg S, Jorde R: **Serum 1,25-dihydroxy vitamin D is inversely associated with body mass index**. *Eur J Nutr* 2008, **47**(2):87-91.
- 296. Ogden CL, Lamb MM, Carroll MD, Flegal KM: **Obesity and socioeconomic status in adults: United States, 2005-2008**. *NCHS Data Brief* 2010(50):1-8.

- 297. Skramm I, Moen AE, Bukholm G: Nasal carriage of Staphylococcus aureus: frequency and molecular diversity in a randomly sampled Norwegian community population. *APMIS* 2011, 119(8):522-528.
- 298. Skramm I, Moen AE, Alm-Kristiansen K, Bukholm G: **Nasal carriage of Staphylococcus aureus:** which sequence types do orthopedic surgical healthcare workers carry? *Infect Control Hosp Epidemiol* 2007, **28**(6):737-739.
- 299. Aamot HV, Blomfeldt A, Skramm I, Muller F, Monecke S: Molecular characterisation of methicillin-sensitive Staphylococcus aureus from deep surgical site infections in orthopaedic patients. Eur J Clin Microbiol Infect Dis 2012, 31(8):1999-2004.
- 300. Szklo M, Nieto FJ: **Beyond the basics.** Jones and Bartlett Publishers. Sudbury, Massachusetts. 2007,pp. 110; 116-133; 151; 227.
- 301. Haldorsen B, Olsen K, Ericson Sollid J, Husom Haukland H, Sundsfjord A, Simonsen G, Breivik S, Larsen G, Furberg AS: Nasal and throat Swab cultures for the assessment of Staphylococcus aureus colonization and carriage. The Tromsø Staph and Skin Study. Abstract number 1150. 19th European Congress of Clinical Microbiology and Infectious Diseases (ECCMID). May 2009. Helsinki. Finland.
- 302. Verhoeven PO, Grattard F, Carricajo A, Lucht F, Cazorla C, Garraud O, Pozzetto B, Berthelot P: An algorithm based on one or two nasal samples is accurate to identify persistent nasal carriers of Staphylococcus aureus. Clin Microbiol Infect 2012, 18(6):551-557.
- 303. Nadas J, Putz Z, Kolev G, Nagy S, Jermendy G: Intraobserver and interobserver variability of measuring waist circumference. *Medical science monitor : international medical journal of experimental and clinical research* 2008, **14**(1):CR15-18.
- 304. Vatanparast H, Chilibeck PD, Cornish SM, Little JP, Paus-Jenssen LS, Case AM, Biem HJ: **DXA-derived abdominal fat mass, waist circumference, and blood lipids in postmenopausal women**. *Obesity (Silver Spring)* 2009, **17**(8):1635-1640.
- 305. Leekha S, Diekema DJ, Perencevich EN: **Seasonality of staphylococcal infections**. *Clin Microbiol Infect* 2012, **18**(10):927-933.
- 306. Wang Y, Jacobs EJ, McCullough ML, Rodriguez C, Thun MJ, Calle EE, Flanders WD: **Comparing** methods for accounting for seasonal variability in a biomarker when only a single sample is available: insights from simulations based on serum **25**-hydroxyvitamin d. *Am J Epidemiol* 2009, **170**(1):88-94.
- 307. Lamaro-Cardoso J, de Lencastre H, Kipnis A, Pimenta FC, Oliveira LS, Oliveira RM, Nouer SS, Aires-de-Sousa M, Milheirico C, Andrade AL: Molecular epidemiology and risk factors for nasal carriage of staphylococcus aureus and methicillin-resistant S. aureus in infants attending day care centers in Brazil. *J Clin Microbiol* 2009, **47**(12):3991-3997.
- 308. Hill BA: The environment and disease: association or causation? *Proc R Soc Med.*; 1965, **58**: 295-300.
- 309. Emonts M, Uitterlinden AG, Nouwen JL, Kardys I, Maat MP, Melles DC, Witteman J, Jong PT, Verbrugh HA, Hofman A *et al*: **Host polymorphisms in interleukin 4, complement factor H, and C-reactive protein associated with nasal carriage of Staphylococcus aureus and occurrence of boils.** *J Infect Dis* **2008, 197**(9):1244-1253.
- 310. Kinsman OS, McKenna R, Noble WC: **Association between histocompatability antigens (HLA)** and nasal carriage of Staphylococcus aureus. *J Med Microbiol* 1983, **16**(2):215-220.
- 311. Klein SL: The effects of hormones on sex differences in infection: from genes to behavior. Neurosci Biobehav Rev 2000, **24**(6):627-638.
- 312. Beery TA: **Sex differences in infection and sepsis**. *Crit Care NursClinNorth Am* 2003, **15**(1):55-62.
- 313. Simanski M, Dressel S, Glaser R, Harder J: **RNase 7 protects healthy skin from Staphylococcus aureus colonization**. *J Invest Dermatol* 2010, **130**(12):2836-2838.
- 314. Biswas B, Yenugu S: **Antimicrobial responses in the male reproductive tract of lipopolysaccharide challenged rats**. *Am J Reprod Immunol* 2011, **65**(6):557-568.

- 315. Han JH, Kim MS, Lee MY, Kim TH, Lee MK, Kim HR, Myung SC: **Modulation of human beta-defensin-2 expression by 17beta-estradiol and progesterone in vaginal epithelial cells**. *Cytokine* 2010, **49**(2):209-214.
- 316. Schauber J, Oda Y, Buchau AS, Yun QC, Steinmeyer A, Zugel U, Bikle DD, Gallo RL: **Histone** acetylation in keratinocytes enables control of the expression of cathelicidin and CD14 by 1,25-dihydroxyvitamin D3. *J Invest Dermatol* 2008, 128(4):816-824.
- 317. Gombart AF, Borregaard N, Koeffler HP: **Human cathelicidin antimicrobial peptide (CAMP)** gene is a direct target of the vitamin D receptor and is strongly up-regulated in myeloid cells by **1,25**-dihydroxyvitamin D3. *FASEB J* 2005, **19**(9):1067-1077.
- 318. Wang TT, Nestel FP, Bourdeau V, Nagai Y, Wang Q, Liao J, Tavera-Mendoza L, Lin R, Hanrahan JW, Mader S *et al*: **Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression**. *J Immunol* 2004, **173**(5):2909-2912.
- 319. Weber G, Heilborn JD, Chamorro Jimenez CI, Hammarsjo A, Torma H, Stahle M: **Vitamin D** induces the antimicrobial protein hCAP18 in human skin. *J Invest Dermatol* 2005, 124(5):1080-1082.
- 320. Turner J, Cho Y, Dinh NN, Waring AJ, Lehrer RI: **Activities of LL-37, a cathelin-associated antimicrobial peptide of human neutrophils**. *Antimicrob Agents Chemother* 1998, **42**(9):2206-2214.
- 321. Chesney RW: **Vitamin D and The Magic Mountain: the anti-infectious role of the vitamin**. *J Pediatr* 2010, **156**(5):698-703.
- 322. Zeigler CC, Persson GR, Wondimu B, Marcus C, Sobko T, Modeer T: **Microbiota in the oral subgingival biofilm is associated with obesity in adolescence**. *Obesity (Silver Spring)* 2012, **20**(1):157-164.
- 323. Kotani K, Tokunaga K, Fujioka S, Kobatake T, Keno Y, Yoshida S, Shimomura I, Tarui S, Matsuzawa Y: **Sexual dimorphism of age-related changes in whole-body fat distribution in the obese**. *Int J Obes Relat Metab Disord* 1994, **18**(4):207-202.
- 324. Meli R, Pacilio M, Raso GM, Esposito E, Coppola A, Nasti A, Di Carlo C, Nappi C, Di Carlo R: Estrogen and raloxifene modulate leptin and its receptor in hypothalamus and adipose tissue from ovariectomized rats. *Endocrinology* 2004, **145**(7):3115-3121.
- 325. Ford ES, Zhao G, Tsai J, Li C: **Associations between concentrations of vitamin D and concentrations of insulin, glucose, and HbA1c among adolescents in the United States**. *Diabetes Care* 2011, **34**(3):646-648.
- 326. Knekt P, Laaksonen M, Mattila C, Harkanen T, Marniemi J, Heliovaara M, Rissanen H, Montonen J, Reunanen A: **Serum vitamin D and subsequent occurrence of type 2 diabetes**. *Epidemiology* 2008, **19**(5):666-671.
- 327. Pujol M, Pena C, Pallares R, Ariza J, Ayats J, Dominguez MA, Gudiol F: **Nosocomial Staphylococcus aureus bacteremia among nasal carriers of methicillin-resistant and methicillin-susceptible strains**. *Am J Med* 1996, **100**(5):509-516.
- 328. Grimnes G, Figenschau Y, Almas B, Jorde R: Vitamin D, insulin secretion, sensitivity, and lipids: results from a case-control study and a randomized controlled trial using hyperglycemic clamp technique. *Diabetes* 2011, 60(11):2748-2757.
- 329. Sneve M, Figenschau Y, Jorde R: **Supplementation with cholecalciferol does not result in weight reduction in overweight and obese subjects**. *Eur J Endocrinol* 2008, **159**(6):675-684.
- 330. Nielsen NO, Skifte T, Andersson M, Wohlfahrt J, Soborg B, Koch A, Melbye M, Ladefoged K: Both high and low serum vitamin D concentrations are associated with tuberculosis: a case-control study in Greenland. *Br J Nutr* 2010, **104**(10):1487-1491.
- 331. Køber T: **Samfunnsspeilet 2-2011. Statistics Norway.** Oslo. 2011. Available: http://www.ssb.no/a/samfunnsspeilet/utg/201102/ssp.pdf. Accessed 2013 April 12.
- 332. Increase in health workers-2010. Statistics Norway 2011. Oslo. 2012 Available: http://www.ssb.no/english/subjects/06/01/hesospers_en/ Accessed 2013 April 12.

- 333. Miller M, Cook HA, Furuya EY, Bhat M, Lee MH, Vavagiakis P, Visintainer P, Vasquez G, Larson E, Lowy FD: **Staphylococcus aureus in the community: colonization versus infection**. *PLoS One* 2009, **4**(8):e6708.
- 334. Davis MF, Iverson SA, Baron P, Vasse A, Silbergeld EK, Lautenbach E, Morris DO: Household transmission of meticillin-resistant Staphylococcus aureus and other staphylococci. *Lancet Infect Dis* 2012, **12**(9):703-716.
- 335. Halablab MA, Hijazi SM, Fawzi MA, Araj GF: **Staphylococcus aureus nasal carriage rate and associated risk factors in individuals in the community**. *Epidemiol Infect* 2010, **138**(5):702-706.
- 336. Siegel JD, Rhinehart E, Jackson M, Chiarello L, Health Care Infection Control Practices Advisory C: **2007 Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Health Care Settings**. *American journal of infection control* 2007, **35**(10 Suppl 2):S65-164.

Paper I

Paper II

Paper III

Appendix A

Questionnaire from the 6th Tromsø Study

English translation



The form will be read electronically. Please use a blue or black pen You can not use comas, use upper-case letters.

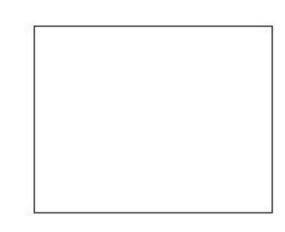
	2007 - 2008 Confidential	
1	HEALTH AND DISEASES How do you in general consider your own health to be?	Below you find a list of various problems. Have you experienced any of this during the last week (including today)? (Tick once for each complaint) No Little Pretty Very complaint complaint much much
	☐ Very good	Sudden fear without reason \(\sigma \square \
	☐ Neither and annied	Felt afraid or
	☐ Neither good nor bad	anxious
	□ Bad □ Variebad □	Faintness or dizziness
	☐ Very bad ☐	Felt tense or upset
2	How is your health compared to others in your age?	Tend to blame yourself
	☐ Much better	Sleeping problems
	☐ A little better	Depressed, sad
	☐ About the same	Feeling of being useless,
	☐ A little worse	worthless
	☐ Much worse Age first	Feeling that everything
3	Do you have, or have you had? Yes No time	regard to the future
	A heart attack	HEE AT HEALTH CERVICES
	Angina pectoris (heart cramp)	USE OF HEALTH SERVICES Have you during the last 12 months visited:
	Cerebral stroke/brain hemorrhage \Box	If YES; how many times?
	Atrial fibrillation	Yes No No. of times
	High blood pressure	General practitioner (GP)
	Osteoporosis	Psychiatrist/psychologist
	Asthma	(other than general practitioner/psychiatrist)
	Chronic bronchitis/Emphysema/COPD	Physiotherapist
	Diabetes	Chiropractor
	Psychological problems (for which you have sought help)	Alternative practitioner (homeopath, acupuncturist, foot zone therapist,
	Hypothyroidism	herbal medicine practitioner, laying on hands practitioner, healer, clairvoyant, etc.)
	Kidney disease, not including urinary tract infection (UTI)	Dentist/dental service
	Migraine	Have you during the last 12 months been to
4	Do you have persistent or constantly recurring pain that has lasted for 3 months or more?	a hospital? Yes No No. of times
	☐ Yes ☐ No	Admitted to a hospital
5	How often have you suffered from sleeplessness during	Had consultation in a hospital without admission;
	the last 12 months?	At psychiatric out-patient clinic
	Never, or just a few times□ 1-3 times a month	At another out-patient clinic 📙 📙
	Approximately once a week	Have you undergone any surgery during the last 3 years?
	☐ More that once a week	☐ Yes ☐ No —
		•

FAMILY AND FRIENDS USE OF MEDICINES 10 Do you currently use, or have you used some of 13 Who do you live with? (Tick for each question and give the number) the following medicines? (Tick once for each line) Yes No Number Never Spouse/partner used Now Earlier Other people older than 18 years.. \square Blood pressure lowering drugs П People younger than 18 years Cholesterol lowering drugs ... П Drugs for heart disease Tick for the relatives who have or have had Parents Children Siblings Diuretics \Box Drugs for A heart attack osteoporosis A heart attack before age of 60 \square Insulin Angina pectoris (heart cramp) Tablets for diabetes Cerebral stroke/brain haemorrhage The drugs for hypothyroidism Thyroxine/levaxin Osteoporosis Gastric/duodenal ulcers How often have you during the last 4 weeks used the following medicines? (Tick once for each line) Asthma Diabetes Not used Less than Every in the last every week, but Dementia 4 weeks week Daily not daily Psychological problems Painkillers on Substance abuse prescription П Painkillers non-15 Do you have enough friends who can give you prescription help when you need it? Sleeping pills ☐ Yes Tranquillizers Do you have enough friends whom you can talk confidentially with? Antidepressants .. How often do you normally take part in organised gatherings, e.g. sport clubs, political State the name of all medicines -both those on prescription and non-prescription drugs- you meetings, religious or other associations? have used regularly during the last 4 weeks. Do not include vitamins, minerals, herbs, natural ☐ Never, or just a few times a year remedies, other nutritional supplements, etc. 1-2 times a month Approximately once a week More than once a week WORK, SOCIAL SECURITY AND INCOME What is the highest level of education you have completed? (Tick once) Primary/secondary school, modern secondary school Technical school, vocational school, 1-2 years senior high school ☐ High school diploma College/university less than 4 years ☐ College/university 4 years or more If there is not enough space for all medicines, continue on a separate sheet. 19 What is your main activity? (Tick once) When attending you will be asked whether you have used antibiotics or painkillers the last 24 Full time work ☐ Housekeeping hours. If you have, you will be asked to provide the ☐ Part time work ☐ Retired/benefit recipient name of the drug, strength, dose and time of use. Unemployed Student/military service

20	Do you receive any of the following benefits? Old-age, early retirement or survivor pension Sickness benefit (on sick leave) Rehabilitation benefit Full disability pension Partial disability pension Unemployment benefits Transition benefit for single parents Social welfare benefits	26	How hard do you exercise on average? Easy- do not become short-winded or sweaty You become short-winded and sweaty Hard- you become exhausted For how long time do you exercise every time on averages Less than 15 minutes 30-60 minutes More than 1 hour
21	What was the household's total taxable income last year? Include income from work, pensions, benefits and similar ☐ Less than 125 000 NOK ☐ 401 000-550 000 NOK ☐ 125 000-200 000 NOK ☐ 551 000-700 000 NOK ☐ 201 000-300 000 NOK ☐ 701 000 -850 000 NOK ☐ 301 000-400 000 NOK ☐ More than 850 000 NOK	28	How often do you drink alcohol? Never Monthly or less frequently 2-4 times a month 2-3 times a week 4 or more times a week
22	Do you work outdoor at least 25% of the time, or in cold buildings (e.g. storehouse/industry buildings)? Yes No	29	How many units of alcohol (a beer, a glass of wine or a drink) do you usually drink when you drink alcohol? 1-2 5-6 10 or more 7-9
23	If you have paid or unpaid work, which statement describes your work best? Mostly sedentary work (e.g. office work, mounting) Work that requires a lot of walking (e.g. shop assistant, light industrial work, teaching) Work that requires a lot of walking and lifting (e.g. postman, nursing, construction) Heavy manual labour	30	in one occasion? Never Less frequently than monthly Monthly Weekly Daily or almost daily
24	 □ Reading, watching TV, or other sedentary activity. □ Walking, cycling, or other forms of exercise at least 4 hours a week (include walking or cycling to work, Sunday-walk/stroll, etc.) □ Participation in recreational sports, heavy gardening, etc. (note:duration of activity at least 4 hours a week) □ Participation in hard training or sports competitions, regularly several times a week 	34	5 (8)
25	How often do you exercise? (With exercise we mean for example walking, skiing, swimming or training/sports) Never Less than once a week Once a week 2-3 times a week Approximately every day		Age in years How many years in all have you smoked daily? Number of years Do you use or have you used snuff or chewing tobacco? No, never Yes, sometimes Yes, previously Yes, daily

	DIET		QUESTIONS FOR WOMEN		
38	38 Do you usually eat breakfast every day?		Are you pregnant at the moment?		
	☐ Yes ☐ No		\square Yes \square No \square Uncertain		
39	How many units of fruit or vegetables do you eat	47	How many children have you given birth to?		
39	on average per day? (units means for example a fruit, a cup of juice, potatoes, vegetables)		Number		
	Number of units	48	If you have given birth, fill in for each child:		
	Tumber of units		birth year, birth weight and months of breastfeeding (Fill in the best you can)		
40	How many times a week do you eat warm dinner?		Months of		
	Number		Child Birth year Birth weight in grams breastfeeding		
41	How often do you usually eat these foods?				
	(Tick once for each line)		2		
	0-1 2-3 1-3 4-6 1-2 times/ times/ times/ times/ time		3		
	mth mth week week day	′	4		
	Potatoes		5		
	Pasta/rice		6		
	Processed meat	49	Have you during pregnancy had high blood		
	(sausages, hamburger, etc.) \square \square \square \square \square		pressure?		
	Fruits, vegetables, berries		☐ Yes ☐ No		
	Lean fish	50	If yes, during which pregnancy?		
	Fatty fish	50	☐ The first ☐ Second or later		
42	How much do you usually drink the following? (Tick once for each line) 1-6 1 2-3 4 or more glasses glasses glasses glasses	51 e	Have you during pregnancy had proteinuria? ☐ Yes ☐ No		
	never /week /day /day /day	52	If yes, during which pregnancy?		
	Milk, curdled milk,		☐ The first ☐ Second or later		
	yoghurt	53	Were any of your children delivered prematurely (a month or more before the due date) because of preeclampsia?		
	•		☐ Yes ☐ No		
43	How many cups of coffee and tea do you drink daily? (Put 0 for the types you do not drink daily)	54	If yes, which child?		
	Number of cups		1st child 2nd child 3rd child 4th child 5th child		
	Filtered coffee				
	Boiled coffee (coarsely ground coffee for brewing)	55	How old were you when you started menstruating?		
	Other types of coffee		Age		
44	How often do you usually eat cod liver and roe? (i.e. "mølje")	56	Do you currently use any prescribed drug influencing the menstruation?		
	☐ Rarely/never ☐ 1-3 times/year☐ 4-6 times/y	year	Oral contraceptives, hormonal intrautrine or similar Yes No		
	\square 7-12 times/year \square More than 12 times/year		Hormone treatment for menopausal problems \square Yes \square No		
45	Do you use the following nutritional supplements?		When attending you will get supplementary		
+	Daily Sometimes Note that Cod liver oil or fish oil capsules]	questions about menstruation and any use of hormones. Write down on a sheet of paper the names of all the hormones you have used and bring it with you. You will also be asked whether your menstruation have ceased and possibly when and why.		

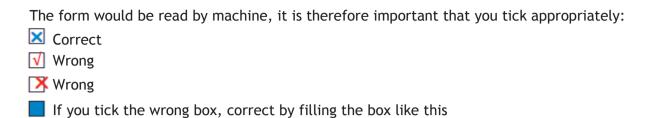








FILL OUT THE FORM IN THIS WAY:



Write the numbers clearly 1234567890

7 4 Correct 7 4 Wrong

Use only black or blue pen, do not use pencil or felt tip pen

1. DESCRIPTION OF YOUR HEALTH STATUS

By placing a tick in one box in each group below, please indicate which statements best describe your own health state today:

To allow you to show us how good or bad your state of health is we have made a scale (almost like a thermometer) where the best state of health you can imagine is marked 100 and the worst 0. We ask you to show your state of health by drawing a line from the box below to the point on the scale that best fits your state of health.

I.DI Mobility		Best imaginable
I have no problems in walking		health state — 100
about		± .00
I have some problems in walking about		‡
☐ I am confined to bed		100
		± /3
LD2 Self-care		‡
☐ I have no problems with self-care		± 80
☐ I have some problems washing or		±
dressing myself		‡
I am unable to wash or dress myself		+ 70
		‡ ‡
		<u> </u>
Usual activities (e.g. work, study, housework,		+ 60
family or leisure activities)		<u>‡</u>
\sqcup I have no problems with performing my	Your own health	‡ ‡
usual activities	state today	± 50
I have some problems with performing my usual activities		80 70 60 40 30 20
I am unable to perform my usual		± 40
activities		± 40
		‡
1.04 Pain and discomfort		± 30
☐ I have no pain or discomfort		‡
☐ I have moderate pain or discomfort		Ī
☐ I have extreme pain or discomfort		+ 20
		<u> </u>
		‡
Anxiety and depression		± 10
☐ I am not anxious or depressed		‡
I am moderately anxious or depressed		‡
I am extremely anxious or depressed		± 0
		Worst imaginable health state
		ricuttii state

3

2. CHILDHOOD/YOUTH AND AFFILIATION

 Where did you live at the age of 1 year? In Tromsø (with present municipal borders) In Troms, but not Tromsø In Finnmark In Nordland Another place in Norway 	What do you consfor one or more altonome.NorwegianSamiKven/FinnishAnother		e lf as? (Tick
Abroad	2.05 How many siblings you have/have you		en do
2.02 How was your family's financial situation during your childhood?	Number of siblings		
☐ Very good ☐ Good	Number of children.		
☐ Difficult ☐ Very difficult	2.06 Is your mother aliv		
 2.03 What is the importance of religion in your life? Very important Somewhat important Not important 	If NO: her age when Is your father alive Yes No If NO: his age when	?	
2.07 What was/is the highest completed education (Tick once for each column)	on for your parents and y Mother	our spouse Father	/partner? Spouse/ partner
7-10 years primary/secondary school, modern so Technical school, vocational school, 1-2 years so High school diploma	enior high school		

3. WELL BEING AND LIVING CONDITIONS

3.01 Below are three statements about satisfaction with statements about views on your own health. Show each of the statements by ticking in the box for the statements by ticking in the box for the statements.	v how you agree or disagree with
(tick once for each statement) Comple disagre	
In most ways my life is close to my ideal	
My life conditions are excellent	
I am satisfied with my life	
I have a positive view of my future health	
By living healthy, I can prevent serious diseases	
Below are four statements concerning your curre working now, the last job you had. (Tick once for	
Complet	
disagre My work is tiring, physically or mentally	e 1 2 3 4 5 6 7 agree
I have sufficient influence on when and how	
my work should be done	
I am being bullied or harassed at workI am being treated fairly at work	
ram being treated fairty at work	
I consider my occupation to have the following s (if you are not currently employed, think about you Very high status Fairly high status Medium status Fairly low status Very low status	
3.04 Have you over a long period experienced any of	<u> </u>
for each line)	Yes, Yes, Yes, No as a child as adult last year
Been tormented, or threatened with violence	
Been beaten, kicked at or victim of other types of violence	e 🗌 💮 💮
Someone in your close family have used alcohol or drugs in such a way that it has caused you worry	
If you have experienced anything of the above, how	much are you affected by that now?
☐ Not affected ☐ Affected to some extent	Affected to a large extent

4. ILLNESS AND WORRIES

4. Have you during the <u>last month</u> experienced any illness or injury?	If you suffer from sleeplessness monthly or more often, what time of the year does it
☐ Yes ☐ No	affect you most? (Put one or more ticks)
	\square No particular time
If YES: have you during the same period?	Polar night time
(Tick once for each line) Yes No	Midnight sun time
Been to a general practitioner	Spring and autumn
Been to a medical specialist	4.06 Have you had difficulty sleeping during
Been to emergency department	the past couple of weeks?
Been admitted to a hospital	Not at all ■ Not at all
Been to an alternative practitioner	No more than usual
(chiropractor, homeopath or similar)	Rather more than usual
4.02 Have you noticed sudden changes in your	☐ Much more than usual
pulse or heart rythm in the <u>last year</u> ?	(an Have your division the last two years folds
☐ Yes ☐ No	4.07 Have you during the last two weeks felt unhappy and depressed?
4.03 Do you become breathless in the following	
situations? (tick once for each question)	No more than usual
V N-	Rather more than usual
When you walk rapidly on level res No ground or up a moderate slope	☐ Much more than usual
When you walk calmly on level	(co. Have your division the last two years folds
ground	4.08 Have you during the last two weeks felt unable to cope with your difficulties?
While you are washing or dressing	Not at all
At rest	No more than usual
Uni De veri courb about deilu for come	Rather more than usual
4.04 Do you cough about daily for some periods of the year?	Much more than usual
Yes No	Mach more than asaac
	4.09 Below, please answer a few questions
If YES: Is the cough usually productive?	<pre>about your memory: (tick once for each question)</pre>
☐ Yes ☐ No	Yes No
	Do you think that your memory has declined?
Have you had this kind of cough for as long	Do you often forget where you
as 3 months in each of the last two years?	have placed your things?
☐ Yes ☐ No	Do you have difficulties finding
400 How often do you suffer from sleenlessness?	common words in a conversation?
4.05 How often do you suffer from sleeplessness? (tick once)	Have you problems performing
Never, or just a few times a year	daily tasks you used to master?
1-3 times a month	Have you been examined for
Approximately once a week	memory problems?
More than once a week	If YES to at least one of the first four questions
indie than once a week	above: Is this a problem in your daily life?
	☐ Yes ☐ No
- 6	-

4.11 Have you during the last last year suffered	4.16 To which degree have you had the following
from pain and/or stiffness in muscles or	complaints during the last 12 months?
joints in your neck/shoulders lasting for	Never Some Much
at <u>least 3 consecutive months</u> ? (tick once for each line)	Nausea
No Little Severe	Heartburn/regurgitation
complaint complaint complaint	t Diarrhoea
Neck, shoulders	Constipation
Arms, hands	Alternating diarrhoea
Upper part of the back	and constipation
The lumbar region	Bloated stomach
Hips, leg, feet	Abdominal pain
Other places	
	4.17 If you have had abdominal pain or
4. Have you suffered from pain and/or	discomfort during the last year: Yes No
stiffness in muscles or joints during the last 4 weeks? (tick once for each line)	Was it located in your upper stomach?.
Link.	Were you bothered as often as once a
No LITTLE Severe complaint complaint complaint	
Neck, shoulders	Do you feel symptoms relief after
Arms, hands	bowel movement?
Upper part of the back	Are the symptoms related to more frequent or rare bowel movements
The lumbar region	than normally?
Hips, leg, feet	Are the symptoms related to more
Other places	loose or hard stool than normally?
Other places	Do the symptoms appear after a meal?
4.12 Have you ever had: Age	4.18 Have you ever had: Age
Yes No last time Fracture in the	Yes No last time
wrist/forearm?	
	Gastric ulcer
Hip fracture?	Duodenal ulcer
4.13 Have you been diagnosed with arthrosis	Ulcer surgery
by a physician?	<u> </u>
☐ Yes ☐ No	4.9 For women: Have you ever had a
4.14 Do you have or have you ever had some	miscarriage? Yes No Do not know
of the following: Never Some Much	
Nickel allergy	If Yes: number of times
Pollen allergy	471 For more Have your partner ever had
Other allergies	4.20 For men: Have your partner ever had a miscarriage?
Other ditergles — — —	Yes No Do not know
4.15 Have you ever experienced infertility	If Yes: number of times
for more than 1 year?	ii res. number of times
Yes No	
If Yes: was it due to: Do not	4.21 Is your diet gluten-free?
Yes No know	YesNoDo not know
	4.22 Have you been diagnosed with
A condition concerning your	Dermatitis Herpetiformis (DH)?
partner?	☐Yes ☐ No ☐ Do not know
_	-+-

4.23 Have you been diagnosed with coeliac disease, based on a biopsy from your intestine taken in a gastroscopy examination? Yes No Do not know	4.30 What is the normal intensity of your headache attacks? Mild (do not hinder normal activity) Moderate (decrease normal activity) Strong (block normal activity)
 4.24 Do you have your natural teeth? Yes No 4.25 How many amalgam tooth fillings do you have/have you had? 0 1-5 6-10 10+ 	 4.3 What is the normal duration of the headache attacks? Less than 4 hours 4 hours - 1 day 1-3 days More than 3 days
4.26 Have you been suffering from headache the last year? Yes No If No: go to section 5, food habits 4.27 What kind of headache are you suffering from?	4.32 If you suffer from headache, when during the year does it affect you most? (tick one or more) No particular time Polar night time Midnight sun time Spring and/or Autumn
Migraine Other headache How many days per month do you suffer from headache? Less than one day 1-6 days 7-14 days More than 14 days	A.33 Before or during the headache, do you have a temporary: Yes No Visual disturbances? (flickering, blurred vision, flashes of light)
4.29 Is the headache attacks <u>usually:</u> (tick once for each line) Yes No Pounding/pulsatory pain	physical activity?

5. FOOD HABITS

5.01 How often do you usually eat	t the following?	(tick once for	r each line)	
		0-1 times			More than 3
		per montl	n per mont	th per week	times per week
Fresh water fish (not farmed)					
Salt water fish (not farmed)					
Farmed fish (salmon, trout, char)					
Tuna fish (fresh or canned)					
Fish bread spread					
Mussels, shells					
The brown content in crabs					
Whale or seal meat					
Pluck (liver/kidney/heart) from					
Pluck (liver/kidney/heart) from	ptarringari/grou	se			
5.02 How many times during the	year do/did yo	u usually eat 1	the followi	ing? (numbe	er of times)
			In ad	Iulthood In	childhood
Mølje (cod or pollack meat, li	ver, and roe)(Nu	mber of times pe	er year)		
Sea gull's egg (Number of eggs pe	r year)				
Reindeer meat (Number of times	per year)				
Local mushroom and wild berrie		nberries/cloudber ber of times per y			
5.03 How many times per month canned (tinned) foods (from Number	metal boxes)?	5.04 Do you ta suppleme	ents?	ns and/or m Sometime	ineral s Never
5.05 How often do you eat?	1-3 tim Never per mo		4-6 times per week	1-2 times 3 per day	times per day or more
		itii pei week	per week	per day	
Dark chocolate	📙 📙				
Light chocolate/milk chocolate					
Chocolate cake					
Other sweets					
5.06 If you eat chocolate, how m	uch do you usu	ally eat each	time?		
Compared with the size of a l		olade (a chocola	te brand in the	e market) and	describe how
much do you eat in relation to i	t. 1⁄4 1⁄2	1	1 ½	2 M	ore than 2
5.07 How often do you drink	1-3 tin		4-6 times	1-2 times	3 times per
cocoa/hot chocolate?	Never per mo	nth per week	per week	per day	day or more

6. ALCOHOL

B.D. How often have you in the last year: Never	Less than monthly	Monthly	Weekly	Daily or almost daily
Not been able to stop drinking alcohol when you have started?				
Failed to do what was normally expected of you because of drinking?				
yourself going after a heavy drinking session? Had feeling of guilt or remorse after				
drinking?				
Not been unable to remember what happened the night before because of your drinking?				
		Never	Yes, but not in the last year	
6.02 Have you or someone else been injured beca drinking?				
Has a relative, friend, physician, or other healt been concerned about your drinking or suggeste down?	d you to cut	:		
7. WEIGH	łT			
the <u>last 6 months</u> ?	Are you s weight? Yes	atisfied w \Box N	ith your pres	ent body
☐ Yes ☐ No If Yes: how many kilograms?			l you be satis	fied with
ii res. now many kitograms:		eal" weigh		- with
7.02 Estimate your body weight when you were 25 years old: Number of kilograms	Number of	kilograms	5	
8. SOLVEN	TS			
How many hours per week, do you do the following leisure- or professional activities: Automobile repair/paint, ceramic work, painting/varnishing/solvents, hair dressing, glazier, electrician. (Put 0 if you do not engage in such leisure or professional activities) Number of hours per week on average	Yes If Yes: How		or preparation	

9. USE OF HEALTH SERVICES

Have you ever experienced that diseases have been insufficiently examined or treated, and this had a serious consequence. Yes, this has happened to me Yes, this has happened to a close relative (child, parents, spouse) No	At the last visit to your GP, did you have a hard time to understand what the doctor(s) told you? Answer on a scale from 0 to 10, where 0 = they were difficult to understand and 10 = they were always easy to understand 0 1 2 3 4 5 6 7 8 9 10
If Yes, was it caused by? (tick once or more): general practitioner emergency medical doctor private practising specialist	How would you rate the treatment or counselling, you got at your last visit to your GP? Answer on a scale from 0 to 10, where 0 = worst treatment or counselling, and 10 = best treatment or counselling 0 1 2 3 4 5 6 7 8 9 10
 hospital doctor other health personnel alternative practitioner more than one person due to deficient 	9.07 During the last 12 months, how much of a problem, if any, was it to get a referral to special examinations (as x-ray, etc.) or to a specialist health care (private
routines and interaction 9.02 Have you ever felt persuaded to accept an examination or treatment that you did not want? Yes No	practising specialist or at hospital)? Not relevant No problem Some problem
If Yes, do you think this has had unfortunate consequences for your health? Yes No	Major problem 9.08 During the last 12 months, how much of a problem, if any, was it to get a referral to physiotherapist, chiropractor, etc.?
 Have you ever complained about a treatment you have received? Have never had a reason for complaining Have considered complaining, but 	Not relevant No problem Some problem Major problem
did not do Have complained verbally Have complained in writing	and Altogether, how much of a problem, if any, was it to get a referral to specialist health care?
How long have you had your current general practitioner/other physician? Less than 6 months 6 to 12 months 12 to 24 months More than 2 years	Not relevantVery difficultSome difficultiesEasyVery easy

9.10	During the last 12 months, have you been examined or treated by the specialist health care? Yes No	9.12	Have you ever, previous to the year 2002, had an operation at a hospital or a specialist clinic? Yes No
	If Yes, did you have a difficult time to understand what the doctor(s) told you? Answer on a scale from 0 to 10, where 0 = they were difficult to understand and 10 = they were always easy to understand	9.13	Have you, during the <u>last 12 months</u> , used herbal or natural medicine? Yes No
	0 1 2 3 4 5 6 7 8 9 10	9.14	Have you, during the <u>last 12 months</u> , used meditation, yoga, qi gong or thai chi as self-treatment?
9.11	How would you rate the treatment or counselling you got at your last visit to a specialist? Answer on a scale from 0 to 10, where 0 = worst treatment or counselling, and 10 = best treatment or counselling		☐ Yes ☐ No
	0 1 2 3 4 5 6 7 8 9 10		

10. USE OF ANTIBIOTICS

Have you used antibiotics during the last 12 m form of tablets, syrups or injections)	<u>nonth</u>	<u>s</u> ? (all p	enicillin	-like me	dicine	in the
Yes No Do not remembe	er					
If YES: What did you get the treatment for? Have you taken many antibiotic treatments, Treatment for each treatment.	eatment 1	Treatment 2	Treatment 3	Treatment 4	Treatmer 5	nt Treatment
 Urinary tract infection (bladder infection, cystitis) Respiratory tract infection (ear, sinus, throat or lung infection, bronchitis) 						
Other Treatment duration: number of days	-					
How did you acquire the antibiotics for treatmental Have you acquired many treatments, tick for ea		e.				
With prescription from a physician/dentist Without contacting a physician/without prescript Purchase from a pharmacy abroad Purchase over the internet Remnants from earlier treatment at home From family/friends Other ways	ion:					
Do you presently have antibiotics at home?	with	ıld you o nout con Yes		_		
If YES:is this after an agreement with your physician for treatment of chronic or frequently recurring disease? Yes No	such Com		n? (mul d	tiple ticl	ks are p	
If No: how did you acquire this antibiotic? (Multiple ticks are possible)	Bron	chitis throat				
Purchased from a pharmacy abroad		sitis				
Purchased over the internet	Feve	er				
Remnants from earlier treatment	Influ	ienza				
From family/friends	Ear i	infectior	۱			
Other ways	Diar	rhoea				
	Urin	ary trac	t infecti	on		
	Othe	er infect	ions			🔲

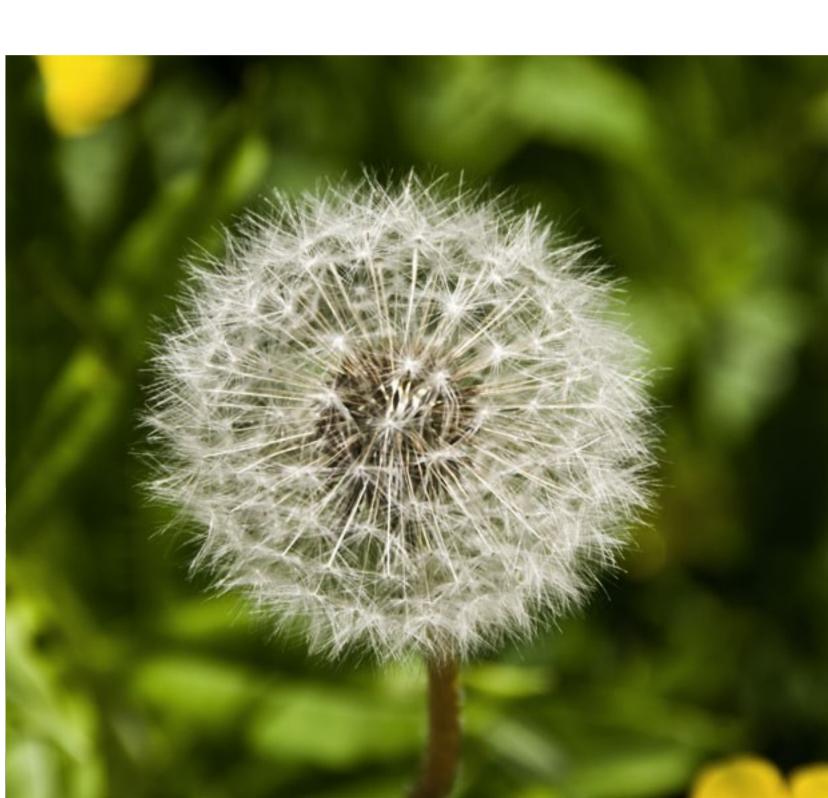
11. YOUR CIRCADIAN RHYTHM

We will ask you some questions about your sleeping habits Have you worked in a shift work schedule during the last 3 months? Yes No Number of days per week which you cannot freely choose when you sleep (e.g. work days)? Then I go to bed at I get ready to fall asleep at Number of minutes I need to fall asleep I wake up at With help of: Alarm clock External stimulus (noise, family members etc.) By myself Number of minutes I need to get up Number of days per week which you can freely choose when you sleep (e.g. free days or holidays Then I go to bed at I get ready to fall asleep at Number of minutes I need to fall asleep I wake up at With help of: Alarm clock External stimulus (noise, family members etc.) By myself Number of minutes I need to get up

12. SKIN AND DERMATOLOGY

or a bath? (tick once)	2.05 Have you often or always any of the following complaints? (tick once for each line
2 or more times daily	Swelling in the ankles or legs, Yes No
\square 1 time daily	particularly in the evenings
4-6 times per week	Varicose veins
2-3 times per week	Eczema (red, itchy rash) on
Once a week	your legs
Less than once a week	Leg pain that is getting worse when you are walking and is relieved when you are standing still
12.02 How often do you usually wash your	-
	2.88 Have you ever had the following diagnoses by a physician? (tick once for each line)
U 0 times	Yes No
1-5 times	Psoriasis
6-10 times	Atopic eczema
☐ 11-20 times	Rosacea
☐ More than 20 times	2.07 Have you recurring large acne/abscesses
Have you ever taken any antibiotics (penicillin and penicillin-like medicines) because of a skin disease, for example	that are tender/painful and often form scars in the following places? (tick once for each line)
infected eczema, acne, non-healing leg	Yes No
ulcers, recurrent abscess?	Armpits
☐ Yes ☐ No	Under the breasts
If Yes: How many times in average per year d	Stomach groove/the navel
you take antibiotics during the period you we	
most affected (tick once)	Around the anus
1-2 3-4 More than 4 times	The groin
Have you or have you ever had the followin skin disorders? (tick once for each line) Yes No	If Yes: Have you ever visited a physician because of abscesses? Yes No
Psoriasis	
Atopic eczema (children's eczema)	If Yes, did you get any of the following
Recurrent hand eczema \Box	treatments? (tick once for each line)
Recurrent pimples/spots for	Yes No
several months	Antibiotic ointment
Leg or foot ulcer that did not heal for 3-4 weeks	Antibiotic tablets
101 3 7 WCCN3	Surgical drainage
If YES on the question concerning leg and/or	A larger surgical intervention
foot ulcer, do you have any leg ulcer today?	including skin removal
☐ Yes ☐ No	Surgical laser treatment 🔲 🔲

Follow-up questions



INFORMATION TO FOLLOW-UP OUESTIONS

The following pages with questions should not be answered by everybody. If you have answered yes to one or more of questions below, we ask you to move on to the follow-up questions on the topic or topics you have answered yes to. The first four topics are from the first questionnaire and the last question is from this form.

We have for the sake of simplicity highlighted topics with different colours so that you will find the questions that applies to you.

If you answered YES to that you have: <u>long-term or recurrent pain that has lasted for 3 months or more</u>, please answer the questions on page 19 and 20. The margin is marked with green.

If you answered YES to that you have undergone any <u>surgery during the last 3 years</u>, please answer the questions on page 21 and 22. The margin is marked with purple.

If you answered YES to that you're <u>working outdoors at least 25% of the time</u>, or in facilities with low temperature, such as warehouse/industrial halls, please answer the questions on page 23. The margin is marked with red.

If you answered YES to that you have used <u>non-prescription pain relievers</u>, please answer questions on page 24. The margin is marked with orange.

If you answered YES to that you have or have ever had <u>skin problems</u> (such as psoriasis, atopic eczema, non-healing leg or foot ulcers, recurrent hand eczema, acne or abscesses), please answer the questions on page 25. The margin is marked with yellow.

If you have answered <u>NO</u> to these five questions, you are finished with your answers. The questionnaire is to be returned in the reply envelope you were given at the survey site. The postage is already paid.

Should you wish to give us written feedback on either the questionnaire or The Tromsø Study in general, you are welcome to that on page 26.

Do you have any questions, please contact us by phone or by e-mail. You can find the contact information on the back of the form. **THANK YOU** for taking the time to the survey and to answer our questions.

13. FOLLOW-UP QUESTIONS ON PAIN

You answered in the first questionnaire that you have protracted or constantly recurrent pain that has lasted for <u>3 months or more</u>. Here, we ask you to describe the pain a little closer.

13.01 How long have you had this pain?	
Number of years months	
How often do you have this pain? Every day Once a week or more Where does it hurt? (Tick for all locations whereurrent pain) Head/face Jaw/temporo-mandibular joint Neck Back Shoulder Arm/elbow Hand	Once a month or more Less than once a month ere you have protracted or constantly Thigh/knee/leg Ankle/foot Chest/breast Stomach Genitalia /reproductive organs Skin Other location
Hip	_ other tocation
What do you believe is the cause of the pain Accident /acute injury Long-term stress Surgical intervention/operation Herniated disk (prolapse) /lumbago Whiplash Migraine/headache Osteoarthritis Rheumatoid arthritis Bechterews syndrome Describe the other cause:	? (Tick for all known causes) Fibromyalgia Angina pectoris Poor blood circulation Cancer Nerve damage/neuropathy Infection Herpes zoster Another cause (describe below) Don't know
 Which kind of treatment have you received treatments you have received) No treatment Analgesic medications/painkillers Physiotherapy/chiropractic treatment Treatment at a pain clinic Surgery 	for the pain? (Tick for all types of pain Psycho-educative/relaxation training/ psychotherapy Acupuncture Complimentary and alternative medicine (homeopathy, healing, aromatherapy, etc. Other treatment

On a scale of 0 to 10, where 0 copossible pain you can imagine:	rrespon	ds to no pain and 10 corresponds to t	he worst
How strong would you say that the pain usually is?		0 1 2 3 4 5 6 7 8 9 10	Worst imaginable pain
How strong is the pain when it is in its strongest Intense?		0 1 2 3 4 5 6 7 8 9 10	Worst imaginable pain
To what degree does the pain interfere with your sleep?	No effect	0 1 2 3 4 5 6 7 8 9 10	Impossible to sleep
To what degree does the pain interfere with performing commor activities at home and at work?	No effect	0 1 2 3 4 5 6 7 8 9 10	Can not do anything

14. FOLLOW-UP QUESTIONS ON SURGERY

In the first questionnaire you answered that you have undergone an operation during $\underline{\text{the last 3 years.}}$

H. How many times have you undergone surge	
Number	
Below, please describe the operation. If you last 3 years, these questions concern the las	have undergone several operations during the t surgery you underwent.
Where in your body did you have surgery? (If you were operated simultaneously in several places in the body, tick more than once) Surgery in the head/neck/back · Head/face	Acute illness/trauma
Surgery in the chest · Heart	The hospital in Harstad
Breasts Another surgery in the chest region	Number of years Months
Surgery in the stomach/pelvis · Stomach/intestines	14.06 Do you have reduced sensitivity in an area near the surgical scar? Yes No
Another surgery in the stomach/pelvis	14.07 Are you hypersensitive to touch, heat or cold in an area near the surgical scar? Yes No
Surgery in the hip/legs · Hip/thigh	II. Does slight touch from clothes, showering or similar cause discomfort/pain?Yes No
Amputation	14.09 If you had pain at the site of surgery before you had surgery, do you have the same type of pain now? Yes No

14.10	The pain at the site of surgery: An 10=worst pain you can imagine	swer (on a sc	ale fro	om 0 t	to 10,	, wher	e 0=no p	+ ain and
H s	How strong pain did you have at the site of surgery before you had surgery	No pain	0 1	2 3	3 4 	5	6 7	8 9 10	Worst imaginable pain
	How strong pain do you normally nave at the site of surgery now	No pain	0 1	2 3	3 4 	5	6 7	8 9 10	Worst imaginable pain
h	How strong pain do you normally nave at the site of surgery when it s most intense	No pain	0 1	2 3	3 4] [5	6 7	8 9 10	Worst imaginable pain

15. FOLLOW-UP QUESTIONS ABOUT WORK IN COLD ENVIRONMENT

In the first questionnaire you answered yes to that you work in cold environments. Here are some follow-up questions that we hope you will answer.

15.01	Do you feel cold at work?	15.05	Have you had itching and/or rash in relation to cold exposure?
	Yes, often		Yes No
	Yes, sometimes		
	☐ No, never	15.0E	Have you during the <u>last 12 months</u> had
	For how long have you been exposed to cold air below 0°C during the last winter?		an accident where cold has been involved, and which required medical treatment? Yes No
	Leisure/hobbies (hours/week)		At work
	Work (hours/week)		In leisure time
	Outdoors, with suitable clothing (hours/week)	15.07	Do you experience any of the following symptoms while you are in a cold environment?
	Outdoors, without suitable clothing (hours/week)		If so, at what temperature do the symptoms occur?
			Yes No Under °C
	Indoors, with no heating (hours/week)		Breathing problems
	In cold, with wet clothing (hours/week)		Wheezy breathing
	Contact with cold objects/tools (hours/week)		Mucus secretion from lungs
			Chest pain
15.03	What ambient temperature prevents you from: Under °C		Disturbance in heart rhythm
	Olidei C		Impaired blood circulation
	Working outdoors		in hands/feet
	Training outdoors		Visual disturbance
			(short term/transient)
	Performing other activities outdoors		Migraine (short term/transient)
15.04	, <u> </u>		Fingers turning white
	frostbite with blisters, sores or skin injury?		(short term/transient)
	☐ Yes ☐ No		Fingers turning blue-red
	If Yes, how many times?		(short term/transient)
15.08	How does cold environments and cold-rela	ted	symptoms influence your performance? Decrease No effect Improve
	Concentration		
	Memory		
	Finger sensitivity (feeling)		
	Finger dexterity (motor)		
	Control of movement (for example tremor)		
	Heavy physical work		
	Long-lasting physical work		
_	23	3	+

16. USE OF NON-PRESCRIPTION PAINKILLERS

In the first questionnaire you answered that you had used non-prescription painkillers (analgesics) in the last 4 weeks. Here are some follow-up questions we hope you will answer.

16.01	What types of non-prescription painkillers have you used?	Phenazone with caffeine: (Antineuralgica, Fanalgin, Fenazon-koffein, Fenazon-koffein sterke)
		☐ Not used
	Paracetamol: (Pamol, Panodil, Paracet, Paracetamol, Pinex)	Less than every week
	☐ Not used	Every week, but not daily
	Less than every week	daily
	Every week, but not daily	How much do you usually take daily
	daily	when you use these medicines? (number of tablets)
	Have accele also concerned by taken also the	
	when you use these medicines? (number of tablets, suppositories)	For which complaints do you use non- prescription painkillers? (multiple ticks are possible)
		Headache
	Acetylsalicylates: (Aspirin, Dispril, Globoid)	☐ Menstrual discomfort
	Not used	☐ Migraine
	Less than every week	Back pain
	Every week, but not daily	Muscle/joint pain
	Daily	☐ Tooth pain
	How much do you usually take daily when you use these medicines? (number of tablets)	Other
		B.03 Do you think you have experienced side
	Ibuprofen: (Ibumetin, Ibuprofen, Ibuprox, Ibux)	effects of some of the medicines? (tick once for each line)
	☐ Not used	, 103 110
	Less than every week	Paracetamol
	Every week, but not daily	Acetylsalicylates
	Daily	Ibuprofen 📙 🖂
	How much do you usually take daily	Naproxen
	when you use these medicines? (number of tablets, suppositories)	Phenazone with caffeine
		16.04 Where do you usually purchase painkillers?
	Naproxen: (Ledox, Naproxen)	Pharmacy
	Not used	Grocery
	Less than every week	Petrol stations
	Every week, but not daily	Abroad
	Daily	☐ Internet
	How much do you usually take daily when you use these medicines?	
	(number of tablets)	IS.IS Do you combine the treatment with the use of painkillers on prescription?
		☐ Yes ☐ No

17. FOLLOW-UP QUESTIONS ABOUT SKIN DISEASES

On page 15 in this questionnaire you answered that you have or have had a skin disease. Here are some follow-up questions we hope you will answer.

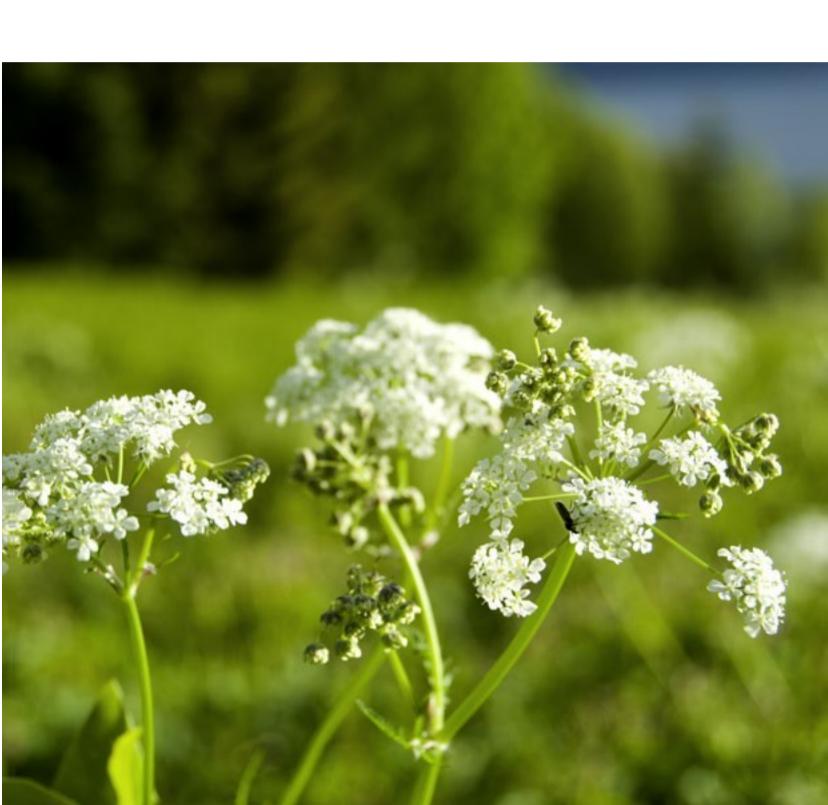
Answer on a scale from 0 to 10, where 0 corresponds to no symptoms and 10 correspond to worst imaginable complaints. If you answered YES to that you have or have had:

Psoriasis How much are you affected by your psoriasis today? How much are you affected by your psoriasis when it is most severe? Atopic eczema How much are you affected by	No worst imaginable of the complaints of the com
your atopic eczema today? · How much are you affected by your atopic eczema when it is most severe?	
 Hand eczema How much are you affected by your hand eczema today? How much are you affected by your hand eczema when it is most severe? 	
Acne How much are you affected by your acne today? How much are you affected by your acne when it is most severe?	
Abscesses · How much are you affected by your abscesses today? · How much are you affected by your abscesses when it is most severe?	
Menstrual periods Pregnancy Other	
How many episodes of abscesses do you usually have per year? (tick once) 0-1 4-6 2-3 More than 6	20-25 years Older than 50 years

FEEDBACK

Should you wish to give us a written feedback on either the questionnaire or The Tromsø Study in general, you are welcome to do it here:				

Thank you for your help





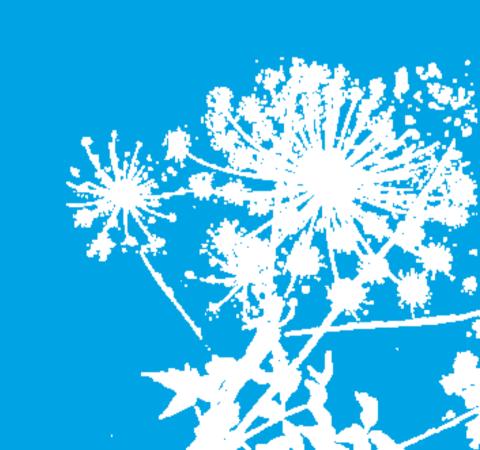
Tromsøundersøkelsen

The Tromsø Study

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9037 TROMSØ

Telephone: 77 64 48 16 Telefax: 77 64 48 31

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Appendix B

Questionnaire from the 6th Tromsø Study

Original questionnaire



	penn. Du kan ikke bruke komma, bruk b		
	2007 – 2008 KONFIDENSIELT		
1	HELSE OG SYKDOMMER Hvordan vurderer du din egen helse sånn i	6	Under finner du en liste over ulike problemer. Har du opplevd noe av dette <u>den siste uken</u> (til og med i dag)? (Sett ett kryss for hver plage)
	alminnelighet?		Ikke Litt Ganske Veldig plaget plaget mye mye
			Plutselig frykt uten grunn
	☐ Dårlig☐ Meget dårlig☐ ☐ Dårlig☐ ☐ Meget dårlig☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐ ☐		Matthet eller svimmelhet \square \square \square \square Føler deg anspent eller
2	Hvordan synes du at helsen din er sammenlignet med andre på din alder?		oppjaget
	☐ Mye bedre		Søvnproblemer
	 □ Litt bedre □ Omtrent lik □ Litt dårligere □ Mye dårligere 		Nedtrykt, tungsindig
3	Alder første Har du eller har du hatt? Alder første Ja Nei gang		Følelse av håpløshet mht. framtida
	Hjerteinfarkt	7	BRUK AV HELSETJENESTER Har du i løpet av de siste 12 måneder vært hos: Hvis JA; Hvor mange ganger? Ja Nei Ant ggr
	Hjerteflimmer (atrieflimmer)		Fastlege/allmennlege
	Astma		Legespesialist utenfor sykehus (utenom fastlege/allmennlege/psykiater)
	Psykiske plager (som du har søkt hjelp for)		Annen behandler (homøopat, akupunktør, fotsoneterapeut, naturmedisiner, håndspålegger, healer, synsk el.l)
	Nyresykdom, unntatt urinveisinfeksjon		Tannlege/tannpleier
4	Migrene	8	Har du i løpet av de siste 12 måneder vært på sykehus? Ja Nei Ant ggr
	smerter som har vart i <u>3 måneder eller mer</u> ? ☐ Ja ☐ Nei		Innlagt på sykehus 🗆 🗆 🗆
5	Hvor ofte har du vært plaget av søvnløshet de siste		Konsultasjon ved sykehus uten innleggelse;
J	12 måneder?		Ved psykiatrisk poliklinikk 🔲 🔲 📗
	Aldri, eller noen få ganger		Ved annen sykehuspoliklinikk \[\square \square \square \square \qq \qqq \qqq \qq
	☐ 1-3 ganger i måneden ☐ Omtrent 1 gang i uken ☐ Mer enn 1 gang i uken	9	Har du gjennomgått noen form for operasjon i løpet av de siste 3 årene? Ja Nei Heritage Her

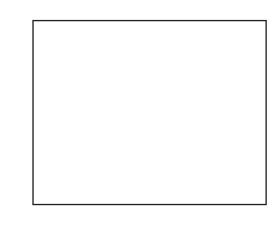
BRUK AV MEDISINER FAMILIE OG VENNER 10 Bruker du, eller har du brukt, noen av følgende 13 Hvem bor du sammen med? (Sett kryss for hvert medisiner? (Sett ett kryss for hver linje) spørsmål og angi antall) Alder → Ja Nei Antall første Ektefelle/samboer..... brukt Nå Før gang Andre personer over 18 år...... Medisin mot høyt blodtrykk... Personer under 18 år..... Kolesterolsenkende medisin.... Medisin mot hjertesykdom.... 14 Kryss av for de slektninger som har eller har hatt Foreldre Søsken Vanndrivende medisin..... Medisin mot beinskjørhet Hjerteinfarkt..... (osteoporose) П Hjerteinfarkt før fylte 60 år..... Insulin..... Angina pectoris (hjertekrampe)..... Diabetesmedisin (tabletter)...... Hjerneslag/hjerneblødning..... Stoffskiftemedisinene Beinskjørhet (osteoporose) Thyroxin/levaxin П Magesår/tolvfingertarmsår..... \Box Hvor ofte har du i løpet av de siste 4 ukene brukt følgende medisiner? (Sett ett kryss pr linje) Astma..... Diabetes Ikke brukt Sjeldnere Hver siste 4 enn hver uke, men Demens..... uker uke ikke daglig Daglig Psykiske plager..... Smertestillende Rusproblemer..... på resept..... Smertestillende 15 Har du nok venner som kan gi deg hjelp reseptfrie..... når du trenger det? Sovemidler..... ☐ Ja ☐ Nei Beroligende П medisiner..... 16 Har du nok venner som du kan snakke fortrolig med? Medisin mot ☐ Ja ☐ Nei depresjon..... 17 Hvor ofte tar du vanligvis del i foreningsvirksomhet 12 Skriv ned alle medisiner – både de med og uten som for eksempel syklubb, idrettslag, politiske lag, resept – som du har brukt regelmessig i siste 4 ukers religiøse eller andre foreninger? periode. (Ikke regn med vitaminer, mineraler, urter, Aldri, eller noen få ganger i året naturmedisin, andre kosttilskudd etc.) 1-2 ganger i måneden ☐ Omtrent 1 gang i uken ☐ Mer enn en gang i uken ARBEID, TRYGD OG INNTEKT 18 Hva er din høyeste fullførte utdanning? (Sett ett kryss) Grunnskole, framhaldsskole eller folkehøyskole Yrkesfaglig videregående, yrkesskole eller realskole Allmennfaglig videregående skole eller gymnas Høyskole eller universitet, mindre enn 4 år ☐ Høyskole eller universitet, 4 år eller mer Får du ikke plass til alle medisiner, bruk eget ark. 19 Hva er din hovedaktivitet? (Sett ett kryss) VED FRAMMØTE vil du bli spurt om du har brukt ☐ Yrkesaktiv heltid ☐ Hjemmeværende antibiotika eller smertestillende medisiner de siste 24 timene. Om du har det, vil vi be om at du oppgir ☐ Yrkesaktiv deltid Pensjonist/trygdet preparat, styrke, dose og tidspunkt Arbeidsledig Student/militærtjeneste

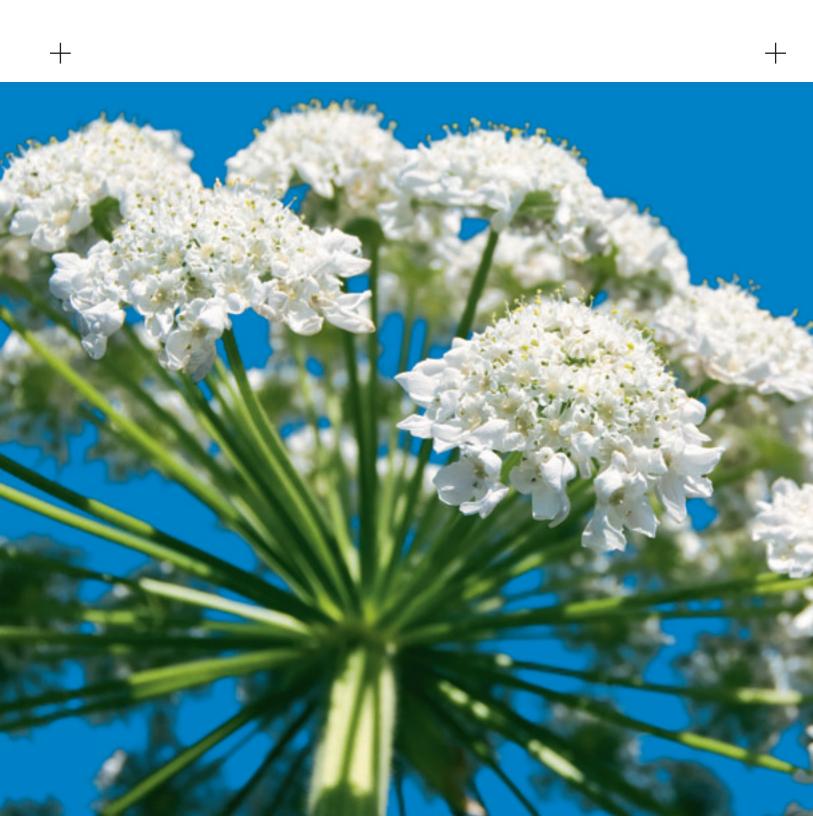
Hvor hardt mosjonerer du da i gjennomsnitt? ☐ Tar det rolig uten å bli andpusten eller svett. ☐ Tar det så hardt at jeg blir andpusten og svett ☐ Tar meg nesten helt ut Hvor lenge holder du på hver gang i gjennomsnitt? ☐ Mindre enn 15 minutter ☐ 30 minutter — 1 time ☐ 15-29 minutter ☐ Mer enn 1 time ALKOHOL OG TOBAKK
Hvor ofte drikker du alkohol? Aldri Månedlig eller sjeldnere 2-4 ganger hver måned 2-3 ganger pr. uke 4 eller flere ganger pr.uke Hvor mange enheter alkohol (en øl, et glass vin, eller
en drink) tar du vanligvis når du drikker? 1-2
Hvor ofte drikker du 6 eller flere enheter alkohol ved en anledning? □ aldri □ sjeldnere enn månedlig □ månedlig □ ukentlig □ daglig eller nesten daglig Røyker du av og til, men ikke daglig? □ la □ Noi
Har du røykt/røyker du daglig? □ Ja, nå □ Ja, tidligere □ Aldri Hvis du har røykt daglig tidligere, hvor lenge er det siden du sluttet? Antall år Hvis du røyker daglig nå eller har røykt tidligere: Hvor mange sigaretter røyker eller røykte du vanligvis daglig? Antall sigaretter □ □ Hvor gammel var du da du begynte å røyke daglig? Antall år Hvor mange år til sammen har du røykt daglig? Antall år Bruker du, eller har du brukt, snus eller skrå? □ Nei, aldri □ Ja, av og til □ Ja, men jeg har sluttet □ Ja, daglig

tuelt når og hvorfor.

-UNDBLAD MEDIA AS, TROMSØ, 77 75 32 50 - ONR 082222







SLIK FYLLER DU UT SKJEMAET:

Skjemaet vil bli lest maskinelt, det er derfor viktig at du krysser av riktig:

☑ Riktig
☑ Galt
☑ Galt

Om du krysser feil, retter du ved å fylle boksen slik

Skriv tydelige tall *1234567890*

 $\begin{array}{c}
7 \downarrow 4 \\
\hline
7 \downarrow 4
\end{array}$ Riktig $\begin{array}{c}
7 \downarrow 4
\end{array}$ Galt

Bruk kun sort eller blå penn, bruk ikke blyant eller tusj

1. BESKRIVELSE AV DIN HELSETILSTAND

Vis hvilke utsagn som passer best på din helsetilstand i dag ved å sette ett kryss i en av rutene utenfor hver av de fem gruppene nedenfor: 1.6 For at du skal kunne vise oss hvor god eller dårlig din helsetilstand er, har vi laget en skala (nesten som et termometer), hvor den beste helsetilstanden du kan tenke deg er markert med 100 og den dårligste med 0. Vi ber om at du viser din helsetilstand ved å trekke ei linje fra boksen nedenfor til det punkt på skalaen som passer best med din helsetilstand.

1.01 Gange	Best tenkelige helsetilstand
	 100
☐ Jeg har litt problemer med å gå omkring	-
☐ Jeg er sengeliggende	‡ + 90
	‡ ‡
1.02 Personlig stell	Ĭ
☐ Jeg har ingen problemer med personlig stell	+ 80
	‡ ‡
Jeg er ute av stand til å vaske meg eller	+ 70
kle meg	‡
1.03 Vanlige gjøremål (f.eks. arbeid, studier,	± 60
husarbeid, familie- eller fritidsaktiviteter)	‡
☐ Jeg har ingen problemer med å utføre mine vanlige gjøremål Nåværen	
☐ Jeg har litt problemer med å utføre mine vanlige gjøremål helsetilsta	and #
Jeg er ute av stand til å utføre mine vanlige gjøremål	± 40
· ····································	+
1.04 Smerte og ubehag	± 30
☐ Jeg har verken smerte eller ubehag	# 1
☐ Jeg har moderat smerte eller ubehag	‡
☐ Jeg har sterk smerte eller ubehag	+ 20
	‡
1.05 Angst og depresjon	± +10
☐ Jeg er verken engstelig eller deprimert	#
Jeg er noe engstelig eller deprimert	‡
☐ Jeg er svært engstelig eller deprimert	\pm_0
	Verst tenkelige helsetilstand

3

2. OPPVEKST OG TILHØRIGHET

2.01 Hvor bodde du da du fylte 1 år? I Tromsø (med dagens kommunegrenser) I Troms, men ikke i Tromsø I Finnmark fylke I Nordland fylke Annet sted i Norge	 2.04 Hva regner du deg selv som? (Kryss av for ett eller flere alternativ) Norsk Samisk Kvensk/Finsk Annet
□ I utlandet	2.05 Hvor mange søsken og barn har du/har du hatt?
2.02 Hvordan var de økonomiske forhold i familien under din oppvekst?	Antall søsken
☐ Meget gode	Antall barn
☐ Gode☐ Vanskelige☐ Meget vanskelige	2.06 Lever din mor?
and the state of t	Hvis NEI: hennes alder ved død
2.03 Hvilken betydning har religion i ditt liv? Stor betydning En viss betydning	Lever din far?
☐ Ingen betydning	Hvis NEI: hans alder ved død
2.07 Hva var/er den høyeste fullførte utdanning til de (sett ett kryss i hver kolonne) Grunnskole 7-10 år, framhaldsskole eller folkehø Yrkesfaglig videregående, yrkesskole eller realsk Allmennfaglig videregående skole eller gymnas. Høyskole eller universitet (mindre enn 4 år)	Ektefelle/ Mor Far samboer øyskole

3. TRIVSEL OG LIVSFORHOLD

3.01	Nedenfor står tre utsagn om tilfredshet med livet som et hele. Deretter står to utsagn om syn på din egen helse. Vis hvor enig eller uenig du er i hver av påstandene ved å sette et kryss i rubrikken for det tallet du synes stemmer best for deg. (sett ett kryss for hvert utsagn)									
		Helt						_		Helt
		uenig	1	2	3	4	5	6	7	enig
	På de fleste måter er livet mitt nær idealet mitt								Ц	
	Mine livsforhold er utmerkede									
	Jeg er tilfreds med livet mitt									
	Jeg ser lyst på min framtidige helse		Ш		Ш		Ш		Ш	
	Ved å leve sunt kan jeg forhindre alvorlige sykdommer									
3.02 Nedenfor står fire utsagn om syn på forhold ved din nåværende jobb, eller hvis du ikke er i arbeid nå, den jobben du hadde sist (sett ett kryss for hvert utsagn) Helt Helt										
	Arbeidet mitt er for belastende, fysisk eller følelsesmessig	uenig	1	2	3	4	5	6	7	enig
	Jeg har tilstrekkelig innflytelse på når og hvordan arbeidet mitt skal utføres									
	Jeg blir mobbet eller trakassert på arbeidsplassen min									
	Jeg blir rettferdig behandlet på arbeidsplassen min									
 Jeg opplever at yrket mitt har følgende sosiale status i samfunnet: (dersom du ikke er i arbeid nå, tenk på det yrket du hadde sist) Meget høy status Ganske høy status Middels status Ganske lav status Meget lav status 										
3.84 Har du over lengre tid opplevd noe av det følgende? (sett ett eller flere kryss for hver linje)										
		Nei	sor	Ja, n bar	'n		Ja, voks	en	Ja siste	•
	Blitt plaget psykisk, eller truet med vold								Г	7
	Blitt slått, sparket eller utsatt for annen type vold									j
	Noen i nær familie har brukt rusmidler på en slik									
	måte at dette har vært til bekymring for deg	🔲								
	Dersom du har opplevd noen av disse forholdene, h Ingen plager Noen plager	nvor my	•	ages o e pla		∕ dett	te <u>nå</u> î	?		

4. SYKDOMMER OG PLAGER

4.01 Har du i løpet av den siste måneden følt deg	Hvis du er plaget av søvnløshet månedlig								
syk eller hatt en skade?	eller oftere, når på året er du mest plaget? (sett ett eller flere kryss)								
∐ Ja □ Nei	☐ Ingen spesiell tid								
Hvis JA: har du i den samme perioden?	☐ Mørketida								
(sett ett kryss for hver linje) . Ja Nei	☐ Midnattsoltida								
Vært hos allmennlege/fastlege	☐ Vår og høst								
Vært hos spesialist	4.06 Har du i de siste par ukene hatt vansker								
Vært på legevakt	med å sove?								
Vært innlagt i sykehus	☐ Ikke i det hele tatt								
Vært hos alternativ behandler	☐ Ikke mer enn vanlig								
(kiropraktor, homøopat eller lignende)	☐ Heller mer enn vanlig								
4.02 Har du merket anfall med plutselig endring i	☐ Mye mer enn vanlig								
pulsen eller hjerterytmen siste året?	and the dealers of the control of the last tellings.								
☐ Ja ☐ Nei	4.07 Har du de siste par ukene følt deg ulykkelig og nedtrykt (deprimert)?								
4.03 Blir du tungpustet i følgende situasjoner?	☐ Ikke i det hele tatt								
(sett ett kryss for hvert spørsmål)	☐ Ikke mer enn vanlig								
Ja Nei	☐ Heller mer enn vanlig								
Når du går hurtig på flatmark eller svak oppoverbakke	☐ Mye mer enn vanlig								
Når du spaserer i rolig tempo på flatmark	4.08 Har du i de siste par ukene følt deg ute av stand til å mestre dine vanskeligheter?								
Når du vasker deg eller kler på deg 🗌 🗌	☐ Ikke i det hele tatt								
Når du er i hvile 🔲 🔲	☐ Ikke mer enn vanlig								
	Heller mer enn vanlig								
4.04 Hoster du omtrent daglig i perioder av året?	☐ Mye mer enn vanlig								
☐ Ja ☐ Nei	,								
Hvis JA: Er hosten vanligvis ledsaget av	4.09 Nedenfor ber vi deg besvare noen spørsmål om din hukommelse: (sett ett kryss for hver								
oppspytt?	spørsmål)								
☐ Ja ☐ Nei	Ja Nei								
	Synes du at din hukommelse har blitt dårligere?								
Har du hatt slik hoste så lenge som i en 3 måneders periode i begge de to siste årene?	Glemmer du ofte hvor du har lagt								
☐ Ja ☐ Nei	tingene dine?								
□ Ja □ INCI	Har du problemer med å finne								
4.05 Hvor ofte er du plaget av søvnløshet?	vanlige ord i en samtale?								
(sett ett kryss)	Har du fått problemer med daglige gjøremål som du mestret tidligere?								
☐ Aldri, eller noen få ganger i året	Har du vært undersøkt for								
☐ 1-3 ganger i måneden	sviktende hukommelse?								
☐ Omtrent 1 gang i uka	Hvis IA på minst ett av de fire første snørs								
☐ Mer enn 1 gang i uka	Hvis JA på minst ett av de fire første spørs- målene ovenfor: Er det et problem i hverdagen?								
	☐ Ja ☐ Nei								
-	-								

<u> </u>	
4.10 Har du i løpet av det siste året vært plaget med smerter og/eller stivhet i muskler og ledd som har vart i <u>minst 3 måneder</u> sammen-	4.16 I hvilken grad har du hatt følgende plager i de siste 12 måneder? Aldri Litt Mye
hengende? (sett ett kryss i hver linje)	Kvalme
Ikke En del Sterkt	Halsbrann/sure oppstøt
plaget plaget plaget	
Nakke, skuldre	Diare U U
Armer, hender	Treg mage
Øvre del av ryggen \Box \Box	Vekslende treg mage
Korsryggen	og diare
	Oppblåsthet
Hofter, ben, føtter \square	Smerter i magen 📙 📙 📙
Andre steder	on the same had an arter sallen about a fee
4.11 Har du vært plaget med smerter og/eller stivhet i muskler og ledd i løpet av de	4.17 Hvis du har hatt smerter i eller ubehag fra magen siste året: Ja Nei
siste 4 ukene? (sett ett kryss i hver linje)	Er disse lokalisert øverst i magen?
Ikke En del Sterkt	Har du hatt plagene så ofte som 1 dag
plaget plaget plaget	i uka eller mer de siste 3 måneder?
Nakke, skuldre	Blir plagene bedre etter avføring? 📙 📙
Armer, hender	Har plagene sammenheng med
Øvre del av ryggen \square \square \square	hyppigere eller sjeldnere avføring
Korsryggen	enn vanlig?
Hofter, ben, føtter	Har plagene noen sammenheng med løsere eller fastere avføring enn vanlig? \
Andre steder	
	Kommer plagene etter måltid? 📙 📙
4.12 Har du noen gang hatt: Alder Ja Nei siste gang	4.18 Har du noen gang hatt: Alder Ja Nei siste gang
Brudd i håndledd/ underarm?	Sår på magesekken 🔲 🔲 🔟
Lårhalsbrudd?	Sår på tolvfingertarmen 🔲 🔲
4.13 Har du fått stilt diagnosen slitasjegikt av lege? ☐ Ja ☐ Nei	Magesår-operasjon
∐ Ja	4.19 Til kvinnen: Har du spontanabortert?
4.14 Har eller har du hatt noen av følgende:	☐ Ja ☐ Nei ☐ Vet ikke
Aldri Litt Mye Nikkelallergi 🗌 🔲	Hvis JA, antall ganger
Pollenallergi	
	4.20 Til mannen: Har din partner noen gang spontanabortert?
Andre allergier	<u> </u>
4.15 Har du opplevd ufrivillig barnløshet i mer	☐ Ja ☐ Nei ☐ Vet ikke
enn 1 år?	Hvis JA, antall ganger
☐ Ja ☐ Nei	
	4.21 Bruker du glutenfri diett?
Hvis JA, skyldtes dette: Vet	☐ Ja ☐ Nei ☐ Vet ikke
Ja Nei ikke	
	4.22 Har du fått stilt diagnosen Dermatitis
Forhold hos deg selv?	Herpetiformis (DH)?
Forhold hos partneren?	∐ Ja ☐ Nei ☐ Vet ikke
	, <u>-</u>

Har du fått stilt diagnosen cøliaki på bakgrunn av en vevsprøve fra tynntarmen tatt under en undersøkelse der du svelget en slange (gastroskopi)?	4.30 Hvor sterk er hodepinen vanligvis? Mild (hemmer ikke aktivitet) Moderat (hemmer aktivitet) Sterk (forhindrer aktivitet)
4.24 Har du egne tenner? Ja Nei 4.25 Hvor mange amalgamfyllinger har du/har du hatt? 0 1-5 6-10 10+	4.31 Hvor lenge varer hodepinen vanligvis? Mindre enn 4 timer 4 timer – 1 døgn 1-3 døgn Mer enn 3 døgn
4.26 Har du vært plaget av hodepine det siste året? Ja Nei Hvis NEI, gå til del 5, kosthold	4.32 Dersom du er plaget av hodepine, når på året er du plaget mest? (sett ett eller flere kryss) Ingen spesiell tid Mørketida Midnattsoltida
 4.27 Hva slags hodepine er du plaget av? Migrene	Vår og/eller høst 4.33 Før eller under hodepinen, kan du da ha forbigående: Synsforstyrrelse? (takkede linjer, flimring, tåkesyn, lysglimt)
4.29 Er hodepinen vanligvis: (sett et kryss for hver linje) Ja Nei Bankende/dunkende smerte	Kvalme og /eller oppkast

5. KOSTHOLD

5.01 Hvor ofte spiser du vanligvis følgende? (sett ett kryss i hver linje)						
			0-1 g per mnd	2-3 g per mnd	1-3 g per uke	Mer enn 3 g per uke
Ferskvannsfisk (ikke oppdrett)						
Saltvannsfisk (ikke oppdrett)					님	
Oppdrettsfisk (laks, røye, ørret)					H	H
Tunfisk (fersk eller hermetisert) Fiskepålegg				H	H	H
Skjell				П	П	Ħ
Den brune innmaten i krabbe			_			
Hvalkjøtt/sel/kobbekjøtt						
Innmat fra rein eller elg			🔲			
Innmat fra rype			📙			
5.02 Hvor mange ganger i året spiser	du/spist	te du vanlig	gvis følgend			din barndom
Mølje (Antall ganger i året)						
Måsegg (Antall egg i året)						
Reinsdyrkjøtt (Antall ganger i året)				<u>L</u>		
Selvplukket sopp og bær (blåbær/t	yttebær/n	nulte) (Antall	ganger i året)			
5.03 Hvor mange ganger i måneden sp hermetiske matvarer (fra metallbo	iser du kser)?	_	Bruker du vit	_	/eller min e Iblant	eraltilskudd?
5.03 Hvor mange ganger i måneden sp hermetiske matvarer (fra metallbo Antall	kser)?	_	_	_		_
hermetiske matvarer (fra metallbo	kser)?		☐ Ja, daglig 1-3 g	g □ 4-6 g.	Iblant 1-2 g.	_
hermetiske matvarer (fra metallbo Antall	kser)? Aldri	1-3 g	☐ Ja, daglig 1-3 g	g □ 4-6 g.	Iblant 1-2 g.	☐ Aldri 3 g. per dag
hermetiske matvarer (fra metallbo Antall	Aldri	1-3 g	☐ Ja, daglig 1-3 g	g □ 4-6 g.	Iblant 1-2 g.	☐ Aldri 3 g. per dag
hermetiske matvarer (fra metallbo Antall	Aldri	1-3 g	☐ Ja, daglig 1-3 g	g □ 4-6 g.	Iblant 1-2 g.	☐ Aldri 3 g. per dag
hermetiske matvarer (fra metallbo Antall 5.05 Hvor ofte spiser du? Mørk sjokolade Lys sjokolade/melkesjokolade	Aldri	1-3 g	☐ Ja, daglig 1-3 g	g □ 4-6 g.	Iblant 1-2 g.	☐ Aldri 3 g. per dag
hermetiske matvarer (fra metallbo Antall 5.05 Hvor ofte spiser du? Mørk sjokolade Lys sjokolade/melkesjokolade Sjokoladekake Andre søtsaker 5.06 Hvis du spiser sjokolade, hvor m	Aldri	1-3 g per mnd	Ja, daglig 1-3 g per uke	4-6 g. per uke	1-2 g. per dag	Aldri 3 g. per dag eller mer
hermetiske matvarer (fra metallbo Antall 5.05 Hvor ofte spiser du? Mørk sjokolade Lys sjokolade/melkesjokolade Sjokoladekake Andre søtsaker	Aldri	1-3 g per mnd	Ja, daglig 1-3 g per uke	4-6 g. per uke	1-2 g. per dag	Aldri 3 g. per dag eller mer
hermetiske matvarer (fra metallbo Antall 5.05 Hvor ofte spiser du? Mørk sjokolade Lys sjokolade/melkesjokolade Sjokoladekake Andre søtsaker 5.06 Hvis du spiser sjokolade, hvor m	Aldri ye pleie Lunsj sjo	1-3 g per mnd	Ja, daglig 1-3 g per uke	4-6 g. per uke	1-2 g. per dag	Aldri 3 g. per dag eller mer
hermetiske matvarer (fra metallbo Antall 5.05 Hvor ofte spiser du? Mørk sjokolade Lys sjokolade/melkesjokolade Sjokoladekake Andre søtsaker 5.06 Hvis du spiser sjokolade, hvor m	Aldri Aldri ye pleie Lunsj sjo	1-3 g per mnd	Ja, daglig 1-3 g per uke	4-6 g. per uke per uke power gang? mye du sp 1 ½ 4-6 g.	Iblant 1-2 g. per dag □ □ □ □ coiser i forh 2 □ 1-2 g.	Aldri 3 g. per dag eller mer Cold til den. Mer enn 2 Cold til den. Mer enn 2
hermetiske matvarer (fra metallbo Antall	Aldri ye pleie Lunsj sjo	1-3 g per mnd	Ja, daglig 1-3 g per uke	4-6 g. per uke per uke power gang? mye du sp 1 ½ 4-6 g.	Iblant 1-2 g. per dag □ □ □ □ coiser i forh 2	Aldri 3 g. per dag eller mer

6. ALKOHOL

6.01 Hvor ofte har du <u>det siste året</u> :	Aldri	Sjeldnere enn månedlig	Månedlig	Ukentlig	Daglig, eller nesten daglig		
Ikke klart å stoppe og drikke alkohol når du først har begynt?							
Ikke klart å gjøre det som normalt forventes av deg fordi du har drukket?							
Trengt en drink om morgenen for å få komme i gang etter en rangel?							
Følt skyld eller anger etter at du har drukket?							
Ikke klart å huske hva som skjedde kvelden før på grunn av at du hadde drukket?							
			Aldri	Ja, men ikke det siste året			
6.02 Har du eller andre noen gang blitt skadet drukket?			ar				
Har en slektning, venn, lege, eller annet hel bekymret for din drikking, eller foreslått at d			et? 🗌				
7	. VEI	KT					
7.01 Har du ufrivillig gått ned i vekt <u>siste 6</u> måneder? Da Nei	7.0	□ Ja	øyd med ve □ Ne				
Hvis JA: Hvor mange kilo?	7.0	4 Hvilken ve trivselsvek		være tilfreds	med (din		
7.02 Anslå din vekt da du var 25 år gammel:		Antall kg					
Antall hele kg							
8. LØSEMIDLER							
Bilreparasjoner/lakkering, keramikkarbeid, maling/lakkering/løsemidler, frisør, glassmestelektriker (Sett 0 om du ikke driver med slike fritids eller yrkesaktiviteter) Antall timer per uke i gjennomsnitt	ter,	Bruker du l Ja Hvis JA, hv	☐ Ne		?		

9. BRUK AV HELSETJENESTER

9.01 Har du noen gang opplevd at sykdom er blitt mangelfullt undersøkt eller behandlet, og at dette har gitt alvorlige følger?	9.05 Ved siste legebesøk hos fastlegen, snakket legen(e) til deg slik at du forsto dem? Svar på en skala fra 0 til 10, hvor 0=de var vanskelige
\square Ja, det har rammet meg selv	å forstå og 10=de var alltid enkle å forstå
Ja, det har rammet en nær pårørende (barn, foreldre, ektefelle/samboer)	0 1 2 3 4 5 6 7 8 9 10
☐ Nei	
Hvis JA, hvor mener du årsaken ligger? (sett ett eller flere kryss): hos fastlege/allmennlege hos legevaktslege hos privatpraktiserende spesialist	9.06 Hvordan vil du karakterisere behandlingen eller rådgivingen du fikk siste gang du var hos lege? Svar på en skala fra 0 til 10, hvor 0= meget dårlig behandling og 10 = meget god behandling 0 1 2 3 4 5 6 7 8 9 10
hos sykehuslege	
hos annet helsepersonell hos alternativ behandler hos flere på grunn av svikt i rutiner og samarbeid Har du noen gang følt deg overtalt til å godta undersøkelse eller behandling som du selv ikke ønsket? Ja Nei	9.07 Har du i løpet av de siste 12 måneder opplevd at det har vært vanskelig å bli henvist til spesielle undersøkelser (som røntgen eller liknende) eller til spesialisthelsetjenesten (privatpraktiserende spesialist eller ved sykehus)? Ikke aktuelt Intet problem Stort problem
Hvis JA, mener du dette har hatt uheldige helsemessige følger? ☐ Ja ☐ Nei	9.08 Har du i løpet av de siste 12 måneder opplevd at det er vanskelig å bli henvist til fysioterapeut, kiropraktor eller liknende?
9.03 Har du noen gang klaget på behandling	☐ Ikke aktuelt
du har fått?	☐ Intet problem
☐ Har aldri vært aktuelt	☐ Noe problem
🗌 Har vurdert å klage, men ikke gjort det	☐ Stort problem
☐ Har klaget muntlig	9.09 Alt i alt, har du opplevd at det er vanskelig
☐ Har klaget skriftlig	eller enkelt å bli henvist til spesialisthelse- tjenesten?
9.04 Hvor lenge har du hatt din nåværende	☐ Ikke aktuelt
fastlege/annen lege?	☐ Meget vanskelig
☐ Mindre enn 6 måneder	☐ Noe vanskelig
☐ 6 til 12 måneder☐ 12 til 24 måneder	☐ Rimelig enkelt
☐ Mer enn 2 år	☐ Meget enkelt
∟ IVICI CIIII∠ al	

9.10 Har du i løpet av de <u>siste 12 måneder</u> vært til undersøkelse eller behandling i spesialisthelsetjenesten?	9.12 Har du noen gang <u>før 2002</u> gjennomgått en operasjon på sykehus eller spesialist- klinikk?
Hvis JA, snakket legen(e) til deg slik at du forstod dem? Svar på en skala fra 0 til 10, hvor 0=de var vanskelige å forstå og 10=de var alltid enkle å forstå	9.13 Har du i løpet av de <u>siste 12 måneder</u> brukt urtemedisin , naturmidler eller naturlegemidler?
0 1 2 3 4 5 6 7 8 9 10	9.14 Har du i løpet av de <u>siste 12 måneder</u> brukt meditasjon, yoga, qi gong eller thai chi som egenbehandling?
Hvordan vil du karakterisere behandlingen eller rådgivningen du fikk siste gang du var hos spesialist? Svar på en skala fra 0 til 10, hvor 0=meget dårlig og 10=meget god	□ Ja □ Nei
0 1 2 3 4 5 6 7 8 9 10	

10. BRUK AV ANTIBIOTIKA

0.01		antibiotika i lø stur eller sprøy		e 12 måne	eder? (all	l penicil	linlikner	nde me	disin i fo	orm av		
	☐ Ja	☐ Nei	☐ Husker ik	ke								
	flere antibiot	fikk du behand ikakurer, sett e	tt kryss for hve	er kur.	Kur 1	Kur 2	Kur 3	Kur 4	Kur 5	Kur 6		
		nfeksjon (blæreb				Ш	Ш	Ш	Ш			
	bronkitt)	nfeksjon (øre-, bi			. 📙							
	· Annet				. <u> </u>							
	Antall dagers	antibiotika ku	r		. 📖							
		ffet du deg ant ett ett kryss for		Har du ta	.tt							
		ra lege/tannleg										
	Uten kontakt	t med lege/ute	n resept:							_		
		ekte fra apotek								닏		
	, , ,	nnom Internett										
		tidligere kur tilg					\vdash			\vdash		
		amilie/venner								님		
	· Andre m	åter			Ц	Ш		Ш	Ш	Ш		
0.02	Har du antibi	iotika hjemme?	•		Kan du t			uke anti	ibiotika	uten å		
	□ Ja	☐ Nei			kontakte □ Ja	e lege ii	Nei					
		ette etter avtale			Hvis JA,	ا ماانده	tiletand	مديناط	ı i cå fal	11		
	å behandle k vendende syl	ronisk eller hyp kdom?	opig tilbake-		behandl				i i sa iai	11		
	□ Ja	□ Nei			Forkjøle		•	•				
	∟ ја	□ IVEI			, Hoste							
	Hvis Nei, hvo	ordan skaffet di	u deg dette		Bronkitt							
	legemiddelet	? (Flere kryss e	r mulig)		Halsbete							
	Kjøpt direkte	fra apotek i ut	landet		Bihulebe							
	Kjøpt over In	ternett								_		
	Rest fra tidlig	gere kur			FeberL Influensa							
	Fått av famili	e/venner			Ørebete	nnelse						
	Andre måter				Diaré					_		
					Blærebe	tennels	e					
					Andre ir							

11. DIN DØGNRYTME

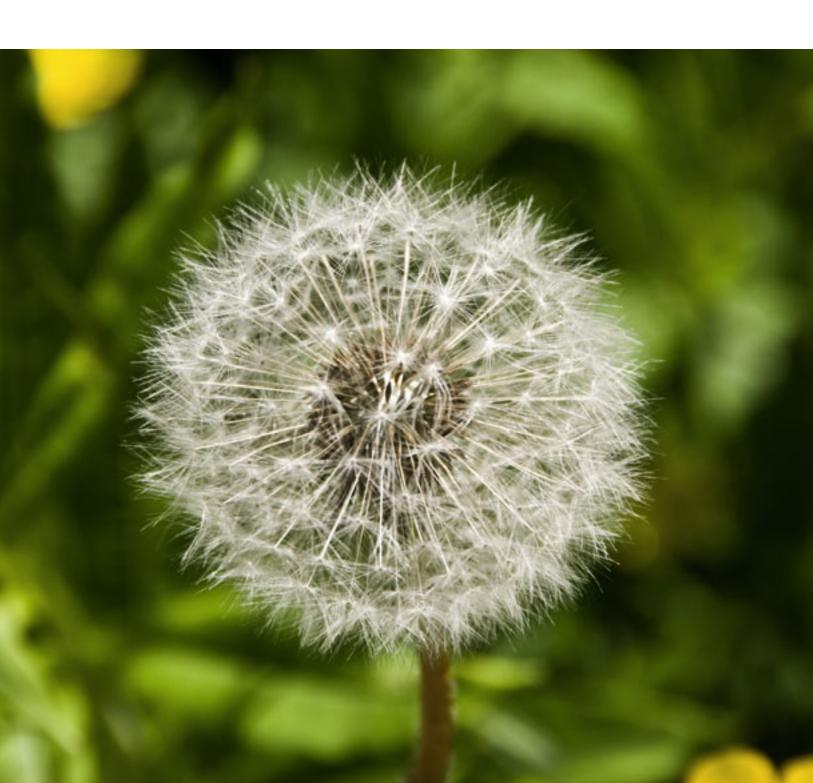
Vi vil stille deg noen spørsmål som handler om dine søvnvaner. 11.01 Har du hatt skiftarbeid de tre siste månedene? ☐ Ja Nei 11.02 Antall dager i løpet av uken hvor du ikke kan velge fritt når du vil sove (f.eks arbeidsdager)? Da går jeg til sengs klokken Jeg gjør meg klar til å sove klokken..... Antall minutter jeg trenger på å sovne Jeg våkner klokken Ved hjelp av: ☐ Vekkeklokke ☐ annen ytre påvirkning (støy, familie etc) ☐ av meg selv Antall minutter jeg trenger på å stå opp..... 11.03 Antall dager i løpet av uken hvor du <u>fritt</u> kan velge når du vil sove (f.eks helger eller fridager) Da går jeg til sengs klokken Jeg gjør meg klar til å sove klokken Antall minutter jeg trenger på å sovne Jeg våkner klokken Ved hjelp av: ☐ Vekkeklokke ☐ annen ytre påvirkning (støy, familie etc) ☐ av meg selv

Antall minutter jeg trenger på å stå opp.....

12. HUD OG HUDSYKDOMMER

	Hvor ofte dusjer eller bader du vanligvis? (sett ett kryss)	12.05	Har du ofte eller bestandig noen av følgende plager? (sett ett kryss for hy	ver linie)
	☐ 2 eller flere ganger daglig		8 p	Ja Nei
	☐ 1 gang daglig		Hevelse i ankler og legger, særlig om kvelden	
	4-6 ganger per uke		Åreknuter	
	2-3 ganger per uke		Eksem (rødt, kløende utslett) på	
	☐ 1 gang per uke		leggene	🔲 🔲
	sjeldnere enn 1 gang per uke		Smerter i beina når du går, men som forsvinner når du står stille	🔲 🔲
12.02	Hvor ofte vasker du vanligvis hendene med		The last section of the following by the section of	
	såpe i løpet av <u>en dag</u> ? (sett ett kryss)	12.06	Har du noen gang fått følgende diag av lege? (sett ett kryss for hver linje)	noser
	☐ 0 ganger		<u></u>	Ja Nei
	☐ 1-5 ganger		Psoriasis	
	6-10 ganger		Atopisk eksem	
	☐ 11-20 ganger		Rosacea	📙 📙
	Mer enn 20 ganger	12 በ7	Har du tilbakevendende store kviser	/
	Har du noen gang fått antibiotikakur (penicillin og liknende medisin) på grunn av en hudlidelse, for eksempel betent eksem, kviser, leggsår som ikke vil gro,	12.07	verkebyller som er ømme/smertefull og som ofte tilheler med arr på følge steder? (sett ett kryss for hver linje)	le
	tilbakevendende verkebyll?		Armhulene	📙 📙
	☐ Ja ☐ Nei		Under brystene	
	Hvis JA, hvor mange ganger i gjennomsnitt		Magefolden/navlen	🔲 🔲
	per år fikk du antibiotika i den perioden du		Rundt kjønnsorganet	🔲 🔲
	var mest plaget (sett ett kryss)		Rundt endetarmsåpningen	🔲 🔲
	☐ 1-2 ☐ 3-4 ☐ Mer enn 4 ganger		Lyskene	
12.04	Har du eller har du noen gang hatt følgende hudlidelser? (sett ett kryss for hver linje) Ja Nei Psoriasis		Hvis JA, har du noen gang oppsøkt le grunn av verkebyller? ☐ Ja ☐ Nei	ge på
	Atopisk eksem (barneeksem)		Illuia IA filludu da maan ay falaaada	
	Tilbakevendende håndeksem		Hvis JA, fikk du da noen av følgende behandlinger? (sett ett kryss for hver	linje)
	Tilbakevendende kviser over flere		3 . ,	Ja Nei
	måneder		Antibiotika salve/krem	🗆 🗆
	Legg- eller fotsår som ikke ville gro i		Antibiotika tabletter	🔲 🔲
	løpet av 3-4 uker		Kirurgisk åpning/tømming	🔲 🔲
	Hvis JA på spørsmål om legg-og/eller fotsår,		Større kirurgisk inngrep med	
	har du leggsår i dag?		fjerning av hud	
	☐ Ja ☐ Nei		Kirurgisk laserbehandling	. 🔲 🔲

Oppfølgingsspørsmål



INFORMASJON TIL OPPFØLGINGSSPØRSMÅL

De neste sidene med spørsmål skal ikke besvares av alle. Dersom du har svart ja på ett eller flere av spørsmålene under, ber vi deg om å gå videre til oppfølgingsspørsmål om emnet eller emnene du har svart ja på. De fire første emnene er fra det første spørreskjemaet og det siste spørsmålet er fra dette skjemaet.

Vi har for enkelhetsskyld markert emnene med ulike farger slik at du lett skal finne frem til de spørsmålene som gjelder for deg.

Dersom du svarte JA på at du har: <u>langvarige eller stadig tilbakevendende smerter som har vart i 3 måneder eller mer</u>, ber vi deg svare på spørsmålene på side 19 og 20. Margen er markert med grønn.

Dersom du svarte JA på at du har gjennomgått noen form for <u>operasjon i løpet av de siste 3 årene,</u> ber vi deg svare på spørsmålene på side 21 og 22. Margen er markert med lilla.

Dersom du svarte JA på at du <u>arbeider utendørs minst 25% av tiden</u>, eller i lokaler med lav temperatur, som for eksempel lager/industrihaller, ber vi deg svare på spørsmålene på side 23. Margen er markert med rød.

Dersom du svarte JA på at du har brukt <u>reseptfrie smertestillende medisiner</u>, ber vi deg svare på spørsmålene på side 24. Margen er markert med orange.

Dersom du svarte JA på at du har eller noen gang har hatt <u>plager med hud</u> (som psoriasis, atopisk eksem, legg- eller fotsår som ikke vil gro, tilbakevendende håndeksem, kviser eller verkebyll), ber vi deg svare på spørsmålene på side 25. Margen er markert med gul.

Har du svart <u>NEI</u> på disse fem spørsmålene, er du ferdig med besvarelsen din. Spørreskjemaet returneres i svarkonvolutten du fikk utlevert på undersøkelsen. Portoen er allerede betalt.

Skulle du ønske å gi oss en skriftlig tilbakemelding om enten spørreskjema eller Tromsøundersøkelsen generelt, er du hjertelig velkommen til det på side 26.

Har du noen spørsmål, kan du ta kontakt med oss på telefon eller på e-post. Du finner kontaktinformasjon på baksiden av skjemaet. **TUSEN TAKK** for at du tok deg tid til undersøkelsen og til å svare på spørsmålene fra oss.

13. OPPFØLGINGSSPØRSMÅL OM SMERTE

Du svarte i det første spørreskjemaet at du har langvarige eller stadig tilbakevendende smerter som har vart i <u>3 måneder eller mer</u>. Her ber vi deg beskrive de smertene litt nærmere.

13.01	Hvor lenge har du hatt disse smertene? Antall år måneder disse smertene?		
13.02	Hvor ofte har du vanligvis disse smertene? Hver dag En eller flere ganger i uken	☐ En eller flere ganger i må ☐ Sjeldnere enn 1 gang i m	
13.03	Hvor er det vondt? (Kryss av for alle steder der du smerter) Hode/ansikt Kjeve/kjeveledd Nakke Rygg Skulder Arm/albue Hånd Hofte	nar langvarige eller stadig tilb Lår/kne/legg Ankel/fot Bryst Mage Underliv/kjønnsorganer Hud Annet sted	akevendende
13.04	Hva mener du er årsaken til smertene? (Kryss av f Ulykke/akutt skade Langvarig belastning Kirurgisk inngrep/operasjon Skiveutglidning (prolaps)/lumbago Nakkesleng (whiplash) Migrene/hodepine Slitasjegikt (artrose) Leddgikt Bechterews sykdom Beskriv annen årsak:	r <u>alle</u> kjente årsaker) Fibromyalgi Angina pectoris (hjertekrand) Dårlig blodsirkulasjon Kreft Nerveskade/nevropati Infeksjon Helvetesild Annen årsak (beskriv under	
13.05	Hvilke former for behandling har du fått for smertsmertebehandling du har mottatt) Ingen behandling Smertestillende medisiner Fysioterapi/kiropraktikk Behandling ved smerteklinikk Operasjon	ene? (Kryss av for <u>alle</u> typer Smerteskole/avspenning. Akupunktur Alternativ behandling (ho aromaterapi, m.m.) Annen behandling	

13.06	På en skala fra 0 til 10, der 0 tilsvar du kan forestille deg:	er ingen	sme	erte	og '	10 t	ilsv	arer	der	ı ve	rst t	enke	elig	ze smerten
	Hvor sterke vil du si at smertene vanligvis er?	Ingen smerte	0	1	2	3	4	5	6	7	8	9	10	Verst tenkelige smerte
	Hvor sterke er smertene når de er på sitt sterkeste?	Ingen smerte	0	1	2	3	4	5 	6	7	8	9	10	Verst tenkelige smerte
	I hvor stor grad påvirker smertene søvnen din?	Påvirker ikke	0	1	2	3	4	5	6	7	8	9	10	Umulig å få sove
	I hvor stor grad hindrer smertene deg i å utføre vanlige aktiviteter hjemme og i arbeid?	Påvirker ikke	0	1	2	3	4	5	6 _	7	8	9	10	Kan ikke gjøre noe

14. OPPFØLGINGSSPØRSMÅL OM OPERASJON

I det første spørreskjemaet svarte du at du har gjennomgått en operasjon i løpet av <u>de siste 3</u> <u>årene</u>.

14.01 Hvor mange operasjoner har du Antall	0,	
Nedenfor ber vi deg beskrive ope av de siste 3 årene gjelder disse s	erasjonen. De pørsmålene	ersom du har gjennomgått flere operasjoner i løpet den siste operasjonen du gjennomgikk.
14.02 Hvor i kroppen ble du operert? (I samtidig ble operert flere steder i settes flere kryss) Operasjon i hode/nakke/rygg · Hode/ansikt	kroppen,	14.03 Bakgrunn for operasjonen: Akutt sykdom/skade
Rygg Operasjon i brystregionen Hjerte Lunger Bryster Annen operasjon i brystregionen Operasjon i mage/underliv Mage/tarm Lyskebrokk Urinveier/kjønnsorganer		Sykehuset i Tromsø
· Galleblære/galleveier · Annen operasjon i mage/ underliv	_	14.07 Er du overfølsom for berøring, varme eller kulde i et område nær operasjonsarret? ☐ Ja ☐ Nei
Operasjon i hofte/ben · Hofte/lår · Kne/legg · Ankel/fot · Amputasjon Operasjon i skulder og arm · Skulder/overarm · Albue/underarm · Hånd · Amputasjon		 14.08 Kan lett berøring av klær, dusj og lignende fremkalle ubehag/smerte? Ja Nei 14.09 Hvis du hadde smerter på operasjonsstedet før du ble operert, har du samme type smerte nå? Ja Nei 14.09 Hvis du hadde smerter på operasjonsstedet før du ble operert, har du samme type smerte nå? Ja Nei

14.10	Smerte fra operasjonsstedet: Svar på tenkelige smerte	en skala	a fra	0 ti	l 10), hv	or ()=in	gen	ı sm	erte	og	
	Hvor sterke smerter hadde du fra operasjonsstedet <u>før</u> operasjonen	Ingen smerte	0	1	2	3	4	5	6	7	8	9	Verst tenkelige smerte
	Hvor sterke smerter har du vanligvis fra operasjonsstedet <u>nå</u>	Ingen smerte	0	1	2	3	4	5	6	7	8	9	Verst tenkelige smerte
	Hvor sterke smerter har du nå fra operasjonsstedet når smertene er på det sterkeste	Ingen smerte	0	1	2	3	4	5	6	7	8	9	Verst tenkelige smerte

15. OPPFØLGINGSSPØRSMÅL OM ARBEID I KALDT KLIMA

I det første spørreskjemaet svarte du ja på at du arbeidet i kaldt klima. Her er noen oppfølgingsspørsmål vi håper du vil svare på.

15.01		15.05	5 Har du opplevd kløe og/eller utslett i forbindelse med kulde?	
	☐ Ja, ofte☐ Ja, noen ganger		☐ Ja ☐ Nei	
	☐ Nei, aldri	15 0.4	6 Har du i løpet av de <u>siste 12 måneder</u> vært	
15.02	Hvor lenge har du vært utsatt for kalde omgivelser under 0°C sist vinter?	13.00	involvert i ulykke som krevde medisinsk behandling der kulde var en viktig faktor? Ja Nei	i
	Fritid/hobby (timer/uke)		På arbeid	
	Arbeid (timer/uke)			
	Utendørs, godt kledd (timer/uke)	15.07	 Opplever du noen av følgende symptomer mens du oppholder deg i kalde omgivelser? I så fall, ved hvilken temperatur oppstår 	
	Utendørs, tynnkledd (timer/uke)		symptomene? Ja Nei Under °C	
	Innendørs, uten oppvarming (timer/uke)		Ja Nei Offder C	7
	I kalde omgivelser, med våte klær		Pusteproblemer	_
	(timer/uke)		Pipende pust	
	Kontakt med kalde gjenstander/ verktøy (timer/uke)		Slim fra lungene]
15.02	Hvilken omgivelsestemperatur		Brystsmerter	
10.00	forhindrer deg i å: Under °C		Forstyrrelse i hjerterytmen	
	Arbeide utendørs		Nedsatt blodsirkulasjon i hender/føtter]
	Trene utendørs		Synsforstyrrelse (kortvarig/forbigående)	
	Utføre andre aktiviteter utendørs		Migrene (kortvarig/forbigående)	
15.04	Har du hatt forfrysninger siste 12 måneder, med blemmer, sår eller skader i huden? ☐ Ja ☐ Nei		Hvite fingre (kortvarig/forbigående)]
	☐ Ja ☐ Nei Hvis JA, hvor mange ganger?		Blå, blå-røde fingre (kortvarig/forbigående)	
15.08	Hvordan påvirker kalde omgivelser og kulde	relate	erte symptomer din yteevne? Nedsatt Uforandret Forbedret	t
	Konsentrasjon. Hukommelse. Fingerfølsomhet (følelse) Fingerferdighet (motorikk) Kontroll av bevegelse (for eksempel skjelving) Tungt fysisk arbeid Langvarig fysisk arbeid			

23

16. BRUK AV RESEPTFRIE SMERTESTILLENDE LEGEMIDLER

I det første spørreskjemaet svarte du at du hadde brukt reseptfrie smertestillende legemidler de siste 4 ukene. Her er noen oppfølgingsspørsmål vi håper du vil svare på.

16.01	Hvilke typer reseptfrie smertestillende legemidler har du brukt?	Fenazon med koffein: (Antineuralgica ,Fanalgin Fenazon-koffein, Fenazon-koffein sterke)
	Paracetamol: (Pamol, Panodil, Paracet,	☐ Ikke brukt
	Paracetamol, Pinex)	☐ Sjeldnere enn hver uke
	☐ Ikke brukt	☐ Hver uke, men ikke daglig
	☐ Sjeldnere enn hver uke	☐ Daglig
	Hver uke, men ikke daglig	Hvor mye tar du vanligvis daglig når du bruker midlene?
	Daglig	(Antall tabletter)
	Hvor mye tar du vanligvis daglig	
		Mot hvilke plager bruker du reseptfrie smertestillende midler: (Flere kryss er mulig)
		☐ Hodepine
	Acetylsalisylsyre: (Aspirin, Dispril, Globoid)	☐ Menssmerter
	☐ Ikke brukt	☐ Migrene
	☐ Sjeldnere enn hver uke	Ryggsmerter
	Hver uke, men ikke daglig	☐ Muskelsmerter/leddsmerter
	☐ Daglig	☐ Tannsmerter
	Hvor mye tar du vanligvis daglig når du bruker midlene? (Antall tabletter)	☐ Annet
	Ibuprofen: (Ibumetin, Ibuprofen, Ibuprox, Ibux) Ikke brukt	Mener du å ha opplevd bivirkninger av noen av legemidlene? (sett ett kryss for hver linje) Ja Nei
		Paracetamol
	☐ Sjeldnere enn hver uke	Acetylsalisylsyre
	☐ Hver uke, men ikke daglig	Ibuprofen
	☐ Daglig Hvor mye tar du vanligvis daglig	Naproksen
	når du bruker midlene? (Antall tabletter, stikkpiller)	Fenazon med koffein
	16.	4 Hvor pleier du å kjøpe slike legemidler?
	Naproksen: (Ledox, Naproxen)	Hvor pleier du å kjøpe slike legemidler?☐ Apotek
	Naproksen: (Ledox, Naproxen)	Apotek
	Naproksen: (Ledox, Naproxen) Ikke brukt	☐ Apotek ☐ Dagligvare
	Naproksen: (Ledox, Naproxen) Ikke brukt Sjeldnere enn hver uke	☐ Apotek ☐ Dagligvare ☐ Bensinstasjon
	Naproksen: (Ledox, Naproxen) Ikke brukt Sjeldnere enn hver uke Hver uke, men ikke daglig	□ Apotek□ Dagligvare□ Bensinstasjon□ Utenlands

17. OPPFØLGINGSSPØRSMÅL OM HUDSYKDOMMER

På side 15 i dette spørreskjemaet svarte du at du har eller har hatt en hudsykdom. Her er noen oppfølgingsspørsmål vi håper du vil svare på.

Svar på en skala fra 0 til 10, der 0 tilsvarer ingen plager og 10 tilsvarer verst tenkelige plager. Dersom du svarte JA på at du har eller har hatt:

	9. k 9					Verst
17.01	Psoriasis · Hvor mye plaget er du av din psoriasis i dag?	Ingen plager	0 1 2 3	4 5 6	7 8 9 10	tenkelige plager
	· Hvor mye plaget er du av din psoriasis når den er som verst?					
17.02	Atopisk eksem					
	 Hvor mye plaget er du av ditt atopiske eksem i dag? Hvor mye plaget er du av ditt 					
	atopiske eksem når det er som verst?					
17.03	Håndeksem					
	· Hvor mye plaget er du av ditt håndeksem i dag?					
	 Hvor mye plaget er du av ditt håndeksem når det er som verst? 					
17 N.	Kviser					
17104	· Hvor mye plaget er du av dine kviser i dag?					
	· Hvor mye plaget er du av dine kviser når de er som verst?					
17.05	Verkebyller					
	· Hvor mye plaget er du av dine verkebyller i dag?					
	· Hvor mye plaget er du av dine verkebyller når de er som verst?					
17.06	Her er en liste over faktorer som kan te å utløse eller forverre verkebyller, kryss		17.08 Hvor gamn første gang		da du fikk vei	kebyller
	for hva du synes gjelder for deg:	a Nei	☐ 0-12 år		🔲 26-35 åı	•
	Stress/psykisk påkjenning	7	∐ 13-19 å		∐ 36-50 åı	
	Trange/tette klær	iП	☐ 20-25 å	år	☐ Over 50	år
	Menstruasjonssyklus		17.09 Dersom du	ı ikke leng	er har verkeh	ıller hvor
	Svangerskap				agene forsvan	
	Annet		🔲 0-12 år		26-35 åı	
17 07	Hvor mange utbrudd av verkebyller har o	du	☐ 13-19 å		∐ 36-50 åı	
1/.U/	vanligvis i løpet av ett år? (sett ett kryss)		□ 20-25 8	ar	☐ Over 50	ar
	□ 0-1 □ 4-6					
_	☐ 2-3 ☐ Mer enn 6	25				4

TILBAKEMELDING

Skulle du ønske å gi oss en skriftlig tilbakemelding om enten spørreskjema eller Tromsøundersøkelsen generelt, er du hjertelig velkommen til det her:					

Takk for hjelpen!





Tromsøundersøkelsen

Tromsøundersøkelsen Institutt for samfunnsmedisin, Universitetet i Tromsø 9037 TROMSØ

telefon: 77 64 48 16 **telefaks:** 77 64 48 31

epost: tromsous@ism.uit.no www.tromso6.no



Appendix C

Interview-questions related to the Tromsø Staph and Skin Study in the 6^{th} Tromsø Study

English translation

Interview-questions related to The Tromsø Staph and Skin Study

Interview Phase 1

ANTIBIOTICS:

Have you taken any antibiotics (tablets, injections or oral suspensions, nasal ointments, eye drops or eye ointment) the last 24 hours? Yes / No

If you have taken any antibiotics the last 24 hours:

- 1. What brand (included strength) did you take?
- 2. Administration form?
- 3. Numbers of tablets, milliliter of suspension etc. last time?
- 4. Total number of tablets, doses of suspensions etc during the last 24 hours?
- 5. How many hours since you took the last dose?

LIGHT/SUN EXPOSURE:

Have you used a solarium or any form of light therapy during the last 7 days? Yes / No

Have you been travelling to the south (for instance South-Europe) during the last 4 weeks? Yes / No

INFECTIONS AND EXPOSURE TO HEALTHCARE SERVICES:

Have you been suffering from persistent coughing during the last 24 hours? Yes / No

Have you been suffering from nasal discharge during the last 24 hours? Yes / No

Have you been hospitalized during the last 12 months? Yes / No

Does anyone in your household work in health care services (hospital, nursing home, senior care service, GP's office, public health center)? Yes / No

Have you ever had a tonsillectomy? Yes / No / Don't know

Interview Phase 2

ANTIBIOTICS:

Have you taken any antibiotics (tablets, injections or oral suspensions, nasal ointments, eye drops or eye ointment) the last 24 hours? Yes / No

If you have taken any antibiotics the last 24 hours:

- 1. What brand (included strength) did you take?
- 2. Administration form?
- 3. Numbers of tablets, milliliter of suspension etc. last time?
- 4. Total number of tablets, doses of suspensions etc during the last 24 hours?
- 5. How many hours since you took the last dose?

LIGHT/SUN EXPOSURE:

Have you used a solarium or any form of light therapy during the last 7 days? Yes / No

Have you been travelling to the south (for instance South-Europe) during the last 4 weeks? Yes / No

INFECTIONS AND EXPOSURE TO HEALTHCARE SERVICES:

Do you work in health care services (hospital, nursing home, senior care service, GP's office, public health center)? Yes / No

Have you been suffering from persistent coughing during the last 24 hours? Yes / No

Have you been suffering from nasal discharge during the last 24 hours? Yes / No

Have you been hospitalized since last attendance (survey)? Yes / No

Appendix D

Interview-questions related to the Tromsø Staph and Skin Study in the 6^{th} Tromsø Study

Original questionnaire

Intervjuspørsmål til Tromsø Staph and Skin Study

Intervju Fase 1:

ANTIBIOTIKA:

Har du brukt antibiotika enten i form av tabletter/mikstur, injeksjoner, nesesalve, hudsalve, øyedråper eller øyesalve i løpet av de siste 24 timene? Ja / Nei

Hvis du har tatt antibiotika siste 24 timer:

- 1. Hvilken type (inkludert styrke) har du brukt?
- 2. Hvilken inntaksmåte?
- 3. Hvor mange tabletter, milliliter suspensjon etc. tok du ved siste dose?
- 4. Hvilken totaldose av tabletter, suspensjon etc. har du tatt siste 24 timer?
- 5. Hvor mange timer er det siden du tok siste dose?

SOLARIUM / SOLEKSPONERING:

Har du tatt solarium eller lysbehandling de siste 7 dagene? Ja / Nei

Har du vært på reise til sydligere breddegrader (tilsvarende Sør-Europa) i løpet av de siste 4 ukene? Ja / Nei

INFEKSJONER / EKSPONERING FRA HELSEVESENET:

Har du vært forkjøla i halsen i løpet av de siste 24 timene? Ja / Nei

Har du vært forkjøla i nesen i løpet av de siste 24 timene? Ja / Nei

Har du vært innlagt på sjukehus i løpet av de siste 12 månedene? Ja / Nei

Arbeider noen i din husstand i helsevesenet (sjukehus, sjukehjem, hjemmetjenesten, legekontor, helsestasjon)? Ja / Nei

Har du tidligere fått fjernet mandlene? Ja / Nei / Vet ikke

Intervju Fase 2:

ANTIBIOTIKA:

Har du brukt antibiotika enten i form av tabletter/mikstur, injeksjoner, nesesalve, hudsalve, øyedråper eller øyesalve i løpet av de siste 24 timene? Ja / Nei

Hvis du har tatt antibiotika siste 24 timer:

- 1. Hvilken type (inkludert styrke) har du brukt?
- 2. Hvilken inntaksmåte?
- 3. Hvor mange tabletter, milliliter suspensjon etc. tok du ved siste dose?
- 4. Hvilken totaldose av tabletter, suspensjon etc. har du tatt siste 24 timer?
- 5. Hvor mange timer er det siden du tok siste dose?

SOLARIUM/ SOLEKSPONERING:

Har du tatt solarium eller lysbehandling de siste 7 dagene? Ja / Nei

Har du vært på reise til sydligere breddegrader (tilsvarende Sør-Europa) i løpet av de siste 4 ukene? Ja / Nei

INFEKSJONER/ EKSPONERING FRA HELSEVESENET:

Arbeider du i helsevesenet (sjukehus, sjukehjem, hjemmetjenesten, legekontor, helsestasjon)? Ja / Nei

Har du vært forkjøla i halsen i løpet av de siste 24 timene? Ja / Nei

Har du vært forkjøla i nesen i løpet av de siste 24 timene? Ja / Nei

Har du vært innlagt på sjukehus siden siste fremmøte/undersøkelse? Ja / Nei



