

Humoral and cellular immune responses in mice after airway administration of *Bacillus thuringiensis* Cry1Ab and MON810 *cry1Ab*-transgenic maize

Monica Andreassen ^{a,b,*}, Elena Rocca ^a, Thomas Bøhn ^{a,d}, Odd-Gunnar Wikmark ^a, Johnnie van den Berg ^c, Martinus Løvik ^e, Terje Traavik ^{a,d} and Unni Cecilie Nygaard ^b

^a GenØk - Centre for biosafety, Tromsø, Norway

^b Norwegian Institute of Public Health, Oslo, Norway

^c North-West University, Potchefstroom, South-Africa

^d UiT The Arctic University of Norway, Tromsø, Norway

^e Norwegian University of Science and Technology, Trondheim, Norway

*Corresponding author

Monica Andreassen	NIPH, MIVM, PO Box 4404, 0403 Oslo, Norway monica.andreassen@fhi.no , +47 21076257
Elena Rocca	Gneisveien 34, 1430 Ås, Norway elenarocca76@gmail.com , +47 90960481
Thomas Bøhn	GenØk – Centre for biosafety, PO Box 6418, 9294 Tromsø, Norway, thomas.bohn@uit.no , +47 77 64 45 41
Odd-Gunnar Wikmark	GenØk – Centre for biosafety, PO Box 6418, 9294 Tromsø, Norway, odd-gunnar.wikmark@uit.no , +47 77646634
Johnnie van den Berg	NWU, Private Bag X 6001, Potchefstroom, 2520, South Africa, Johnnie.VanDenBerg@nwu.ac.za , +27 18299 2376
Martinus Løvik	Nordengveien 85D, 0755 Oslo, martloev@online.no , + 47 71184711
Terje Traavik	GenØk – Centre for biosafety, PO Box 6418, 9294 Tromsø, Norway, terje.traavik@uit.no , +47 77644379
Unni C. Nygaard	NIPH, MIVM, PO Box 4404, 0403 Oslo, Norway unni.cecilie.nygaard@fhi.no , +47 21076435

ABSTRACT

Genetically modified (GM) crops may bring new proteins with immunogenic and allergenic properties into the food and feed chains. The most commonly grown GM maize, MON810 expresses a modified version of the insecticidal Cry1Ab protein originating in the soil bacterium *Bacillus thuringiensis* (Bt). Immune reactions following inhalation of pollen and debris from such plants have been scarcely studied. We exposed BALB/c mice to purified Cry1Ab proteins and Cry1Ab-containing MON810 plant materials by intranasal installation. No anti-Cry1Ab antibodies were detected following exposure to the plant materials. Exposure to purified Cry1Ab resulted in specific anti-Cry1Ab IgG1 and IgE production, indicating inherent immunogenicity and allergenicity. Mice exposed to leaf extracts from both MON810 and unmodified maize demonstrated influx of lymphocytes and eosinophils in the bronchoalveolar lavage, and increased cytokine release in mediastinal lymph node cells. The results indicate that the airway exposure to Cry1Ab proteins may be a route of practical relevance.

Key words: *Bacillus thuringiensis*; MON810; Cry1Ab; IgE; airway allergy; mouse models

1. INTRODUCTION

Genetically modified (GM) crops are created by direct transfer, insertion and ligation into the plant genome of a transgene, coding for a protein that may confer an advantageous trait, e.g. pest insect resistance, to the recipient plant (reviews in Traavik and Lim Li, 2007). As many GM crops bring new proteins into the food and feed chains, concerns have been raised about potential adverse effects in terms of toxicity (Hammond et al., 2013, Domingo and Gine Bordonaba, 2011), impaired nutrition value (EFSA, 2008) and undesirable immune responses (Finamore et al., 2008, Kroghsbo et al., 2008, Adel-Patient et al., 2011, EFSA, 2010, Taylor and Hefle, 2001, Bernstein et al., 2003). In predisposed individuals, immunogenic proteins (i.e. proteins that are able to induce specific humoral and/or cellular immune responses) may be capable of stimulating a type-I hypersensitivity reaction through specific immunoglobulin (Ig) E responses, resulting in allergy against the protein (Poulsen and Hummelshoj, 2007).

The assessment of potential immunogenicity and/or allergenicity of novel proteins expressed in a GM plant is a challenging task, as no single validated test or type of information exists to predict allergic or other immune responses against them (Kimber and Dearman, 2002, EFSA, 2010). For ethical reasons, experimental sensitisation to potential allergens cannot be performed in humans, while epidemiological investigations have not been published so far. Current allergenicity assessment strategies, such as the pepsin resistance test and *in vitro* digestibility tests, the search for sequence and structural similarities between newly expressed proteins and known allergens, as well as *in vitro* IgE binding tests, have severe limitations (Bernstein et al., 2003, Guimaraes et al., 2010). Most importantly, *in vitro* testing cannot completely mimic the complex interaction between the many factors that are involved in the induction of an immune response. The EU scientific guidance document on the assessment of

allergenicity of GM plants (EFSA, 2010) recognises the weight of evidence approach and case by case evaluation to be the most appropriate way of assessing the allergenicity of GM food and feed. Animal models, in particular, are mentioned as helpful to evaluate the potential of a newly expressed protein to sensitise and/or elicit allergic reactions, and to evaluate the immunogenic and/or adjuvant capacity of these proteins.

The most commonly grown GM maize event, MON810, expresses an insecticidal Cry1Ab protein from a genome-inserted *cry1Ab* transgene originating in the soil bacterium *Bacillus thuringiensis* (Bt). The Cry1Ab protein is currently not considered an allergen in part based on the observation that specific IgE responses against Cry1Ab were not detected in exposed food-allergic humans (Nakajima et al., 2007). Moreover, it has been stated for Cry1Ab protoxin that there is no sequence similarity to known allergens (Randhawa et al., 2011), and the protein is rapidly digested in simulated gastric and intestinal fluids (EFSA, 2009). However, sequence similarity, protein source and digestibility alone cannot predict allergenicity. Several studies in mammals and fish have reported effects on the immune system after feeding of MON810-containing diets, including elevated levels of allergic and inflammatory associated cytokines in serum and altered percentage of B and T cells in the gut (Finamore et al., 2008) and a changed proportion of ileal B cells and macrophages (Walsh et al., 2011). Assessments of potential Cry1Ab immunogenicity and/or allergenicity have mostly been performed with purified protein isolated from the naturally occurring *Bt* spore Cry(stal) inclusions, or with a version produced in *E. coli* that is trypsinised to mimic the activated toxin expressed in GM plants. The many important structural and contextual differences between bacterial and plant Cry1Ab may, however, make extrapolations inadequate, and may have led to irrelevant conclusions. In the Bt bacterium, Cry1Ab is expressed as a protoxin that must be activated by enzymatic cleavage in the lepidopteran gut to obtain its toxicity. The

MON810 transgene, on the other hand, encodes only the N-terminal half of the protoxin. Hence it expresses an already activated version of the Cry1Ab toxin, which is probably structurally and immunogenically different from the Bt protoxin (Guerrero et al., 2004, Guerrero et al., 2007, Kitami et al., 2011). Furthermore, transgenic insertions into a maize genome may cause a complex recombination event resulting in expression of “new” unanticipated proteins (Rosati et al., 2008). The different origins of the Cry1Ab protein (bacterium- or plant-expressed), as well as the specific version (protoxin, toxin, size, molecular conformation etc.) or context (purified bacterial, part of plant cell or tissue), thus prerequisite careful consideration and interpretation.

Although the mucous membranes of the respiratory and gastro-intestinal tracts may be the most important portals of exposure (Vazquez-Padron et al., 2000, Kitami et al., 2011) little information exist on the potential immune effects after airway exposure to the Cry1Ab proteins. Inhalation of pollen and plant debris may be a realistic exposure route for humans and domestic as well as wildlife animals in the fields, and also of workers during processing of food and feed. To our knowledge, airway exposure to adequate plant Cry1Ab proteins has not yet been investigated. Purified Cry1A protoxins and trypsin activated toxins have demonstrated specific humoral as well as cellular anti Cry1A immune responses after intranasal administration in mice (Guerrero et al., 2004, Guerrero et al., 2007). The same authors emphasized that the immune responses after intranasal exposure were unique, and suggested that binding of the lectin motifs of Cry proteins to the lectin binding receptors in nasal-associated lymphoid tissue (NALT) might contribute to that (Takata et al., 2000). The immunological potential of airway exposure to Cry1Ab-containing feed debris was directly illustrated, although unintended, when control fed and Cry1Ab-containing rice fed rats kept in the same room all developed Cry1Ab specific antibodies (Kroghsbo et al., 2008). In humans,

Cry1Ab specific antibodies have been detected after occupational inhalation of Bt-based insecticidal sprayings (Bernstein et al., 1999, Doekes et al., 2004). Studies like these indicate that specific immune responses may be induced after inhalation of Cry1Ab proteins.

Given the expansion of cultivation areas for *cry*-transgenic maize varieties, the current knowledge gaps concerning immunogenicity and allergenicity of Cry1Ab proteins in general, and inhalation of such proteins in particular, warrants further research. We hypothesized that airway exposure to Cry1Ab-expressing plant materials may induce immunological reactions. Since the origin/version/context of the Cry1Ab protein may give subtle structural modifications and thus affect the immune reactions, we investigated the various versions of the Cry1Ab protein to elicit immune and/or allergic responses in mice after intranasal exposure to 1) Cry1Ab protoxin, 2) trypsinised protoxin Cry1Ab (trypCry1Ab), 3) MON810 pollen and 4) MON810 leaf extract.

2. MATERIALS AND METHODS

2.1. *Animals*

Five to six-week old female BALB/c mice (Charles River, Sulzfeld, Germany) were allowed to rest for one week. The mice were randomly allocated to groups (8 –10 mice per group) and housed 3 – 5 animals together in cages with Nestpack bedding (Datesand Ltd, Manchester, UK). Pelleted Teklad Global 18% protein rodent diet 2018 (Harlan Laboratories, Madison, Wisconsin, USA) and tap water were given *ad libitum*. The mice were exposed to a 12/12 hour light/dark cycle, room temperature of 21 +/- 2 °C and 55 +/- 10 % humidity. The experiments were performed in conformity with laws and regulations for live animals, and approval was given by the North-West University Office of Ethics Committee (Ethical approval reference: NWU-00025-10-A3 Potchefstroom) and by the Norwegian Animal Research Authority under the Ministry of Agriculture (reference FOTS 4050) for experiment 1 and 2 (described below), respectively.

2.2. *Experiments*

Experiment 1 was conducted at North-West University (NWU) in Potchefstroom, South Africa, in 2011, while experiment 2 was performed at the Norwegian Institute of Public Health (NIPH), Oslo in 2012. Although the two experiments were not supposed to be replicates (different origin/versions of the Cry1Ab protein were tested), the experimental conditions were kept as similar as possible (otherwise stated) and performed by the same investigator (MA).

2.3. Experimental protocol

The mice were anaesthetised and exposed intranasally (i.n.) to 35 μ L of test solutions on days 0, 1, 2, and booster i.n. on days 21, 22 and 23, and the experiments were terminated on day 26 (Figure 1). A 100 μ l blood sample was collected from *vena saphena lateralis* prior to the exposures on days 0 and 21. The mice were exsanguinated by heart puncture under anaesthesia on day 26, and blood and 3 x 0.8 mL bronco-alveolar lavage fluid (BALF) were collected as previously described by Samuelson et al. (2009). BALF samples were kept on ice until processing. In experiment 2, MLNs were located according to Van den Broeck et al. (2006), excised, and single cell suspensions were prepared as described by Hansen et al. (2011).

2.4. Cry1Ab protein

Purified Cry1Ab protoxin isolated from *Bacillus thuringiensis* spores, prepared in PBS (pH 7.4) and lyophilised, was purchased from Abraxis via Analytical Solutions (Banbury, South Africa). Trypsinised Cry1Ab (trypCry1Ab) protein was purchased from Case Western Reserve University, (Ohio, USA, Dr. Marianne Carey). The origin of the *cry1Ab* gene inserted in *E. coli* was the *Bt kurstaki HD-1* strain. The inclusion bodies were solubilized at pH 10.5 in the presence of a reducing agent and the precipitated protoxins were digested by commercial bovine trypsin and subsequently purified by ion exchange HPLC. The relevant fractions were analysed by gel filtration, HPLC and SDS-PAGE, desalted and lyophilized (M. Carey, personal communication). The protein preparations were dissolved in a sterile physiological buffer (HBSS or PBS). The total doses of protoxin Cry1Ab and trypCry1Ab were 60 and 16.5 μ g per mouse, respectively.

2.5. Maize plant material

The maize event MON810 (cultivar DKC78-15B) and its non-GM conventional counterpart (cultivar CRN 3505) were grown under identical conditions in pots in a greenhouse at NWU. The presence of Cry1Ab in MON810 plants was confirmed with the QuickStix kit for Cry1Ab corn leaf & seed (Envirologix, Portland, Maine, USA). On days 0, 1, and 2 of the experiment, individual plant flowers from the two cultivars were carefully enclosed in plastic bags and shaken to dislodge pollen from flowers 2-3 hours before exposure to mice. After sieving twice to remove fragments other than pollen, 5 mg pollen was suspended in 350 μ l sterile HBSS less than 1 hour before exposure. Additional pollen was frozen immediately and kept at minus 80 °C until exposures on day 21, 22 and 23. Frozen pollen was thawed in room temperature 2-3 hours before exposure to mice and liquefied as described above.

For leaf extract preparation, whole leaves were collected from greenhouse-cultivated MON810 (cultivar 6Q-308Bt) and non-GM (cultivar 6Q-121) maize plants at NWU. The leaves were enclosed in wet paper, sent by airmail to Norway. At arrival, the leaves were cut into 1 - 2 cm long pieces before manual grinding using a mortar and pestle. Ten ml HBSS was added to 1 gram of ground leaves and left overnight at 4 °C. After centrifugation at 2000 rpm for 5 min, the supernatant was collected and frozen at -18 until exposure.

The content of plant expressed Cry1Ab in both pollen suspension and leaf extract were determined using a semi-quantitative ELISA kit (Agdia, Elkhart, Indiana, USA) following manufacturer instructions. Standard curves were obtained using progressive dilutions of a

purified trypCry1Ab solution, ranging from 0.03 – 0.96 ng/μl and 3 – 96 ng/μL for the quantification of Cry1Ab in pollen and leaves respectively.

2.6. Anaesthetics

At NWU, an initial dose of 1.4 mL of Halothane (SafeLine Pharmaceuticals (Pty) Ltd, South Africa) was added to an inhalation chamber, and the mice were kept individually in the chambers until sedated. A supplementary dose of 0.4 ml of Halothane was then added once the sedative effect declined. At NIPH, 3.5 % Isofluran gas (Isoba vet; Intervet/Schering-Plough Animal Health, Lysaker, Norway) was administered in surgical O₂ in an inhalation chamber until sedation.

2.7. Intranasal exposure

Each individual mouse was put on its back, as droplets of the test solution were applied with a pipette onto the left and right nostril repeatedly until the complete volume of 35 μL was taken up.

2.8. Detection of endotoxin in test solutions

Endotoxin levels were measured in the pollen suspension (0.131 EU/mL in both MON810 and non-GM), leaf extracts (0.130 and 0.129 EU/mL for MON810 and non-GM, respectively) and the trypCry1Ab solution (0.046 EU/mL) using ToxinSensor™ Chromogenic LAL endotoxin Assay kit (GenScript, Piscataway, New Jersey, USA) according to the manufacturers' description.

2.9. Detection of Cry1Ab-Specific Antibodies

In-house enzyme-linked immunosorbent assay (ELISA) protocols were established to detect specific anti-Cry1Ab IgG1, IgG2a and IgE in mouse sera. For the IgG1 assay, microtiter 96-well plates (Maxisorp, Nunc-Immuno Plate, Thermo Fisher Scientific, Roskilde, Denmark) were coated with 2ng/ μ L purified trypCry1Ab protein per well and incubated for 1 hour at room temperature (RT) and then overnight at 4 °C. Plates were washed with 50 mM Tris/HCl buffer pH8 with 0.05 % Tween 20 (Tris/Tween) and incubated with blocking solution (5 % skimmed milk powder (Fluka Analytical, Sigma-Aldrich Chemie GmbH, Steinheim, Germany) in phosphate buffered saline (PBS)) for 1 hour at room temperature (RT). After subsequent washing, diluted sera (1:50) were added and plates were incubated for 1 hour at RT and again overnight at 4 °C. The plates were washed, 200 ng biotinylated rat anti-mouse IgG1 (Experimental Immunology Unit, University of Louvain, Belgium) was added per well and plates were incubated for 1 hour at RT. After subsequent washing, poly-HRP-streptavidin (Thermo-Scientific, Pierce Biotechnology Inc., Rockford, IL, USA) diluted 1:40000 was added and incubated for 1 hour at RT. The plates were washed and colour development was obtained by adding stabilized chromogen TBM (Invitrogen). The reaction was stopped with 2N H₂SO₄ solution, after incubation in darkness for a maximum of 15 minutes.

Considering the small relative amounts of IgE compared to other classes of Igs, competitive displacement of IgE by the latter must be avoided. Therefore, we used anti-mouse IgE antibodies as capture antibody. Microtiter 96-well plates with 96 wells were coated with 200 ng rat anti mouse IgE antibodies (clone LO-ME-3, Experimental Immunology Unit, University of Louvain, Belgium,) per well, incubated for 1 hour at RT and then overnight at 4 °C. Subsequently, the plates were washed with Tris/Tween and blocked with 5% skimmed

milk in PBS for 1 hour at RT. After subsequent washing, diluted sera (1:10) were added and plates were incubated for 1 hour at RT and then again over night at 4 °C. The following day plates were washed, 300 ng trypCry1Ab were added to each well and incubated for 1 hour at RT. After subsequent washing, 300 ng biotinylated rabbit anti mouse Cry1Ab antibody (Abraxis, Warminster, Pennsylvania, USA) were added per well as detection antibody and plates were incubated for 1 hour at RT. Plates were washed and detection was performed with poly-HRP-streptavidin and stabilized chromogen TBM as described above. The reaction was stopped with 2N H₂SO₄ solution after incubation in darkness for a maximum of 15 minutes.

The relative amount of specific IgG2a is apparently small, and a Cry1Ab-specific IgG2a ELISA protocol was established using the same reasoning as for detection of Cry1Ab-specific IgE. Microtiter plates with 96 wells were coated with 200 ng biotinylated rat anti mouse IgG2a antibodies (clone R19-15, BD Biosciences Pharmingen, San Diego, California, USA) per well, and the protocol for Cry1Ab-specific IgE detection was subsequently followed as described above. To accelerate the reactivity of each step, incubations were performed at 37 °C for the detection of specific anti-Cry1Ab IgG2a.

Standard curves were made from duplicates of diluted serum pools from mice immunized with trypCry1Ab and Al(OH)₃, and included for all antibody assays on each plate. As the amount of specific IgG1, IgE and IgG2a in the standards is unknown, the levels are presented as arbitrary units (AU). Absorbance was measured at 450 nm on a BioTek Elx808 Absorbance Microplate reader with the Gen5™ Microplate Data Collection & Analysis Software (BioTek® Instruments, Inc., Winooski, Vermont, USA).

2.10. *Cytokine analyses*

Cytokine levels in BALF and in supernates from MLN cells were determined by Cytometric Bead Array (CBA) flex set kit from BD Biosciences (San Diego, California, USA). Beads dyed with fluorescence red and covalently coupled to antibodies that capture specific cytokines, allow simultaneous quantification of several cytokines. IL-5, IL-10, IFN γ , TNF α and MCP-1 were measured in BALF from both experiments. IL-10, IFN γ , IL-4, IL-13 and IL-17 were determined in the supernates from MLN cells stimulated with 172 μ g/mL trypCry1Ab and incubated at 37 °C for 120 hours, in experiment 2.

2.11. *Differential counts*

Cells (125 000 cells/mL) from all three BALF collections from each individual mouse were spun down on to cytoslides using a cytocentrifuge (Shandon Scientific Ltd., Astmoor, Runcorn, UK) at 70 g for 6 minutes, and stained with the Hemacolour rapid staining according to the manufacturer's protocol (Merc KGaA, Darmstadt, Germany). A total number of 200 cells were counted on each slide in a light microscope (Axioplan2, Zeiss, Göttingen, Germany) at 100x magnification, differentiating macrophages, eosinophils, neutrophils, lymphocytes and epithelial cells based on their morphological characteristics. Cell differential counts for all animals were performed blindly by the same investigator (MA).

2.12. *Statistical analyses*

Statistical analyses were performed using SigmaPlot 12.3 and Minitab 16 Statistical software. All normally distributed parameters with comparable variances were tested by one-way ANOVA. When necessary, data were log₁₀ transformed to obtain normal distribution and

comparable variances. In the case of significant overall differences, pair-wise comparisons between treatment and control or between pairs of treatments were performed with the Dunnett's *post hoc* test. Data that could not be transformed to normal distribution and comparable variances were tested with the non-parametric Kruskal-Wallis test, using Mann-Whitney *post hoc* tests for pair-wise comparisons between treatments versus control or between pairs of treatments. Differences between outcomes were considered significant when p-values were < 0.05 .

3. RESULTS

3.1. Cry1Ab exposure dose

Based on the measured concentrations (Table 1) the calculated total dose of Cry1Ab given per mice exposed to pollen or leaf extracts was 0.02 µg and 12.86 µg, respectively. Non-GM pollen and leaves had no detectable content of Cry1Ab protein.

3.2. Serum Cry1Ab-specific antibodies

Cry1Ab-specific antibodies were analysed in sera collected prior to the intranasal exposures on days 0 and 21, and at termination on day 26. Cry1Ab-specific antibodies were not detected on day 0. Cry1Ab-specific IgG1 was elevated on day 21 in mice exposed to both Cry1Ab protoxin and trypCry1Ab compared to controls (data not shown). On day 26, mice exposed to Cry1Ab protoxin, but not maize pollen, had significantly higher serum levels of both Cry1Ab-specific IgE and IgG1, and a tendency towards a higher serum level of Cry1Ab-specific IgG2a, compared to the control group (Figure 2a, b, c). Furthermore, mice exposed to trypCry1Ab protein, but not maize leaf extracts, had elevated serum level of Cry1Ab-specific IgG1 and Cry1Ab-specific IgE, on day 26 (figure 2 d, e, f). Cry1Ab-specific IgG2a levels were not determined on day 0 and 21 due to limited amounts of serum, and the levels on day 26 did not significantly differ in mice from either group.

3.3. Cytokine levels in BALF

IL-5, IL-17, MCP-1, TNFα and IFNγ cytokine levels determined in BALF collected on day 26 were very low (figure 3). Although statistically significant differences between the groups were observed, levels of cytokines in mice exposed to Cry1Ab protoxin or MON810 pollen

were not significantly different from those exposed to the non-GM pollen. Cytokine levels in BALF collected on day 26 after exposure of mice to trypCry1Ab and maize leaf extracts were below the detection limit irrespective of the exposure scheme (data not shown).

3.4. Cell differential counts

The percentage of neutrophils, macrophages, lymphocytes and eosinophils were determined in BALF from mice exposed to trypCry1Ab and maize leaf extracts. Mice exposed to MON810 leaf extract had a significantly higher percentage of lymphocytes and eosinophils, and significantly lower percentage of macrophages, than mice in the control group (Figure 4). However, these numbers were not significantly different from what was observed in mice exposed to the non-GM leaf extract. The percentage of neutrophils was not different between any groups of mice irrespective of exposure scheme.

3.5. Ex vivo cytokine secretion from MLN cells

Cytokines were measured in supernates from MLN cells collected on day 26 and stimulated with trypCry1Ab. IL-10 was significantly elevated in mice exposed to MON810 leaf extract compared to mice exposed to non-GM extract, and compared to control mice (Figure 5a). Furthermore, mice exposed to MON810 leaf extract had significantly higher levels of IL-4, IL-13, IFN γ , and IL-17 compared to mice in the PBS-control group, however the levels were not significantly different from mice exposed to non-GM leaf extract (figure 5).

4. DISCUSSION

The MON810 plant material did not elicit humoral immune responses in mice after airway exposure. However, the production of specific IgG1 against the two purified protein versions indicates that Cry1Ab protein as such is capable of provoking immunogenic responses, and the production of specific IgE further indicate the capability of Cry1Ab to induce pro-allergic responses, in mammals. In the field, the airway exposure of Cry1Ab proteins (e.g. through pollen and dust) is a highly relevant route of exposure and our results therefore warrants further studies. Our findings are in agreement with previous investigations showing immunogenic responses after Cry1Ab administration through other routes. For example, Cry1Ab-specific IgG, IgE and IgG2a antibody response were detected in BALB/c mice immunised intraperitoneally (i.p.) with the protoxin Cry1Ab, however this was not the case after intragastric (i.g.) administration (Adel-Patient et al., 2011). The authors did not, like us, detect specific antibody responses after immunisation with a protein extract from MON810 maize, neither via the i.p. nor the i.g. route. On the other hand, our results are only partly in line with the findings of Guerrero et al. (2004). While an i.p. immunisation with a total dose of 150 µg purified protoxin Cry1Ab (produced in recombinant *E.coli*; 133 kDa) resulted in a strong specific IgG antibody response in BALB/c mice, i.n. immunisations with the same dose gave no such response. Furthermore, 150 µg of toxin Cry1Ab (trypsinised protoxin; 70 kDa) induced a strong specific IgG response after both the i.p. and i.n. immunisation route.

The precise measurement of Cry1Ab concentrations in plant material has been proven to be a difficult task, with significant inter-laboratory differences (Szekacs et al., 2012) and huge variations in concentrations reported and methods used. Furthermore the expression levels of Cry1Ab as well as the whole proteome of a given GM plant may vary considerably when it

is grown under different environmental conditions (Agapito-Tenzen et al., 2013). The calculated Cry1Ab content in our plant material was based on values from an adjusted semi-quantitative assay and is thus not absolutely, but reflects a relative difference in the Cry1Ab content in different parts of the plant, suggesting that leaves contain approximately 90 times more Cry1Ab than pollen. Nevertheless, mice exposed to both plant materials received a considerably lower dose of Cry-proteins compared to those exposed to purified protein solutions, which could, at least in part, explain the lack of humoral responses against the plant Cry1Ab. On the other hand, immunisation with low doses of allergens has been shown to induce higher IgE production than high doses (Arps et al., 1998, Hansen et al., 2011), i.e. not supporting a simple positive dose-response relationship. This was also the case in a study by Adel-Patient et al. (2011), where 1 µg protoxin Cry1Ab induced a specific IgE response, while the dose of 100 µg did not. In addition, structural differences between the Cry1Ab proteins we used in our four treatments may be accountable for the difference in immune responses. The trypsin activation of the protein produced in *E. coli* is supposed to give a Cry1Ab that is similar to the activated version expressed in plants, although the degree of similarity has not been confirmed. Also post-translational differences could explain the difference in immune response between purified protein and plant material preparations, as suggested by Prescott et al. (2005). It was demonstrated in a mouse model that transgenic expression of the bean alpha-amylase inhibitor-1 (α AI) in peas led to the synthesis of a protein variant with altered immunogenicity. However, these findings could not be reproduced by Lee et al. (2013). Whether the dissimilar responses to plant and purified protein preparations are due to discrepancy in doses or structural differences can however not be answered by the present study.

To our knowledge, a Cry1Ab specific IgE response has not previously been demonstrated after airway exposure to Cry1Ab proteins. Since the amount of specific IgE antibodies in general are relatively low compared to other classes of immunoglobulins, and thus vulnerable to be outcompeted by immunoglobulins in abundance (Ladics et al., 2010), our sandwich ELISA design trapped Cry1Ab specific IgE on the well surface by coating microtiter plates with anti-IgE monoclonal antibodies, before incubating with trypCry1Ab protein and biotinylated anti-Cry1Ab as the detection antibody. The use of the trypsinised version of the Cry1Ab protein in the assay could theoretically result in lack of detection of the fraction of specific antibodies towards epitopes present on the protoxin or the plant version. However, previous studies suggest that most epitopes are on the N-terminal present on both protoxin and trypsinised versions of Cry1Ab (Guerrero et al., 2004), and our data demonstrate the presence of Cry-specific IgE responses in animals treated with both protein versions.

It should be noted that the clinical relevance of the detected specific IgE was not evaluated. The inbred BALB/c is an “allergy prone” mouse strain that easily adopt the allergy-associated Th2 response pathway. The strain has been employed in previous allergy models and usually requires an adjuvant to elicit an allergic response (Nygaard et al., 2005). Here, a clear specific IgE response against both protoxin and trypsinised Cry1Ab was elicited even without an adjuvant, suggesting that genetically predisposed (or atopic) subjects may develop Cry1Ab IgE antibody response after airway exposure. Furthermore it is important to recognise that the model is optimised to induce allergy as the protocol with three consecutive immunisations and thereafter three booster immunisations is more likely to provoke IgE responses.

Because we observed a specific antibody response against the trypsinised Cry1Ab protein we might also anticipate an effect on cell influx in lungs and cytokine release from MLN cells from mice exposed to the trypCry1Ab protein compared to controls. However, these parameters were not significantly altered by trypCry1Ab. Interestingly, a significant influx of lymphocytes and eosinophils in the broncho-alveolar lavage fluid were evident in the MON810 leaf extract exposed mice compared to controls, and also the cytokine release of IL10, IFN γ , IL4, IL-13 and IL-17 were higher in MLN cells from the MON810 leaf exposed mice. When MLN cells were stimulated *in vitro* with the trypCry1Ab protein, cytokine release was induced only in MON810 treated mice, indicating a specific response to the Cry1Ab epitopes, i.e. suggesting immunogenicity of Cry1Ab protein of plant origin. The increase of cytokines both related to Tregs (IL-10), Th2 (IL-4 and IL-13), Th1 (IFN γ) and Th17 (IL-17) cells, suggest a general immune stimulation rather than skewing of the immune response in a particular direction. In agreement, the antibody response towards the Cry1Ab induced by protoxin and trypCry1A were both associated with Th1 and Th2 associated antibody classes (IgG2a, and IgE and IgG1, respectively). However, neither the cell influx nor the cytokine levels (with IL-10 as the only exception) were significantly different from the non-GM exposed mice, suggesting that the local immune response was elicited in response to the plant material as such, and not related to the Cry1Ab protein.

It has previously been demonstrated that immunisation dose, as well as sex and age, highly influence the allergy outcome (Hansen et al., 2011). The immunogenic and allergenic effects seen in prime aged females (6-11 weeks old) included in the present experiment and in males (10-x weeks old) in (Guerrero et al., 2004) indicate that Cry1Ab proteins could affect the mature immune system. Little is known, however, concerning the impact of these proteins on vulnerable groups. Finamore et al. (2008) reported that the immune systems of young and old

rats were more affected by Cry1Ab containing feed, than prime aged individuals. It would therefore be reasonable to assume that the present findings would be of even greater importance in vulnerable age groups or in hypersensitive individuals.

Although no Cry protein immunogenicity could be observed at the present dose of MON810 pollen or leaf extract, the specific antibody response against the purified Cry1Ab protoxin and toxin preparations demonstrates the principle that these proteins may affect the immune system, i.e. act as immunogens, after intranasal administration. The elicitation of specific IgE antibodies indicates allergenicity of the purified Cry1Ab proteins. However, this was not observed after exposure to (lower doses of) Cry1Ab in plant materials. Notably, the observed immune responses were elicited in the absence of an adjuvant. Given the importance of Bt-transgenic maize as food and feed across the world, a considerable number of individuals may be exposed to Cry1Ab by inhalation, in the field as well as along the food/feed chain. Thus, our results warrant further clarification and testing, both in animal models and in humans, including emphasis on vulnerable age groups and hypersensitive individuals.

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All authors declare no conflict of interest

5. REFERENSES

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Table 1. Total exposure doses of Cry1Ab in MON810 pollen and leaves per mouse.

Source	Cry1Ab ($\mu\text{g}/\text{mg}$)	Plant material (mg) per mouse	Total dose Cry1Ab (μg) per mouse
<i>MON810 pollen</i>	0.00692	3 ^a	0.02
<i>MON810 leaf</i>	0.61251	21 ^b	12.86

^a Exposure 6 days of 500 μg pollen each day.

^b Exposure 6 days of 3.5 mg leaves (in extract form) each day

Figure 1

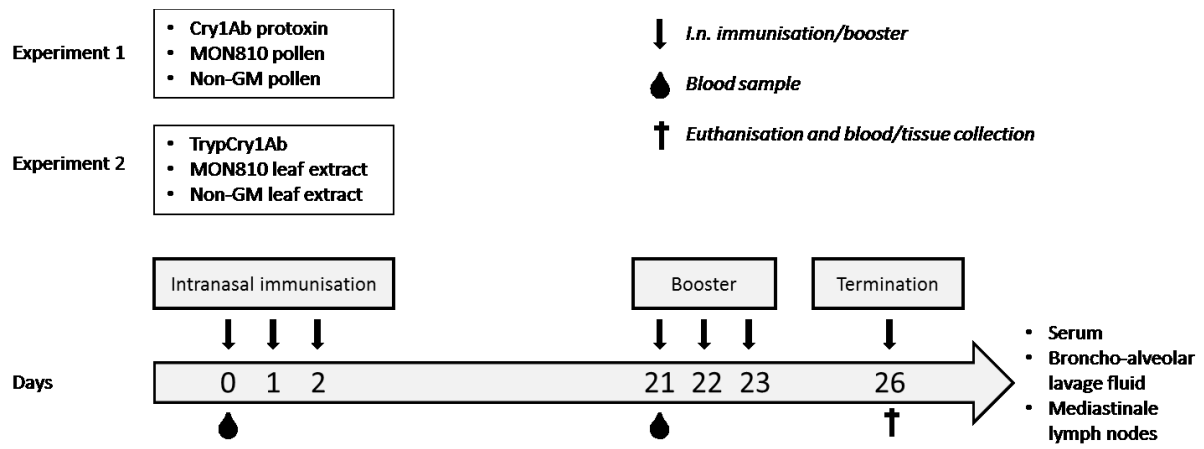


Figure 2

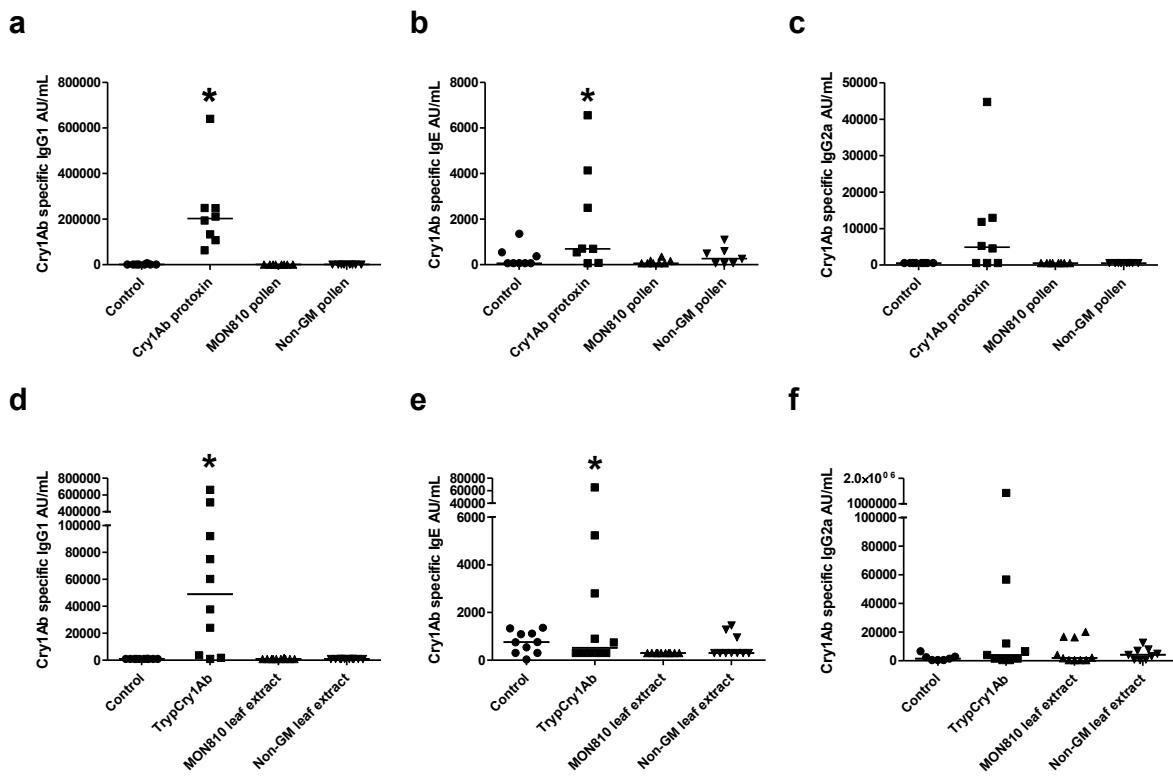


Figure 3

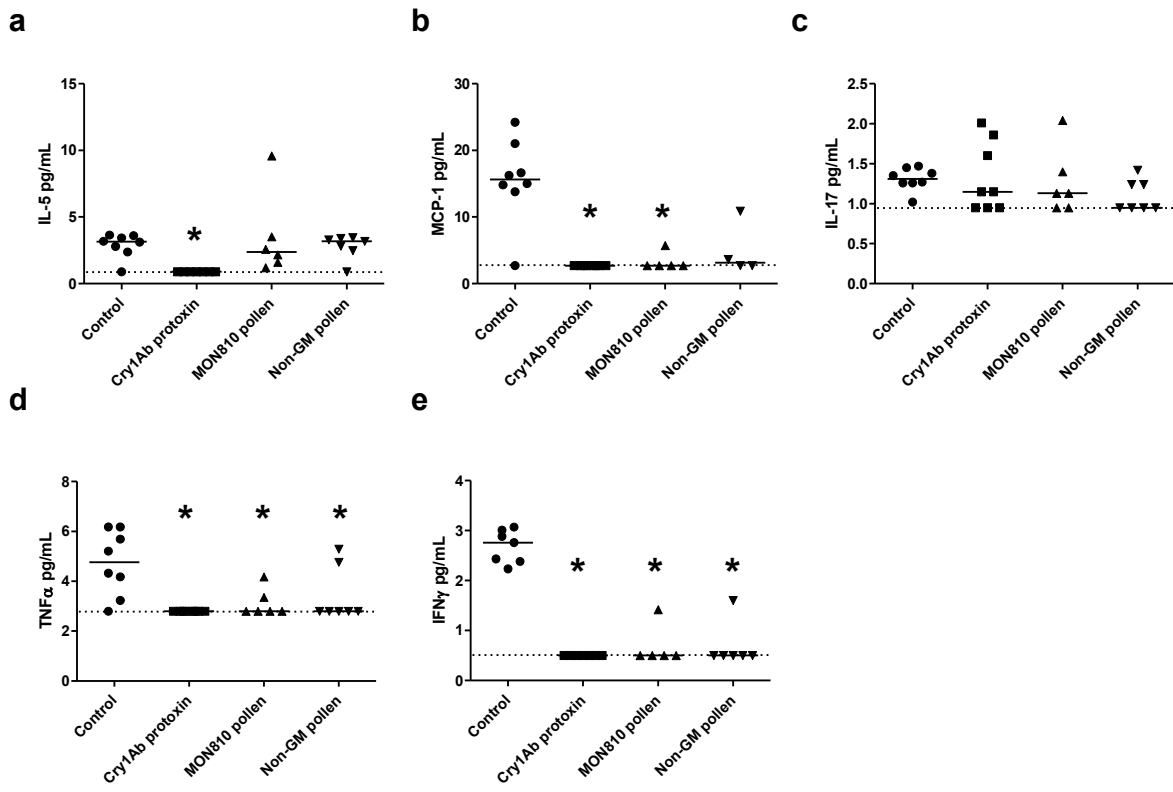


Figure 4

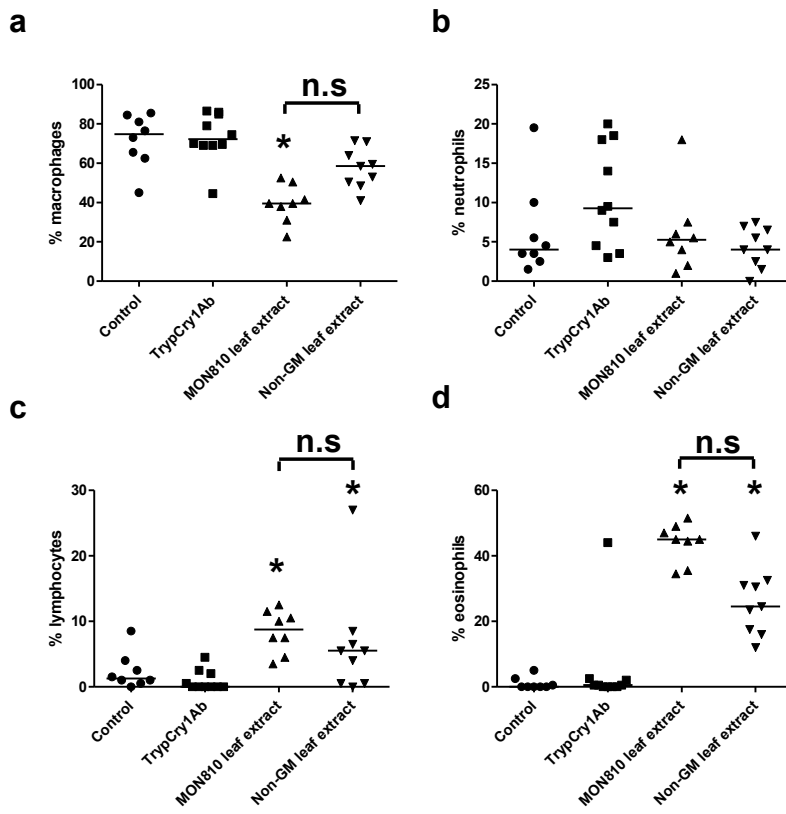


Figure 5

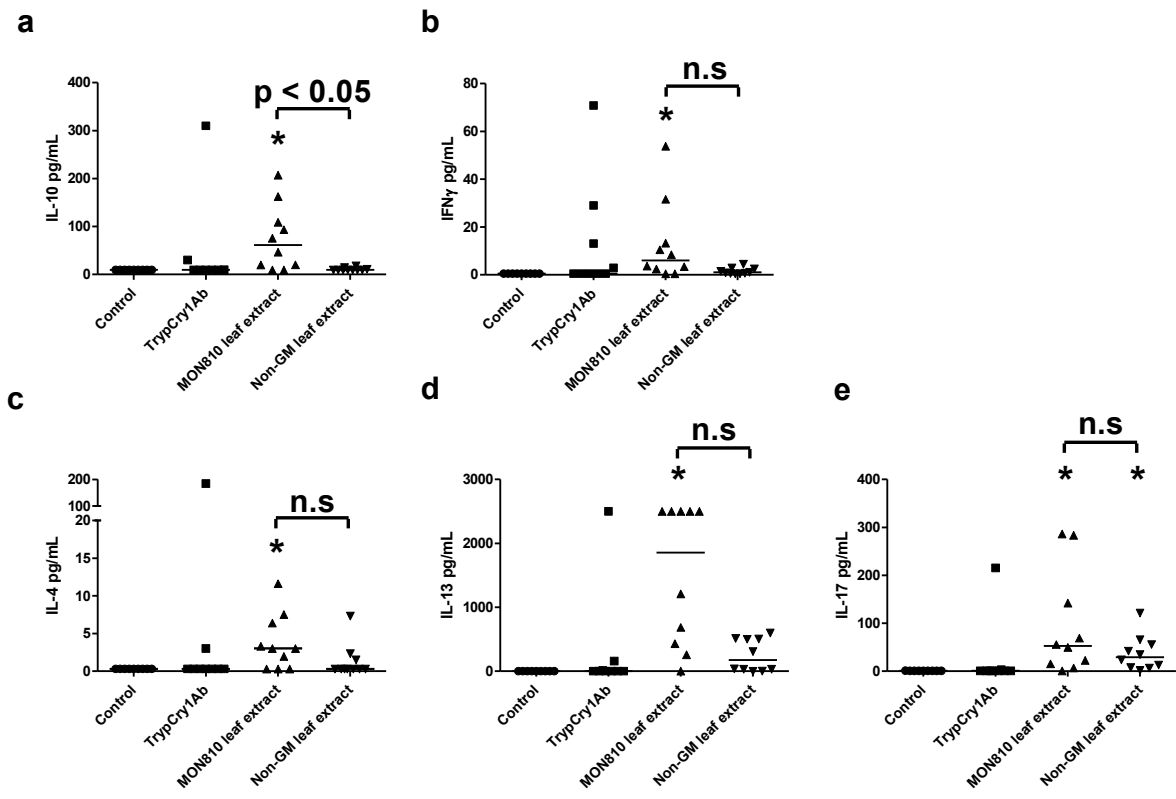


Figure 1. Experimental protocol. Mice were intranasally (i.n.) immunised with Cry1Ab protoxin, MON810 pollen and non-GM pollen (experiment 1), and trypsinised Cry1Ab (trypCry1Ab), MON810 leaf extract and non-GM leaf extract (experiment 2) on days 0, 1, 2, and boosted i.n. with the equivalent test materials on days 21, 22, 23. Blood samples were collected on day 0 and 21, and blood, broncho-alveolar lavage fluids (BALF) and mediastinal lymph nodes (MLN) were collected at termination on day 26.

Figure 2. Cry1Ab specific IgG1 (a), IgE (b) and IgG2a (c) in serum after intranasal immunisation and booster of mice to Cry1Ab protoxin and fresh MON810 maize pollen and non-GM pollen on days 0, 1, 2, 3, 21, 22 and 23. Cry1Ab specific IgG1 (d), IgE (e) and IgG2a (d) in serum after intranasal immunisation and booster of mice to trypsinised Cry1Ab and MON810 leaf extract and non-GM leaf extracts on days 0, 1, 2, 3, 21, 22 and 23. Asterisks (*) denote groups that are significantly different ($p < 0.05$) from control group exposed to physiological buffer only.

Figure 3. Cytokine IL-5 (a), MCP-1 (b), IL-17 (c), TNF α (d) and IFN γ (e) levels in broncho-alveolar lavage fluid (BALF) collected on day 26 after intranasal exposure of mice to Cry1Ab protoxin and fresh maize pollen on days 0, 1, 2, 21, 22 and 23. Each circle represents one individual animal, and bars represent group medians. The dotted lines indicate the lower quantitative detection limit of the assays. Asterisks (*) denote groups that are significantly different ($p < 0.05$) from the control group (CTRL) exposed to physiological buffer only.

Figure 4. Percentages (%) of macrophages (a), neutrophils (b), lymphocytes (c) and eosinophils (d) in the broncho-alveolar lavage fluid (BALF) collected on day 26 after intranasal exposure of mice to trypsinised Cry1Ab (trypCry1Ab), or leaf extracts from the genetically modified maize event MON810 and its conventional counterpart (non-GM) on days 0, 1, 2, 21, 22 and 23. Bars represent

group medians, circles represent one individual animal. Asterisks (*) denote groups that are significantly different ($p < 0.05$) from the control group exposed to physiological buffer only.

Figure 5. Cytokine IL-10 (a), IFN γ (b), IL-4 (c), IL-13 (d) and IL-17 (e) secretion (pg/mL) from mediastinal lymph node (MLN) cells collected on day 26 after intranasal exposure of mice to trypsinised Cry1Ab (trypCry1Ab), or leaf extracts from the genetically modified maize event MON810 or its conventional counterpart (non-GM) on days 0, 1, 2, 21, 22 and 23. Each circle represents one individual animal, and bars represent group medians. The dotted lines indicate the lower quantitative detection limit of the assays. Asterisks (*) denote groups that are significantly different ($p < 0.05$) from the control group exposed to physiological buffer only